



Basic and Application

Inductively coupled plasma
mass spectrometry (ICPMS)



Inductively coupled plasma mass spectrometry (ICP-MS) is a **type of mass spectrometry** which is capable of detecting metals and several non-metals at concentrations as low as one part in 10^{12} (ppt). This is achieved by **ionizing** the sample **with inductively coupled plasma** and then using a mass spectrometer to separate and quantify those ions.

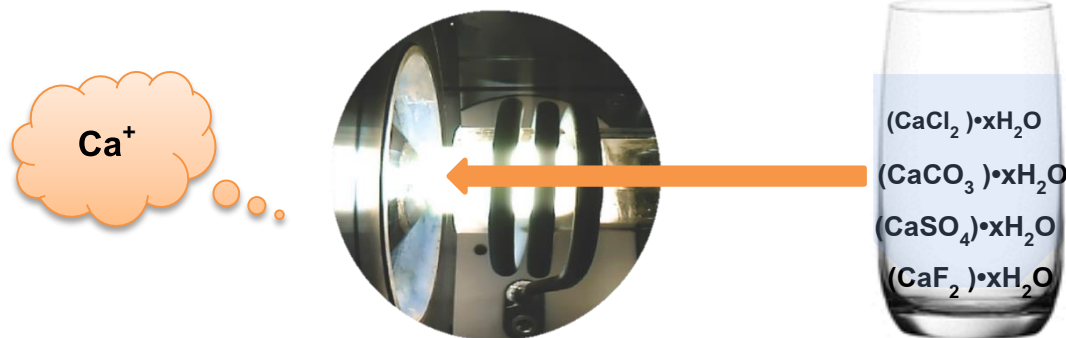
Mission :

Convert all analyses species in sample to each isotope ions which be able to measure by mass spectrometer.

User-friendly operation

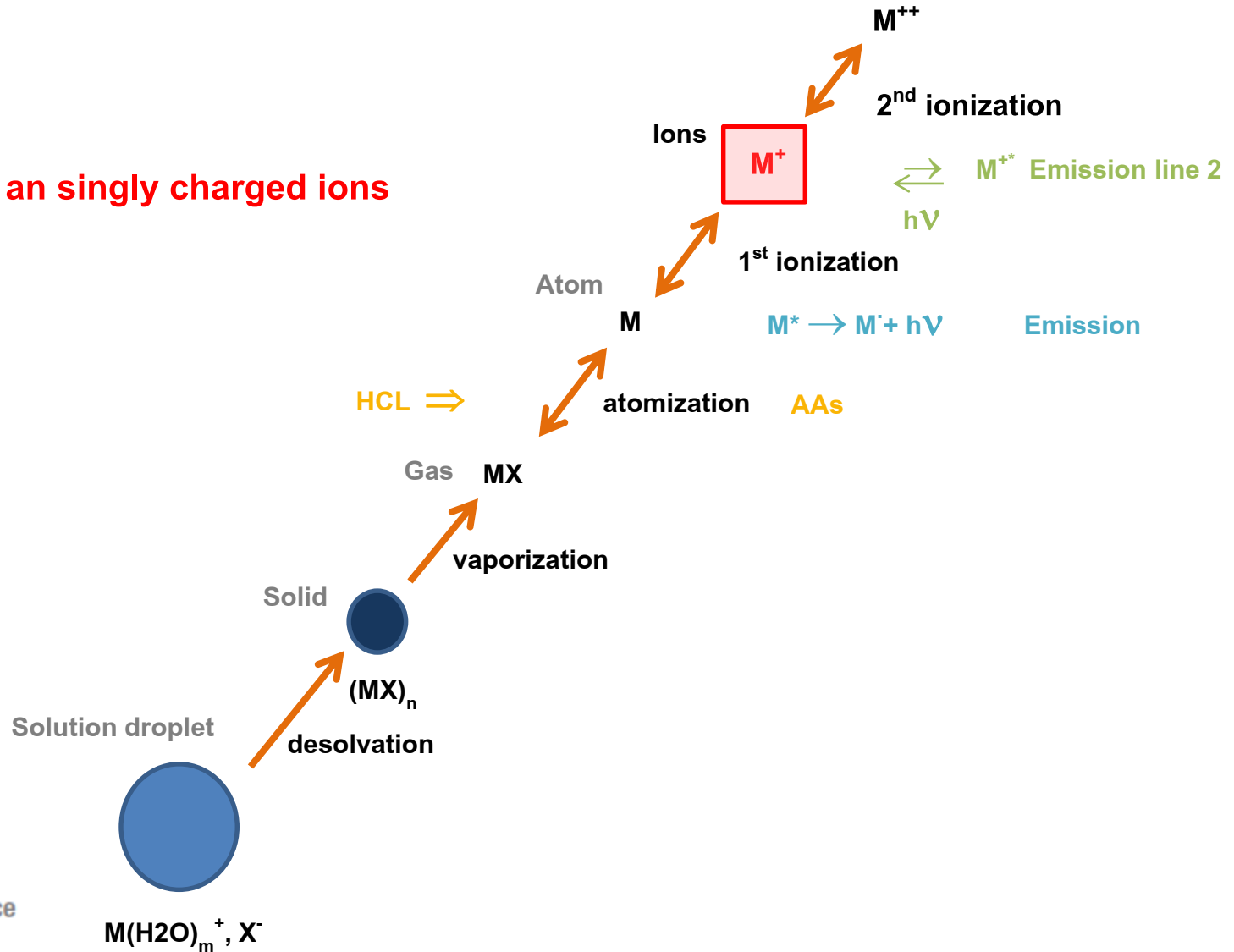
Examples : if we are looking for Calcium in water sample.

More analysis in less time



High uptime, low maintenance

MS need an singly charged ions

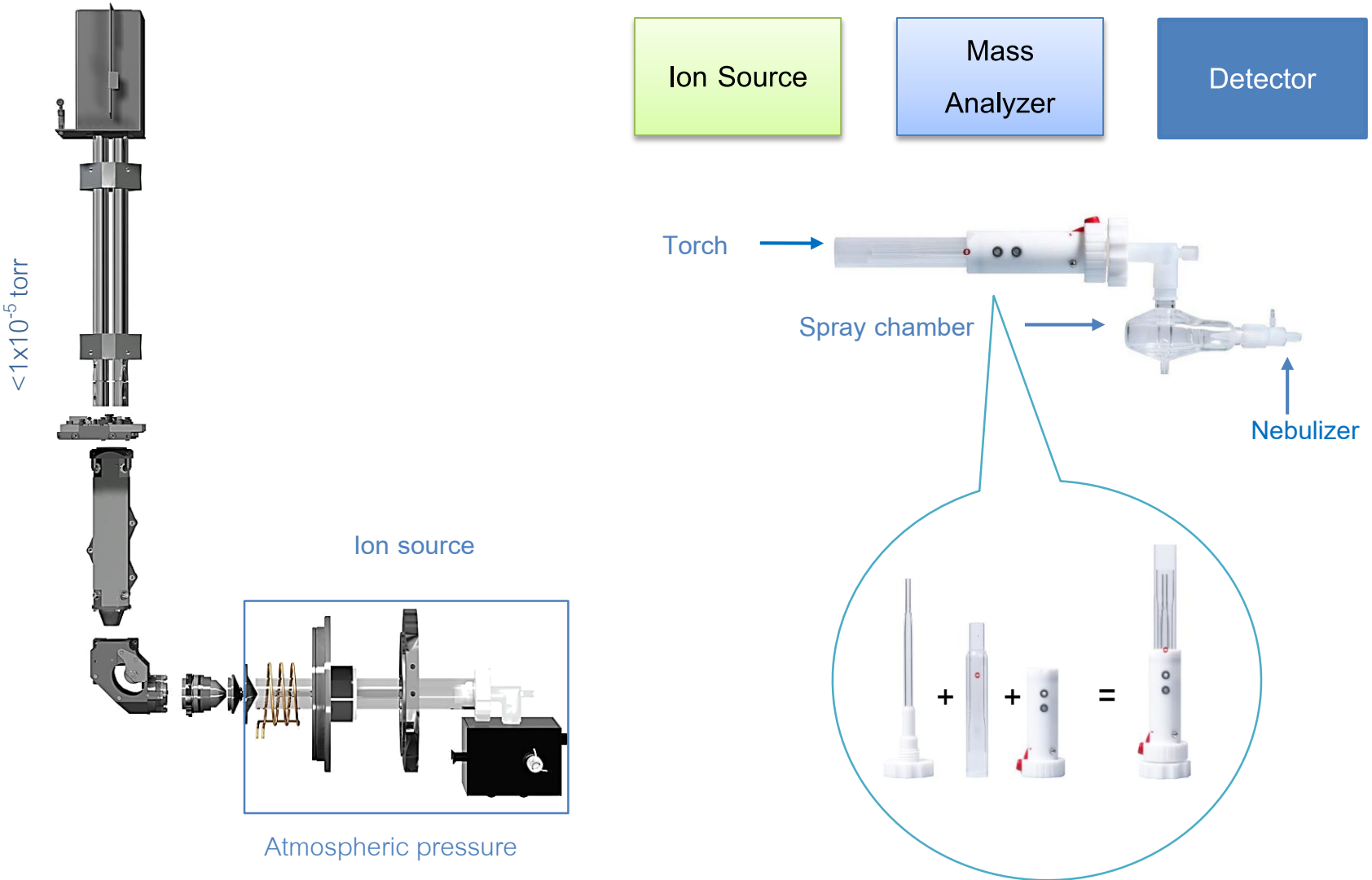


User-friendly operation

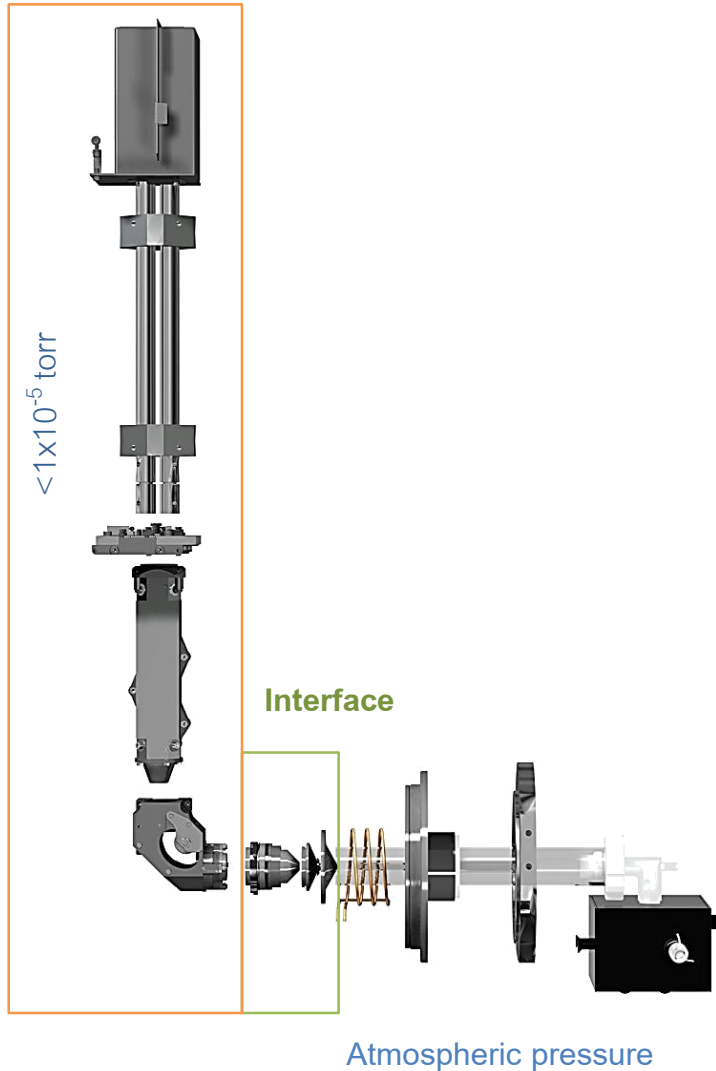
More analysis in less time

High uptime, low maintenance

ICP-MS Instrumental



ICP-MS Instrumental



Ion Source

Mass Analyzer

Detector

How to extract the ions into MS ?
 The positively charged ions that are produced in the plasma are extracted into the vacuum system, via a pair of interface “cones” and the “extraction lens”



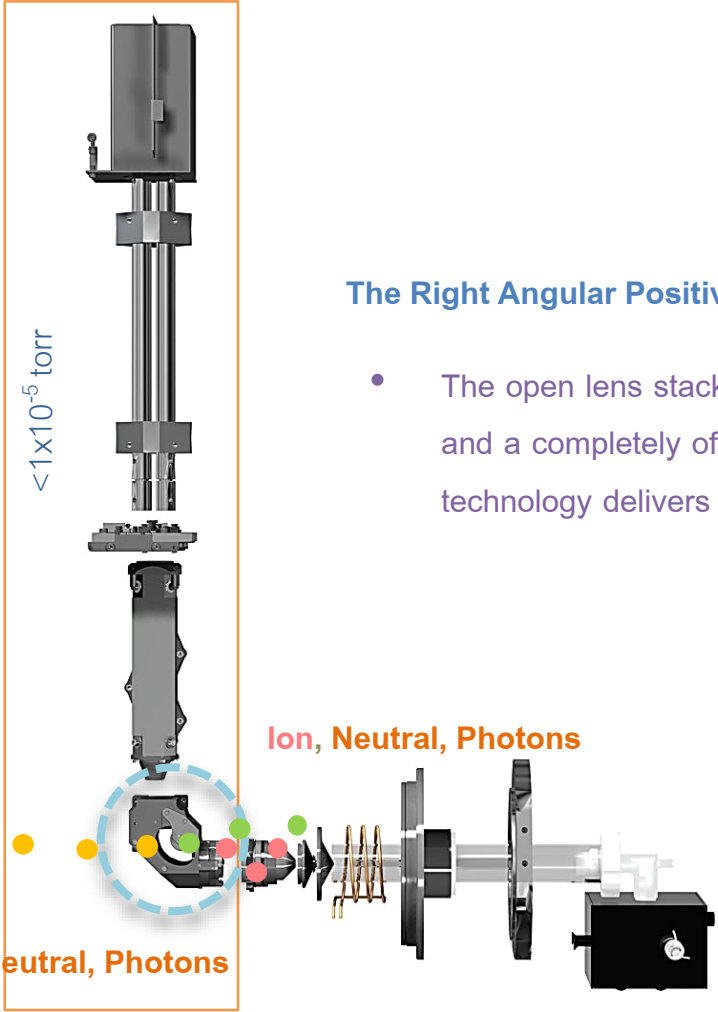
Ion Source

Mass Analyzer

Detector

The Right Angular Positive Ion Deflection (RAPID) lens

- The open lens stack eliminates lens cleaning maintenance and a completely off-axis design together with QCell technology delivers a class leading background noise.

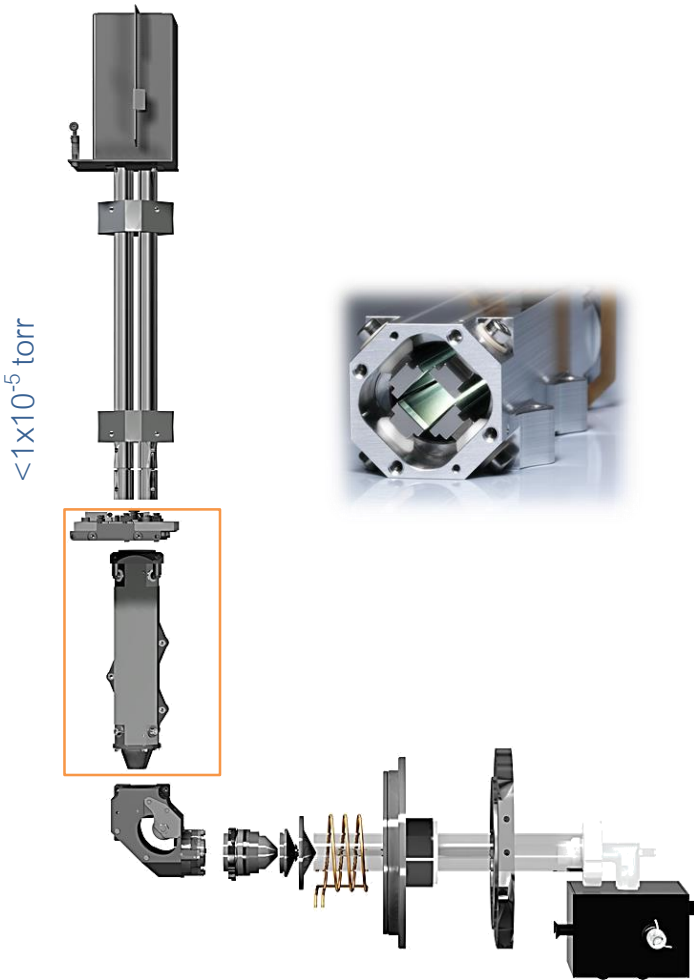


Elimination of neutral species

$^{40}\text{Ar}^{16}\text{O}$
 $^{40}\text{Ca}^{16}\text{O}$

$^{40}\text{Ar}^{35}\text{Cl}$

ANALYTE	POTENTIAL INTERFERENT	PRECURSORS
^{45}Sc	$^{13}\text{C}^{16}\text{O}_2$, $^{12}\text{C}^{16}\text{O}_2\text{H}$, ^{44}CaH , $^{32}\text{S}^{12}\text{CH}$, $^{32}\text{S}^{13}\text{C}$, $^{33}\text{S}^{12}\text{C}$	H, C, O, S, Ca
^{47}Ti	$^{31}\text{P}^{16}\text{O}$, ^{46}CaH , $^{35}\text{Cl}^{12}\text{C}$, $^{32}\text{S}^{14}\text{NH}$, $^{33}\text{S}^{14}\text{N}$	H, C, N, O, P, S, Cl, Ca
^{49}Ti	$^{31}\text{P}^{18}\text{O}$, ^{48}CaH , $^{35}\text{Cl}^{14}\text{N}$, $^{37}\text{Cl}^{12}\text{C}$, $^{32}\text{S}^{16}\text{OH}$, $^{33}\text{S}^{16}\text{O}$	H, C, N, O, P, S, Cl, Ca
^{50}Ti	$^{34}\text{S}^{16}\text{O}$, $^{32}\text{S}^{18}\text{O}$, $^{35}\text{Cl}^{14}\text{NH}$, $^{37}\text{Cl}^{12}\text{CH}$	H, C, N, O, S, Cl
^{51}V	$^{35}\text{Cl}^{16}\text{O}$, $^{37}\text{Cl}^{14}\text{N}$, $^{34}\text{S}^{16}\text{OH}$	H, O, N, S, Cl
^{52}Cr	$^{36}\text{Ar}^{16}\text{O}$, $^{40}\text{Ar}^{12}\text{C}$, $^{35}\text{Cl}^{16}\text{OH}$, $^{37}\text{Cl}^{14}\text{NH}$, $^{34}\text{S}^{18}\text{O}$	H, C, O, N, S, Cl, Ar
^{55}Mn	$^{37}\text{Cl}^{18}\text{O}$, $^{23}\text{Na}^{32}\text{S}$, $^{23}\text{Na}^{31}\text{PH}$	H, O, Na, P, S, Cl, Ar
^{56}Fe	$^{40}\text{Ar}^{16}\text{O}$, $^{40}\text{Ca}^{16}\text{O}$	O, Ar, Ca
^{57}Fe	$^{40}\text{Ar}^{16}\text{OH}$, $^{40}\text{Ca}^{16}\text{OH}$	H, O, Ar, Ca
^{58}Ni	$^{40}\text{Ar}^{18}\text{O}$, $^{40}\text{Ca}^{18}\text{O}$, $^{23}\text{Na}^{35}\text{Cl}$	O, Na, Cl, Ar, Ca
^{59}Co	$^{40}\text{Ar}^{18}\text{OH}$, $^{43}\text{Ca}^{16}\text{O}$, $^{23}\text{Na}^{35}\text{ClH}$	H, O, Na, Cl, Ar, Ca
^{60}Ni	$^{44}\text{Ca}^{16}\text{O}$, $^{23}\text{Na}^{37}\text{Cl}$	O, Na, Cl, Ca
^{61}Ni	$^{44}\text{Ca}^{16}\text{OH}$, $^{38}\text{Ar}^{23}\text{Na}$, $^{23}\text{Na}^{37}\text{ClH}$	H, O, Na, Cl, Ca
^{63}Cu	$^{40}\text{Ar}^{23}\text{Na}$, $^{12}\text{C}^{16}\text{O}^{35}\text{Cl}$, $^{12}\text{C}^{14}\text{N}^{37}\text{Cl}$, $^{31}\text{P}^{32}\text{S}$, $^{31}\text{P}^{16}\text{O}_2$	C, N, O, Na, P, S, Cl
^{64}Zn	$^{32}\text{S}^{16}\text{O}_2$, $^{32}\text{S}_2$, $^{36}\text{Ar}^{12}\text{C}^{16}\text{O}$, $^{38}\text{Ar}^{12}\text{C}^{14}\text{N}$, $^{48}\text{Ca}^{16}\text{O}$	C, N, O, S, Ar, Ca
^{65}Cu	$^{32}\text{S}^{16}\text{O}_2\text{H}$, $^{32}\text{S}_2\text{H}$, $^{14}\text{N}^{16}\text{O}^{35}\text{Cl}$, $^{48}\text{Ca}^{16}\text{OH}$	H, N, O, S, Cl, Ca
^{66}Zn	$^{34}\text{S}^{16}\text{O}$, $^{32}\text{S}^{34}\text{S}$, ^{33}S , ^{48}C , ^{18}O	O, C, S
^{69}Ga	$^{32}\text{S}^{18}\text{O}_2\text{H}$, $^{34}\text{S}_2\text{H}$, $^{37}\text{Cl}^{16}\text{O}_2$	H, O, S, Cl
^{70}Zn	$^{34}\text{S}^{18}\text{O}_2$, $^{35}\text{Cl}_2$	O, S, Cl
^{75}As	$^{40}\text{Ar}^{34}\text{SH}$, $^{40}\text{Ar}^{35}\text{Cl}$, $^{40}\text{Ca}^{35}\text{Cl}$, $^{37}\text{Cl}_2\text{H}$	H, S, Cl, Ca, Ar
^{77}Se	$^{40}\text{Ar}^{37}\text{Cl}$, $^{40}\text{Ca}^{37}\text{Cl}$	Cl, Ca, Ar
^{78}Se	$^{40}\text{Ar}^{38}\text{Ar}$	Ar
^{80}Se	$^{40}\text{Ar}_2$, $^{40}\text{Ca}_2$, $^{40}\text{Ar}^{40}\text{Ca}$, $^{32}\text{S}_2^{16}\text{O}$, $^{32}\text{S}^{16}\text{O}_3$	O, S, Ar, Ca



Ion Source

Mass Analyzer

Detector

Qcell Collision/Reaction Cell (CRC)

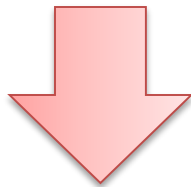
- Flatapole technology for improved transmission
- Low mass cut off filters out unwanted precursor ions
- Single mode interference removal with He
- He KED filters out unwanted polyatomic interferences
- Small CRC volume for fast gas exchange
- Flexibility to work with reactive gases, such as mixtures of O_2 7% dilute H_2 or 1% NH_3
- Non-consumable, zero-maintenance

Atmospheric pressure

He KED filters out unwanted polyatomic interferences

High transmission enables analysis of even low mass analytes in He KED mode

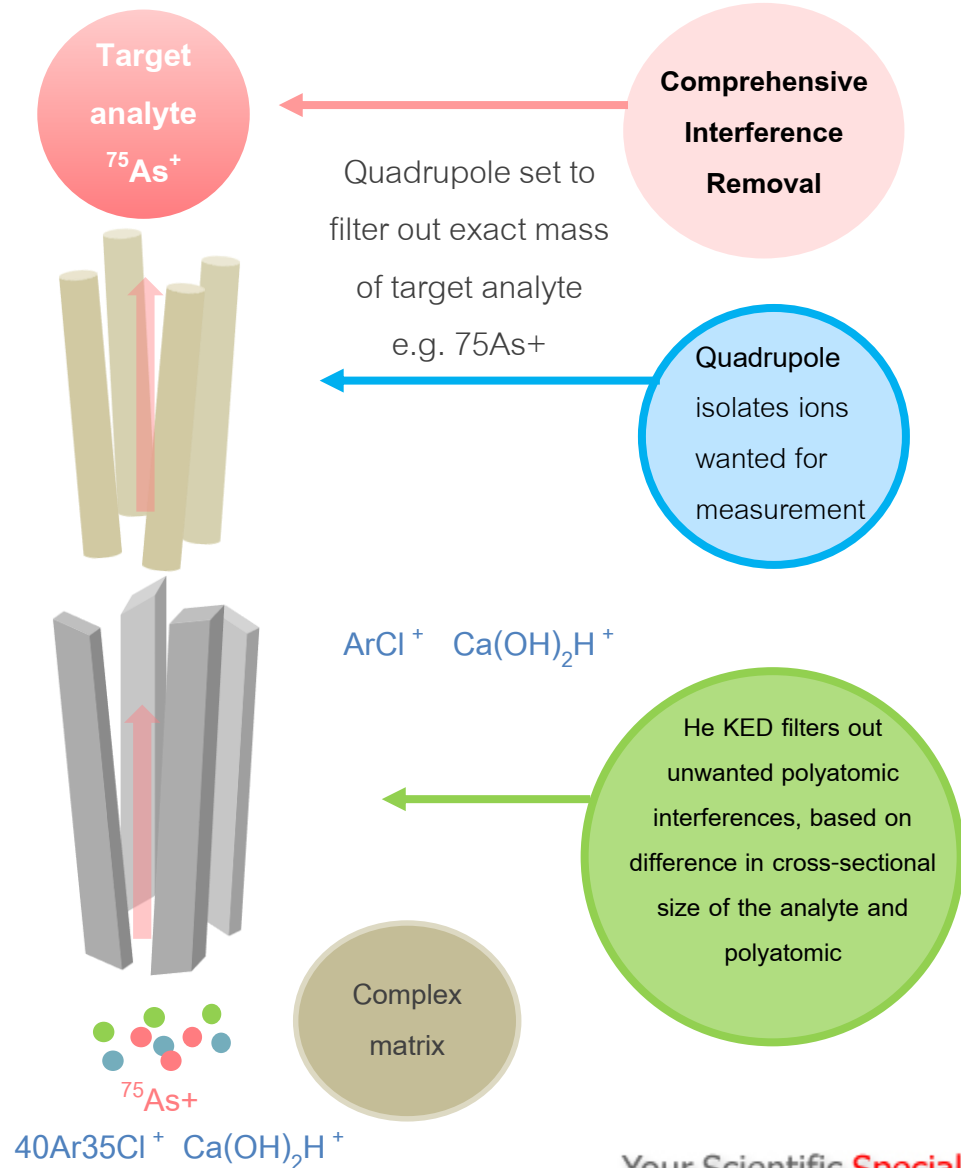
Single measurement mode for all analytes in analytical method



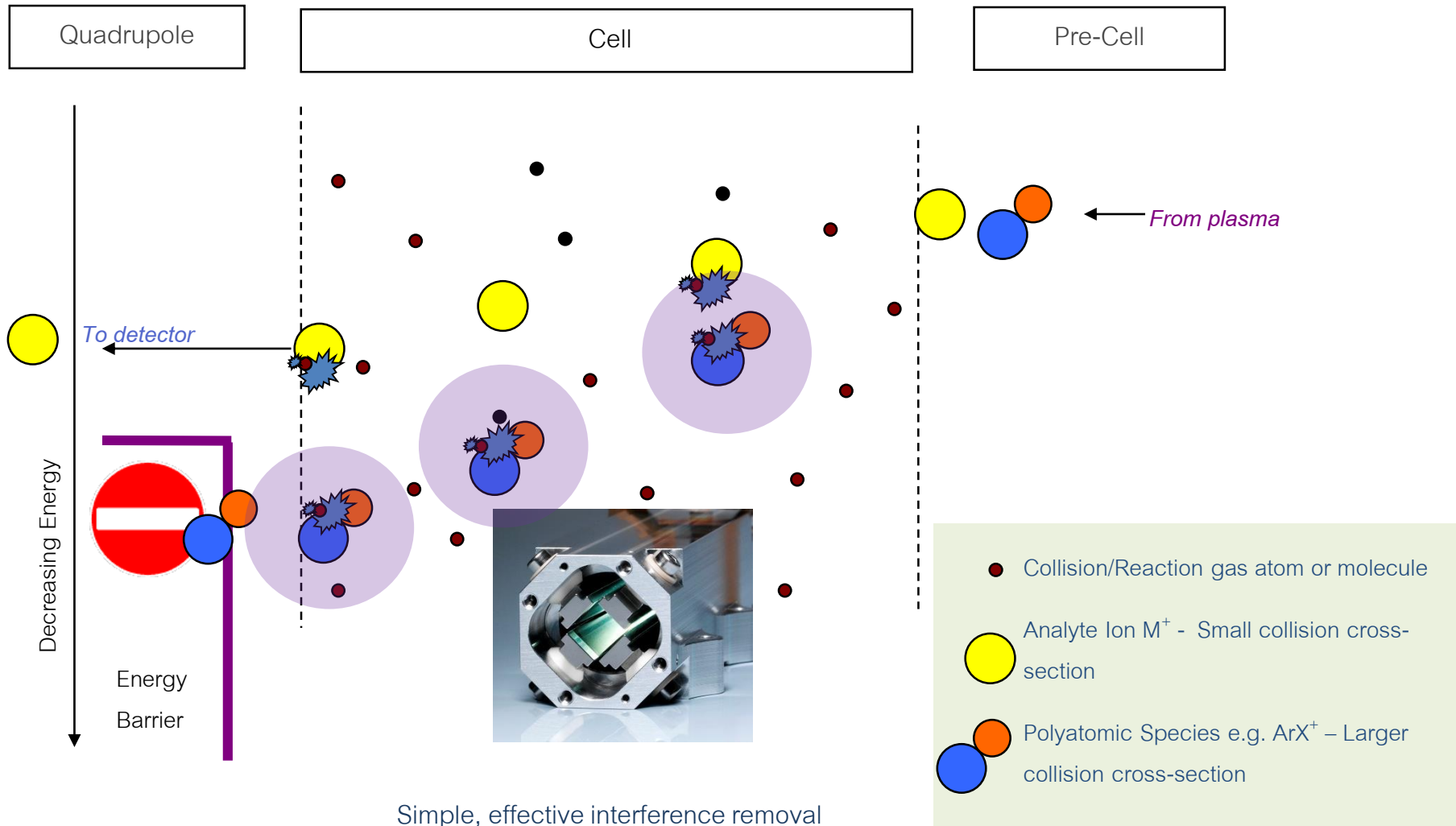
No gas switching makes method more productive and simple

No change of measurement mode effectively eliminates method development

Unique Flatapole Design



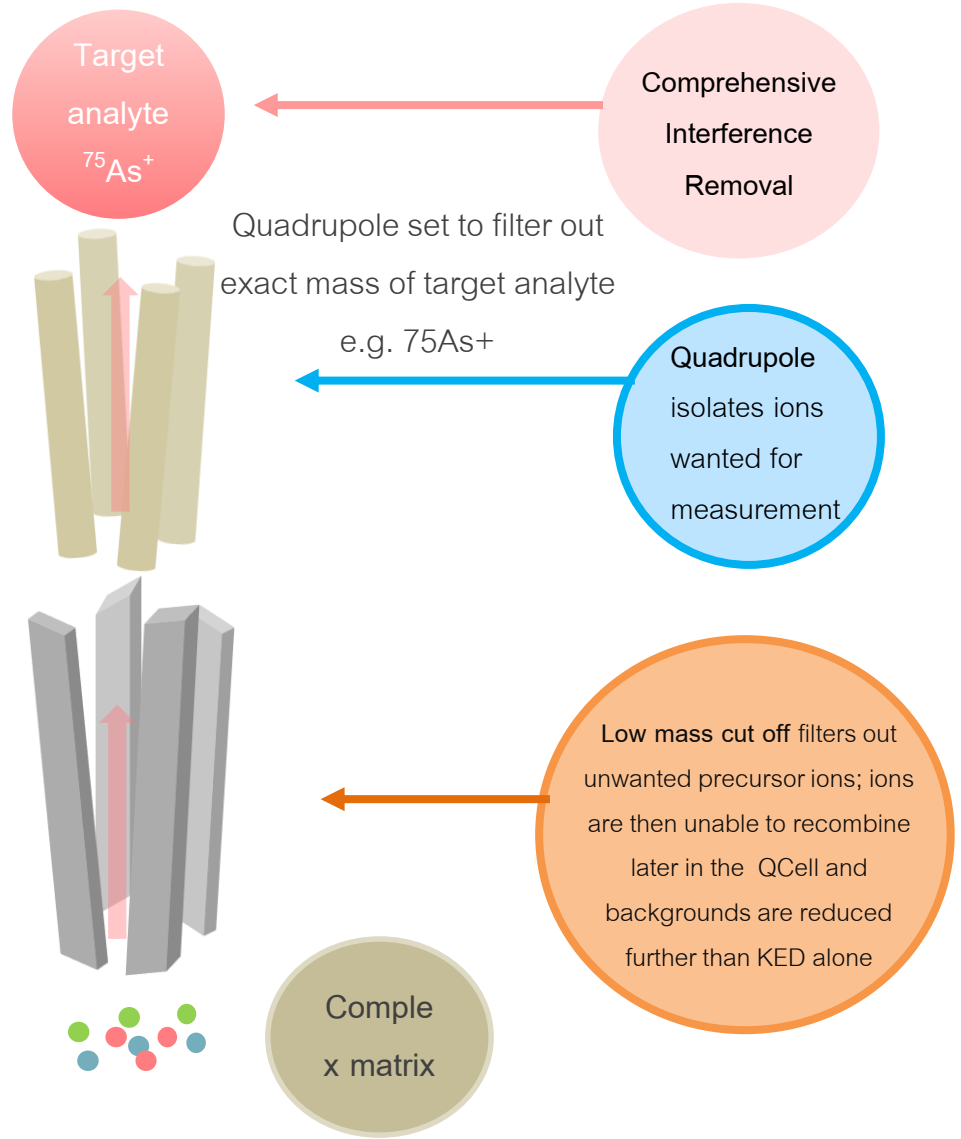
Collisional retardation / energy filtering



QCell flatapole dynamically applies Low Mass Cut Off (LCMO) relative to target analyte

Anal	LMCO	Interferences	Precursors
^{51}V	35	$^{35}\text{Cl}^{16}\text{O}$, $^{37}\text{Cl}^{14}\text{N}$, $^{34}\text{S}^{16}\text{OH}$	H, N, O, S, Cl
^{56}Fe	39	$^{40}\text{Ar}^{16}\text{O}$, $^{40}\text{Ca}^{16}\text{O}$	O, Ar, Ca
^{63}Cu	45	$^{40}\text{Ar}^{23}\text{Na}$, $^{12}\text{C}^{16}\text{O}^{35}\text{Cl}$, $^{31}\text{P}^{32}\text{S}$	C, N, O, Na, P, S, Cl, Ar
^{75}As	47	$^{40}\text{Ar}^{35}\text{Cl}$, $^{40}\text{Ca}^{35}\text{Cl}$, $^{40}\text{Ar}^{34}\text{SH}$, $^{37}\text{Cl}_2\text{H}$	H, S, Cl, Ca, Ar

Unique Flatapole Design

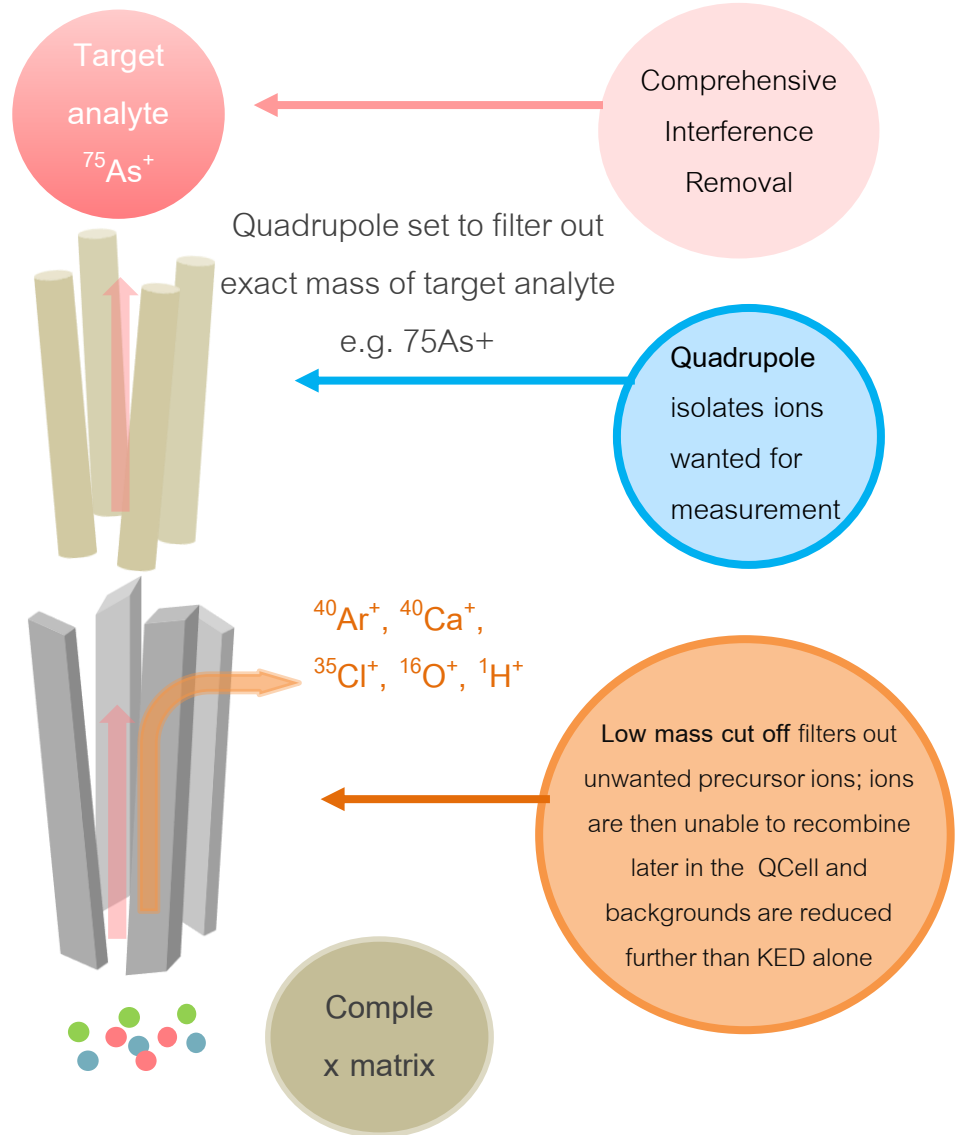


QCell flatapole dynamically applies Low Mass Cut Off (LCMO) relative to target analyte

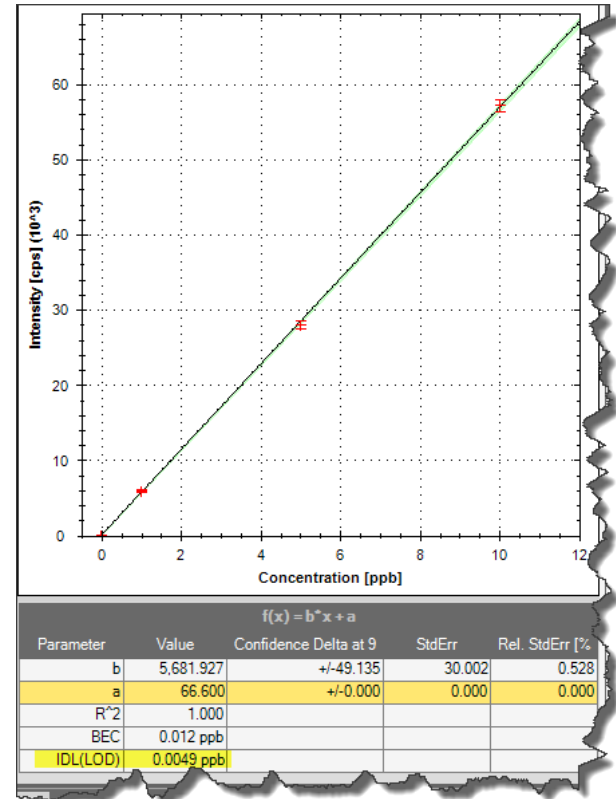
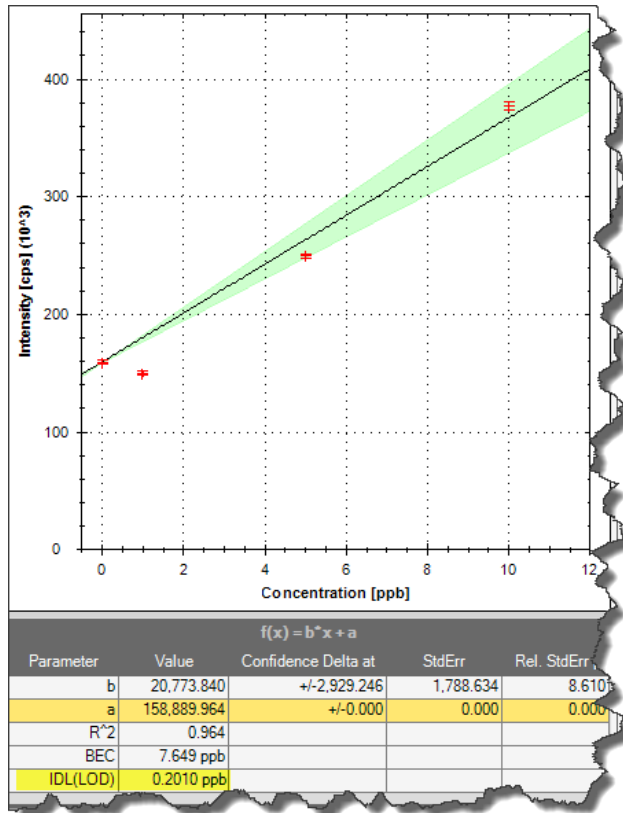
Anal	LMCO	Interferences	Precursors
^{51}V	35	$^{35}\text{Cl}^{16}\text{O}$, $^{37}\text{Cl}^{14}\text{N}$, $^{34}\text{S}^{16}\text{OH}$	H, N, O, S, Cl
^{56}Fe	39	$^{40}\text{Ar}^{16}\text{O}$, $^{40}\text{Ca}^{16}\text{O}$	O, Ar, Ca
^{63}Cu	45	$^{40}\text{Ar}^{23}\text{Na}$, $^{12}\text{C}^{16}\text{O}^{35}\text{Cl}$, $^{31}\text{P}^{32}\text{S}$	C, N, O, Na, P, S, Cl, Ar
^{75}As	47	$^{40}\text{Ar}^{35}\text{Cl}$, $^{40}\text{Ca}^{35}\text{Cl}$, $^{40}\text{Ar}^{34}\text{SH}$, $^{37}\text{Cl}_2\text{H}$	H, S, Cl, Ca, Ar

All unwanted precursors that contribute to interferences are eliminated

Unique Flatapole Design



Calibration curve for ⁷⁵As in a solution containing 0.5% HCl

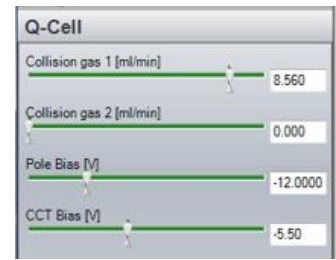
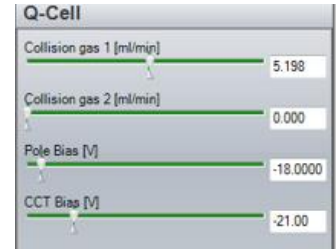
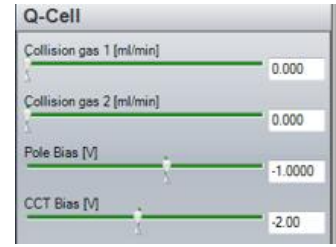
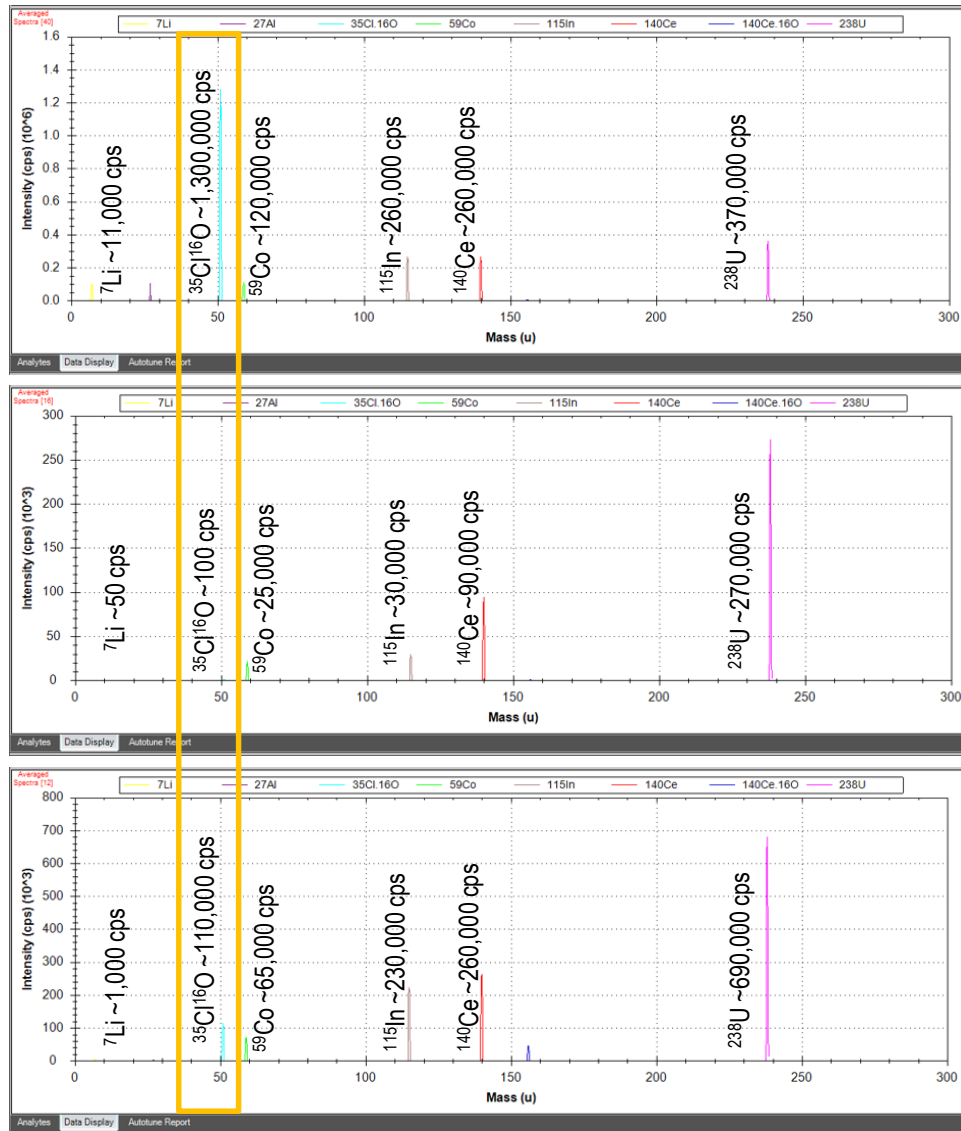


STD mode: Polyatomic interference leads to poor IDL and elevated BEC

KED mode: Polyatomic interference removed
IDL below 5 ppt

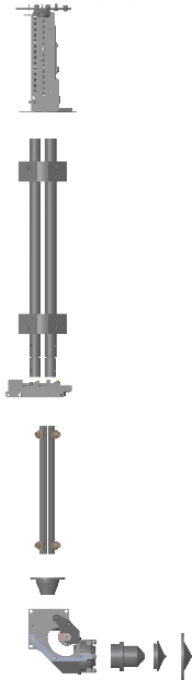


Element	ppb
Li	1
Co	1
In	1
Ba	1
Ce	1
Bi	1
U	1

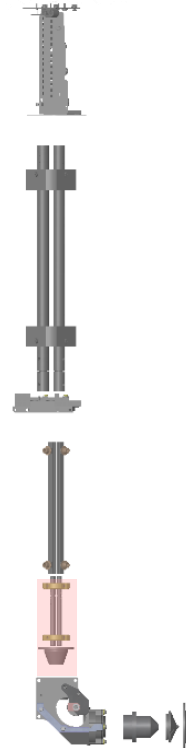


What's the difference between a SQ and TQ-ICP-MS

iCAP RQ ICP-MS



iCAP TQ ICP-MS



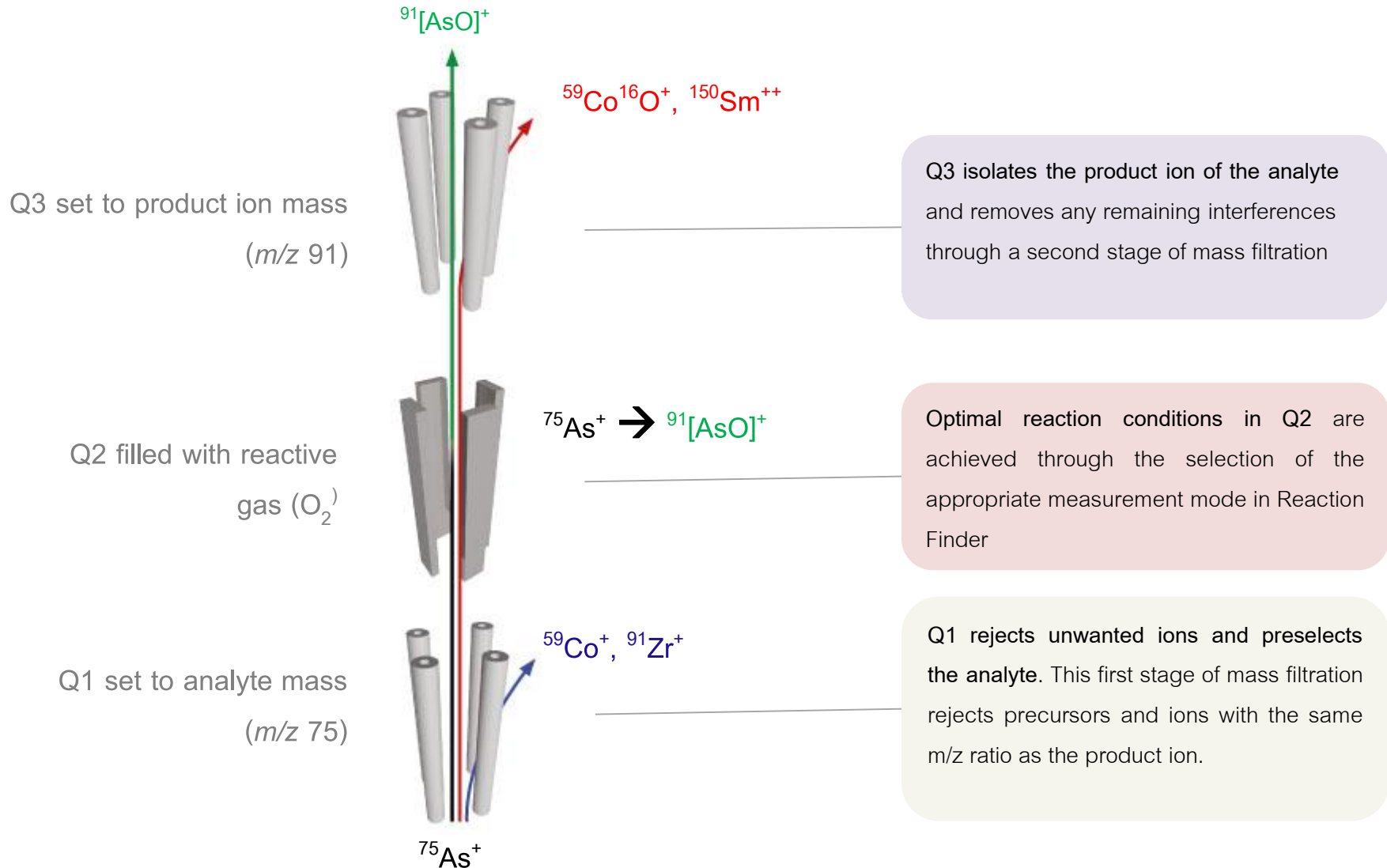
Additional Q1 mass
filter quadrupole

Additional electronics

Additional gases

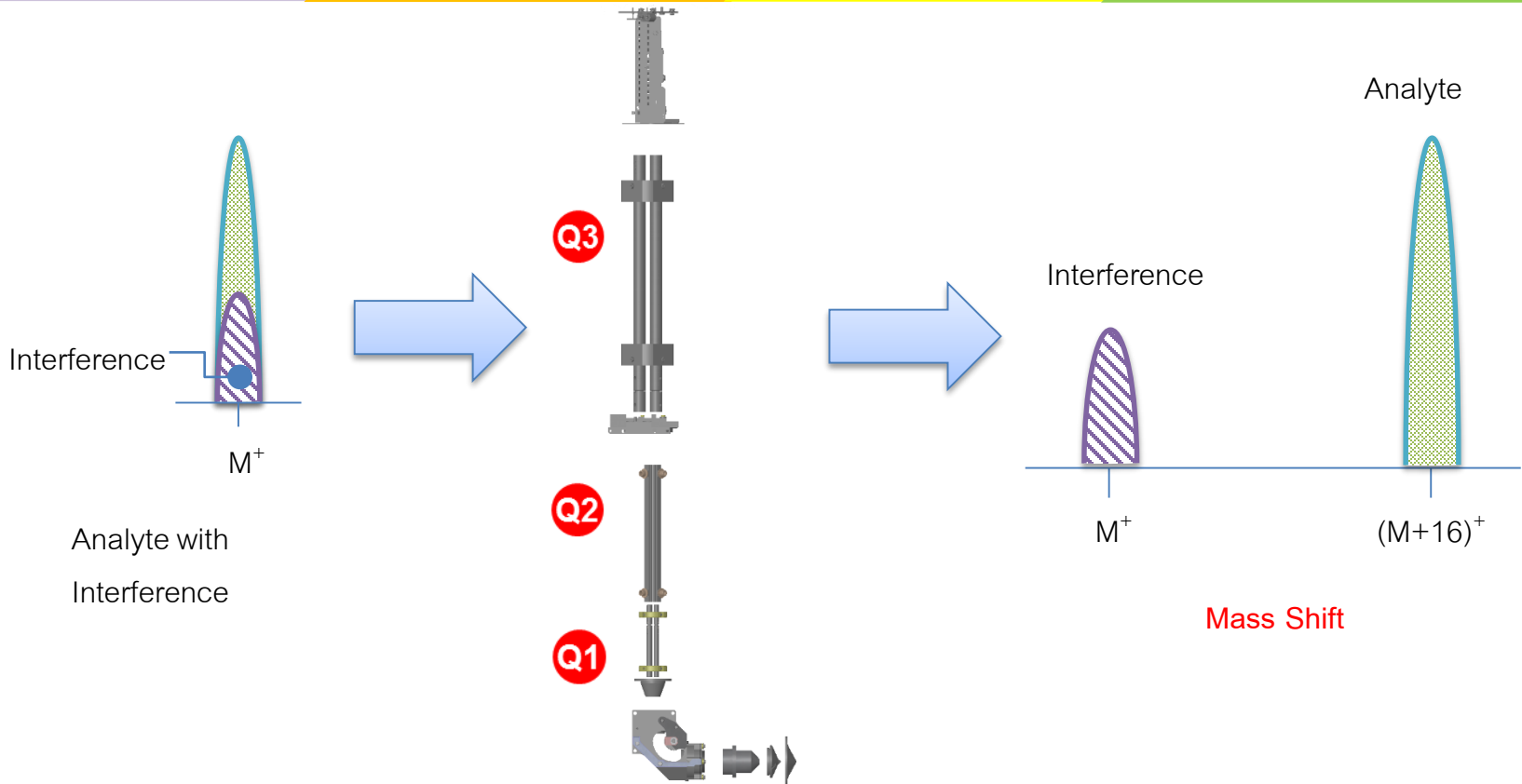
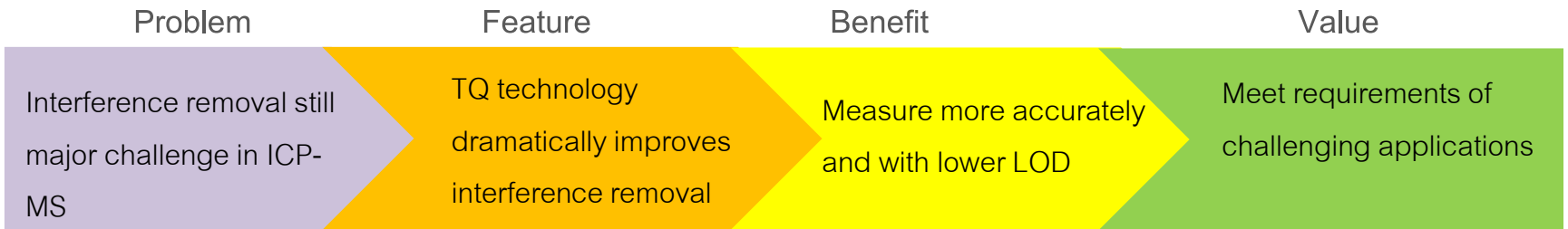
Enhanced software

Same platform – there are more differences than you think!

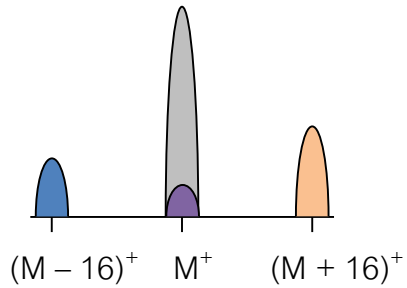


Mass shift interference removal with the iCAP TQ ICP-MS



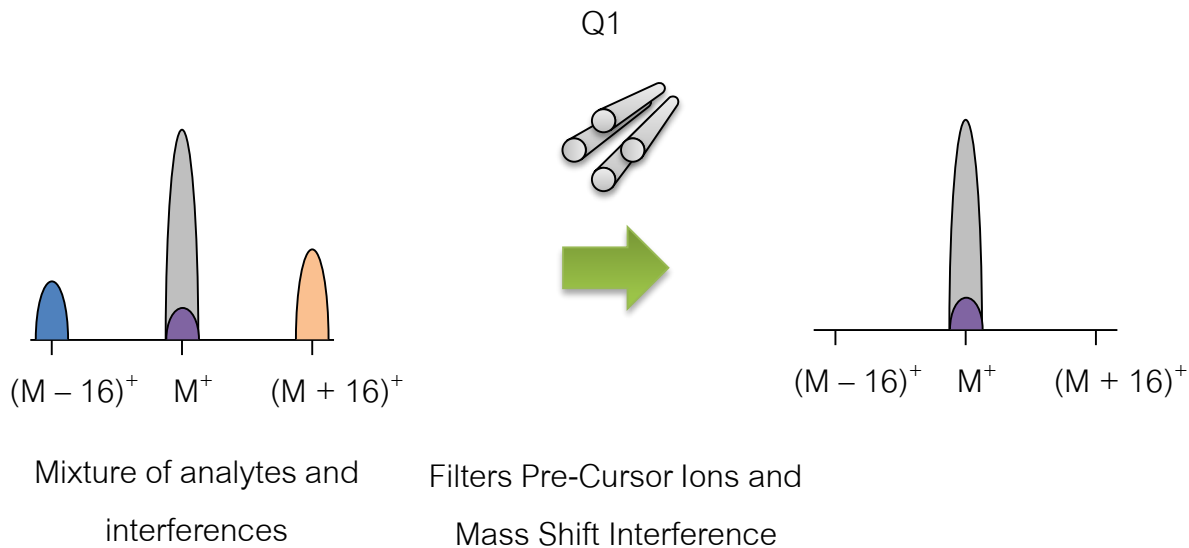


- Precursor Ion
- Analyte
- Isobaric Interference
- Mass Shift Interference



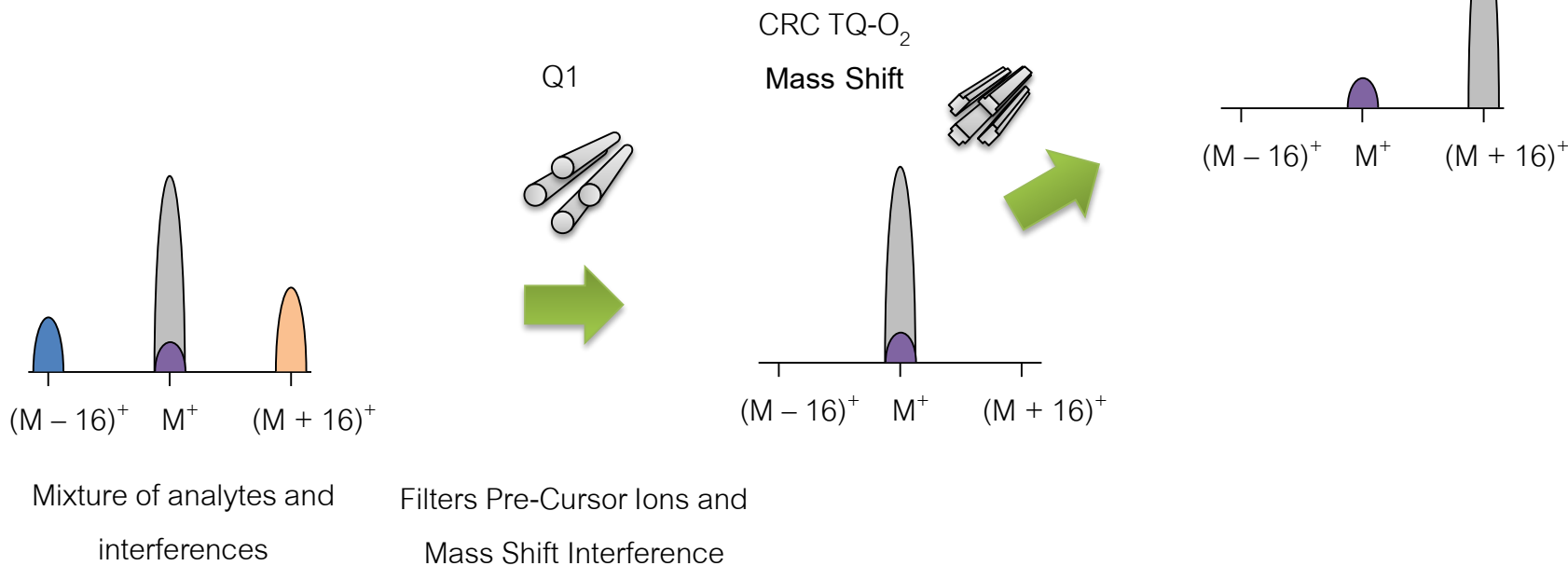
Mixture of analytes and interferences

- Precursor Ion
- Isobaric Interference
- Analyte
- Mass Shift Interference



Interference removal using TQ Reaction Chemistry (with O₂)

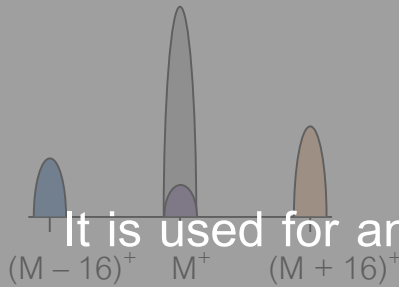
- Precursor Ion
- Isobaric Interference
- Analyte
- Mass Shift Interference



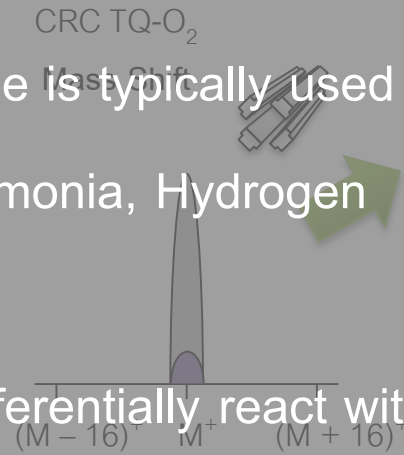
- Precursor Ion
- Analyte
- Isobaric Interference
- Mass Shift Interference

TQ Mass Shift mode is typically used with:

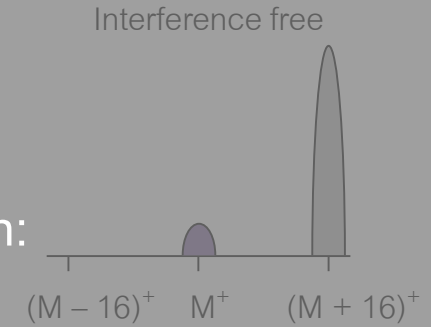
Oxygen, Ammonia, Hydrogen



Mixture of analytes and interferences



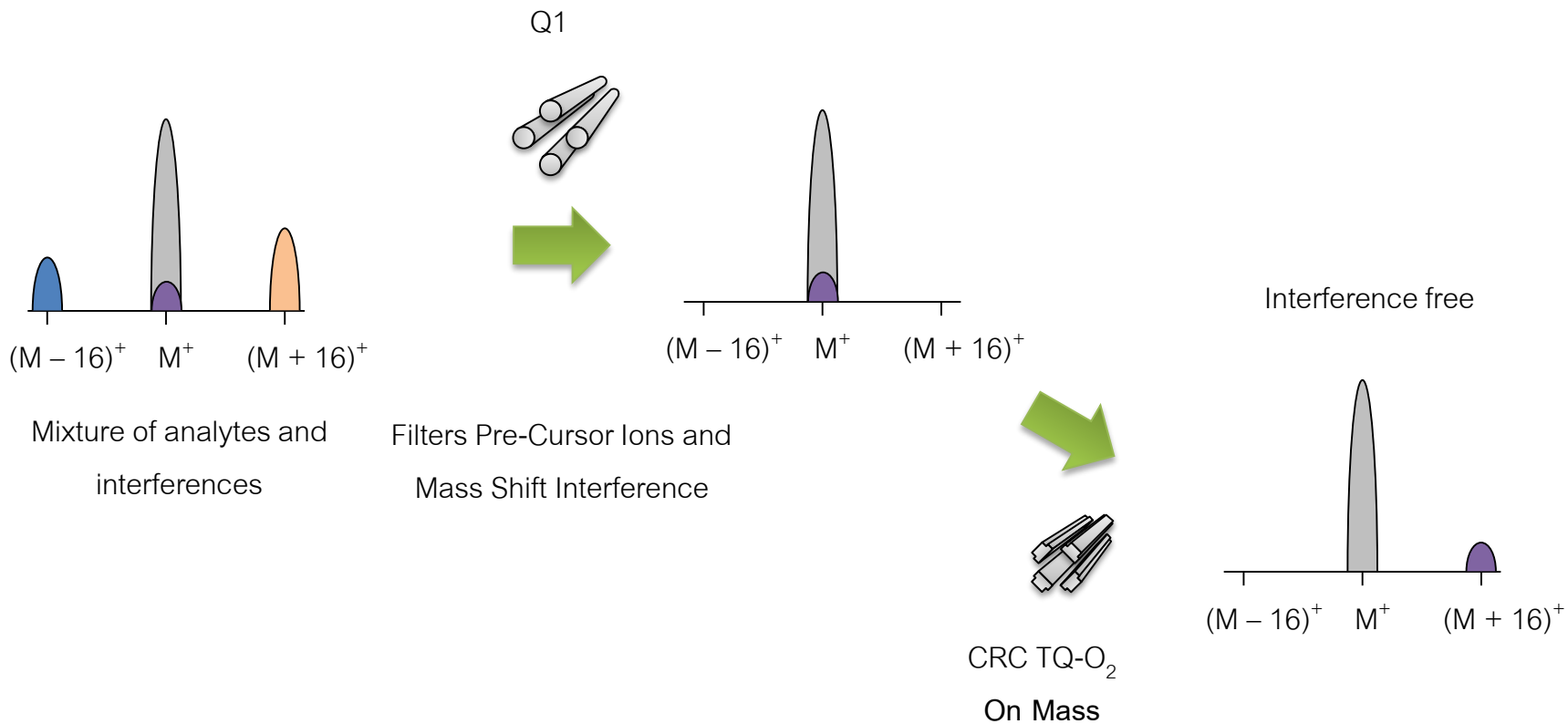
Filters Pre-Cursor Ions and Mass Shift Interference



It is used for analytes which preferentially react with the reactive gas


Interference removal using TQ Reaction Chemistry (with O₂)

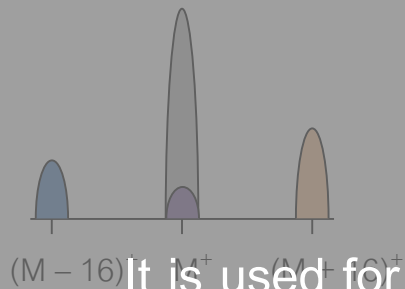
- Precursor Ion
- Isobaric Interference
- Analyte
- Mass Shift Interference



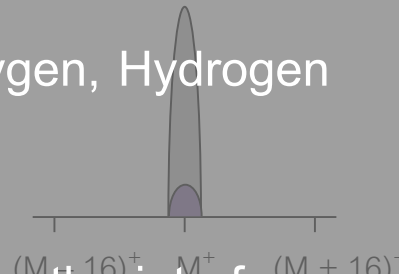
- Precursor Ion
- Isobaric Interference
- Analyte
- Mass Shift Interference

Q1
TQ On Mass mode is typically used with:

 Oxygen, Hydrogen

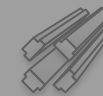


Mixture of analytes and interferences

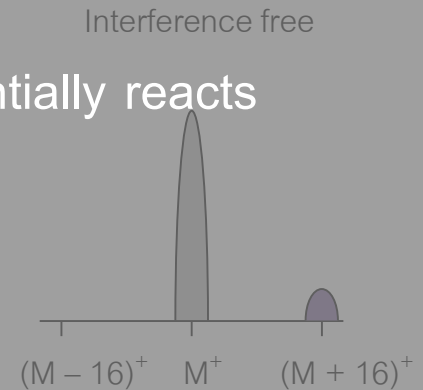


Filters Pre-Cursor Ions and Mass Shift Interference

It is used for analytes where the interference preferentially reacts with the reactive gas



CRC TQ-O₂
On Mass



Interference free



Environmental Analysis – As, Se

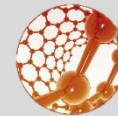


Clinical Research - Ti



Metallurgy

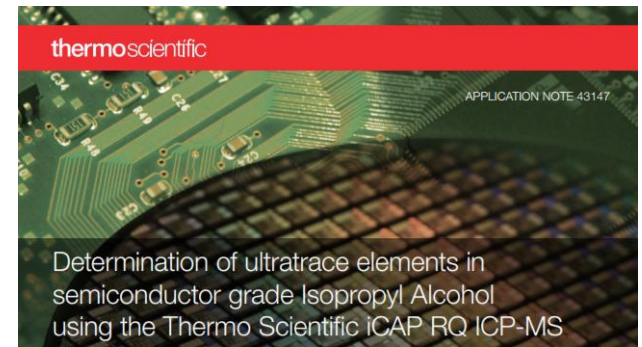
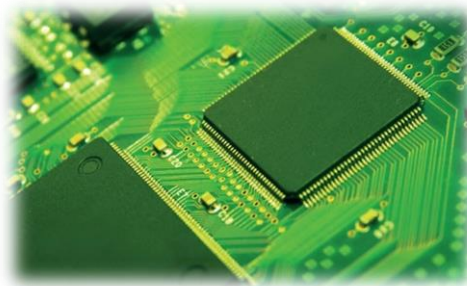
Advanced Applications



Speciation Analysis

Laser Ablation - Imaging

- Isopropyl alcohol (IPA) is used to solvent clean wafers during production in the semiconductor industry
- IPA has been considered a difficult matrix to analyze directly by ICP-MS
 - high volatility
 - low viscosity
 - high carbon content
- Kinetic energy discrimination (KED) can remove carbon based interferences from the sample matrix and argon based interferences from the ICP



Authors

Tomoko Vincent¹, Vicki Wu², Julian D. Wills³, Lothar Rottmann¹,
¹Thermo Fisher Scientific, Germany, ²Joy Allied Technology, Taiwan.

Keywords

Cold plasma, ICAP RQ, ICP-MS, IPA, organics, semiconductor

Goal

To determine ultratrace metal concentrations in semiconductor grade isopropyl alcohol (IPA). Use cold plasma to reduce background equivalent concentrations (BEC) and detection limits (LoD) to demonstrate reproducible ultratrace (ng-L⁻¹) measurements. Demonstrate the reliable switching between hot and cold plasma within a single measurement to maximize sample throughput.

Introduction

Isopropyl alcohol (IPA) is used to solvent clean wafers during production in the semiconductor industry. As IPA comes into direct contact with wafer surfaces, it must be controlled for its trace metal purity. Because of its high elemental sensitivity, ICP-MS is widely used in quality control analyses of materials used in the semiconductor industry. A direct ICP-MS technique for the analysis of IPA would provide a useful control for ultratrace (ng-L⁻¹) levels of analytes in IPA and avoid any contamination caused by sample preparation.

IPA has historically been considered a difficult matrix to analyze directly by ICP-MS due to its high volatility, low viscosity and high carbon content. In this study instead of using kinetic energy discrimination (KED) to remove carbon based interferences from the sample matrix and argon based interferences from the ICP, cold plasma was employed. With this approach the ICP ion source is run at a significantly lower power, effectively suppressing the ionization of argon and carbon and therefore eliminating interfering polyatomic species that would otherwise interfere with target analyte ions. This approach is particularly effective for the alkali metals and permits their direct analysis at the ultratrace concentration levels required by the semiconductor industry.

Parameter	Value
Hot Plasma Power	1350 W
Cold Plasma Power	800 W
Spraychamber	Quartz cyclonic
Peltier Temperature	-10 °C
Hot Plasma Nebulizer Gas Flow	0.7 L·min ⁻¹
Cold Plasma Nebulizer Gas Flow	1.0 L·min ⁻¹
Oxygen Gas Flow	50 mL·min ⁻¹
Nebulizer	MicroFlow PFA-100 (self-aspirating)
Injector	1.0 mm I.D., quartz
Interface	Platinum sampler and high sensitivity platinum skimmer
Dwell Time	100 ms per peak, 10 sweeps

Identifier	Dwell time (s)	Measurement mode	Channels	Spacing (u)
7Li (Cold Plasma)	0.1	Cold Plasma	1	0.1
23Na (Cold Plasma)	0.1	Cold Plasma	1	0.1
24Mg (Cold Plasma)	0.1	Cold Plasma	1	0.1
40Ca (Cold Plasma)	0.1	Cold Plasma	1	0.1
39K (Cold Plasma)	0.1	Cold Plasma	1	0.1
27Al (Cold Plasma)	0.1	Cold Plasma	1	0.1
▶ 74Ge (Hot Plasma)	0.1	Hot Plasma	1	0.1

Advanced Parameters	
Number of sweeps:	<input type="text" value="10"/>
Measurement order:	<input type="text" value="Cold Plasma"/> <input type="text" value="Hot Plasma"/>

- Standard concentration : 20, 50, 100 and 200 ng/L
- IPA sample was spiked with 100 ng/L
- Measurement of **26 elements** at ultratrace concentrations in IPA was achieved in **less than 5 minutes**. (sample uptake, analysis and washout as well as the switching time between hot and cold plasma within each measurement)
- Low ionization potential (IP) elements : Li, Na, Mg, K, Ca, Al as well as first row transition metals; Cr, Fe are measured with low backgrounds and high sensitivities in **cold plasma**
- Higher IP elements are analyzed in hot plasma

Mg m/z 24

Parameter	Value
b	155.459
a	120.667
R	1.000
BEC	0.776 ppt
IDL(LOD)	0.541 ppt

Calibration data

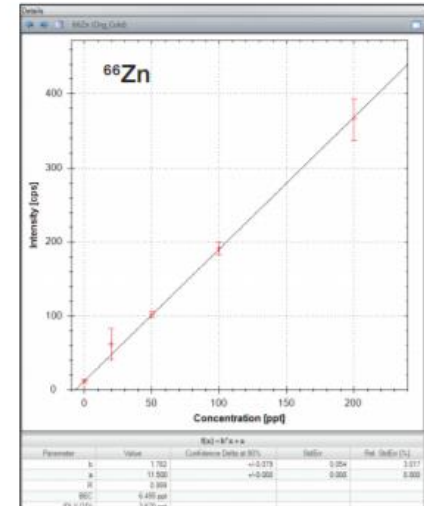
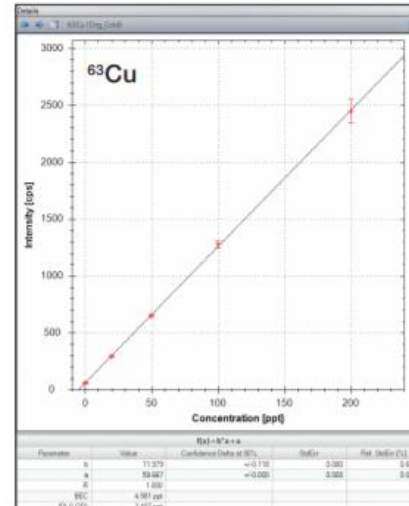
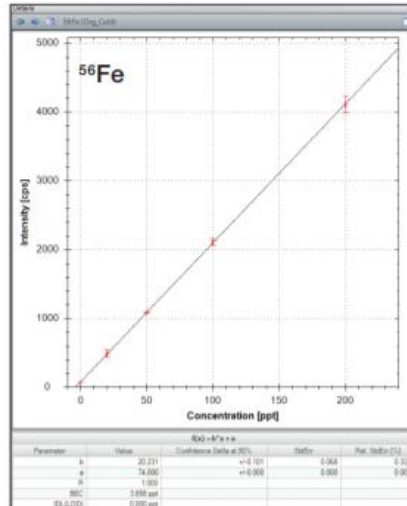
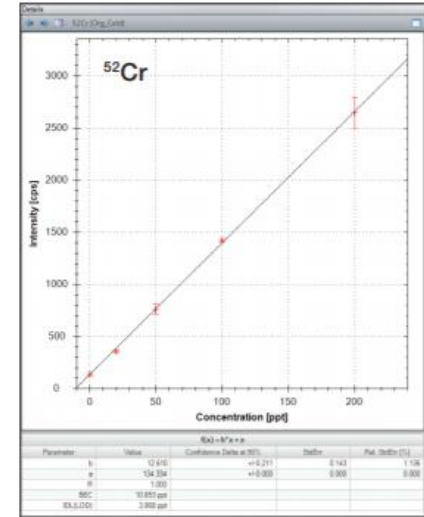
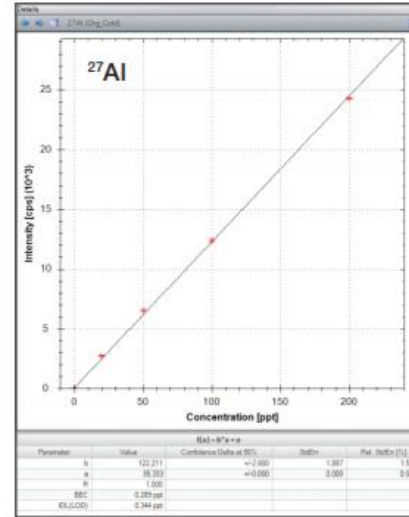
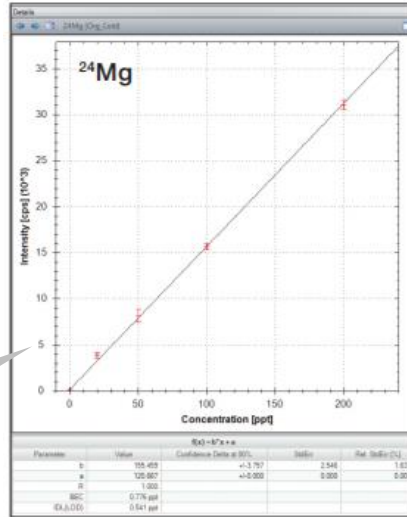


Figure 3. Cold plasma calibration curves with points 20, 50, 100 and 200 (ng·L⁻¹).

- Mg at m/z 24 in IPA under hot plasma conditions is complicated by a carbon dimer species (12C₂)

- Cold plasma can remove interference at m/z 24 and provide the **sub ng/L detection limit (LOD)** and background equivalent concentration (BEC)

	Analysis Mode	BEC (ng·L ⁻¹)	LoD (ng·L ⁻¹)	Recovery (%)
⁷ Li	Cold	0.031	0.007	99%
¹¹ B	Hot	107	29	89%
²³ Na	Cold	10	0.8	101%
²⁴ Mg	Cold	0.78	0.54	101%
²⁷ Al	Cold	0.29	0.34	97%
³⁹ K	Cold	14	0.31	97%
⁴⁰ Ca	Cold	7.4	3.1	118%
⁵¹ V	Hot	4.0	1.4	100%
⁵² Cr	Cold	11	3.9	97%
⁵⁵ Mn	Cold	7.3	1.6	97%
⁵⁶ Fe	Cold	3.7	< 0.001	98%
⁵⁸ Ni	Cold	0.40	0.52	97%
⁵⁹ Co	Cold	1.9	1.2	103%
⁶³ Cu	Cold	5.0	2.4	106%
⁶⁶ Zn	Cold	6.5	3.7	109%
⁷⁴ Ge	Hot	5.1	3.9	90%
⁷⁵ As	Hot	7.6	11	107%
¹⁰⁷ Ag	Cold	0.41	0.42	116%
¹¹¹ Cd	Hot	0.26	0.39	104%
¹¹⁵ In	Cold	0.01	0.04	100%
¹²⁰ Sn	Hot	0.10	0.001	103%
¹³⁸ Ba	Hot	0.014	0.014	102%
¹⁹⁷ Au	Hot	1.0	1.4	86%
²⁰⁵ Tl	Cold	0.07	0.22	100%
²⁰⁸ Pb	Cold	0.13	0.39	99%
²⁰⁹ Bi	Cold	0.001	0.001	98%

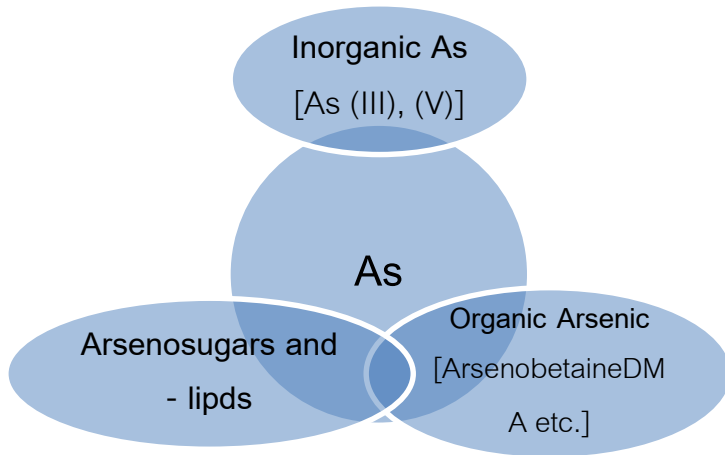
- ✓ Fast, reliable, in measurement switching between hot and cold plasma even for volatile organic solvents



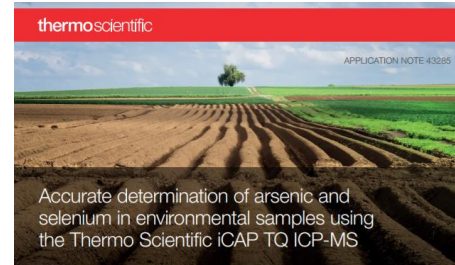
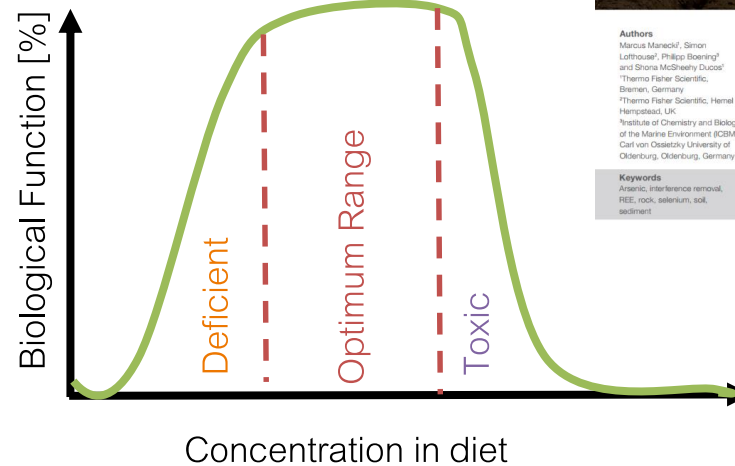
Thermo Scientific iCAP RQ ICP-MS.

The role of As and Se in the environment

Arsenic: A potential hazard in the food chain



Selenium: An essential nutrient



Authors
 Marcel Manekf, Simon Lofthouse¹, Philipp Boering² and Shona McSheehy Duco¹
¹Thermo Fisher Scientific, Bremen, Germany
²Thermo Fisher Scientific, Hemel Hempstead, UK
³Institute of Chemistry and Biology of the Marine Environment (ICBM), Carl von Ossietzky University of Oldenburg, Oldenburg, Germany

Goal
 To demonstrate the accurate determination of arsenic and selenium in sediments and rocks that contain elevated levels of rare earth elements using triple quadrupole ICP-MS.

Introduction
 Due to the impact arsenic and selenium can have in the environment at low levels, as a toxin or essential nutrient respectively, it is important to be able to quantify them accurately.

Selenium for example is an essential element that is necessary for normal thyroid function and due to its antioxidant properties, is associated with several health benefits. Diseases associated with selenium deficiency such as Keshan disease and symptoms of hypothyroidism, are most commonly found in areas where levels of selenium in soil are particularly low. Supplementation as a remedy is common practice and is not isolated to humans. Understanding where soil selenium deficiencies occur for example supports the correct supplementation of cattle grazing in those areas to prevent white muscle disease (a cattle specific selenium deficiency disease).

Keywords
 Arsenic, interference removal, REE, rock, selenium, soil, sediment

ThermoFisher
 SCIENTIFIC

- Many different chemical forms (species) are known that strongly differ in toxicity and bioavailability
- Plants such as rice are well known for high accumulation of As from soils
- Knowledge of Se content in soil may prevent Se deficiency in both human and animal populations

- Calibration standards of arsenic and selenium : Concentrations of 0.2, 0.5, 1, 2 and 5 $\mu\text{g/L}$
- 1 ppm of Dy, Gd, Nd, Sm and Tb added
- 35 mg of a marine sediment sample, collected from the deep Pacific Ocean + 50 mg of the andesite reference standard AGV-1 and treated in closed PTFE vessels with concentrated HNO_3 overnight to oxidize any organic matter (if present).
- Concentrated HF (1.5 mL) and HClO_4 (1.5 mL) added
- Heated in a hot block for 12 h at 180 °C.
- After digestion, the acids were evaporated on a hot plate at 180 °C to near dryness.
- The residues were re-dissolved, fumed off three times with 6N HCl to near dryness and finally taken up in 10 mL 1N HNO_3
- Prior to analysis, both samples were further 1:10 diluted with 1% HNO_3 /2% MeOH
- Lutetium was added at a concentration of 1 $\mu\text{g}\cdot\text{L}^{-1}$ as an internal standard
- The use of methanol is important in the analysis of arsenic and selenium due to the effect of carbon enhancement in the plasma which increases the ionization of both elements.

For different measurement modes were evaluated:

SQ-STD - Single quadrupole mode with no collision/reaction cell (CRC) gas.

SQ-H₂ - Single quadrupole mode with CRC pressurized with pure hydrogen as reaction gas

SQ-KED - Single quadrupole mode with CRC pressurized with helium as a collision gas and KED applied.

TQ-O₂ Triple quadrupole mode with CRC pressurized with oxygen as a reaction gas, Q1 set to analyte mass (M+) and Q3 set to product ion mass (MO+)



Interferences on As and Se: Ar_2 , ArCl – easy to remove using He KED but if REE are present...

Single Quad ICP-MS: KED

Typically enhances M^{2+} Interferences

Doubly charged ions (e.g. $^{156}\text{Gd}^{++}$) appear at m/z 78 and hence interfere with the detection of $^{78}\text{Se}^+$

As and Se,
m/z 75, 78, 80

Rare Earth Elements, m/z 140-176
→ ^{150}Sm , ^{156}Gd , ^{160}Gd

m/z

Interferences on As and Se: Ar_2 , ArCl – easy to remove using He KED but if REE are present...

Single Quad ICP-MS: KED

Typically enhances M^{2+} Interferences

Solution:

Mass shift As and Se using O_2

Doubly charged ions (e.g. $^{156}\text{Gd}^{++}$) appear at m/z 78 and hence interfere with the detection of $^{78}\text{Se}^+$

As and Se,
m/z 75, 78, 80



$^{75}\text{As}^{16}\text{O}^+$, $^{78}\text{Se}^{16}\text{O}^+$, $^{80}\text{Se}^{16}\text{O}^+$
@ m/z 91, 94 and 96

Rare Earth Elements, m/z 140-176
→ ^{150}Sm , ^{156}Gd , ^{160}Gd

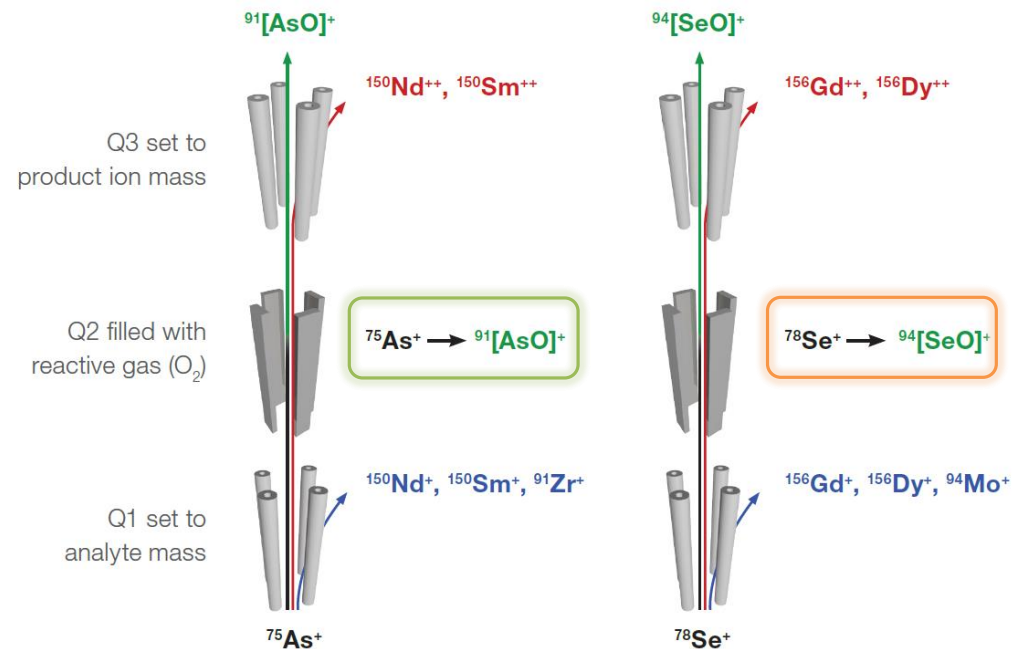
m/z

Mass shift reactions that move the analyte of interest to a different m/z

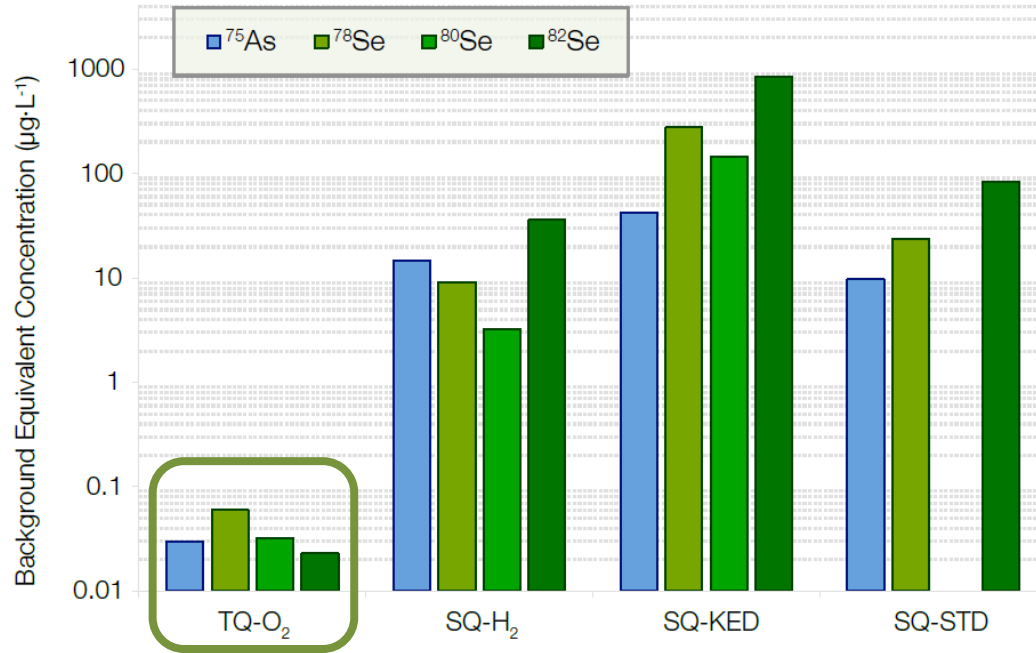
- Control ions entering the cell using Q1
- Use O₂ to efficiently convert As and Se to AsO and SeO in Q2 – the REE⁺⁺ don't react
- Selectively detect AsO and SeO free from REE⁺⁺ interference, using Q3

Type	⁷⁵ As	Method to remove
Polyatomic	⁴⁰ Ar ³⁵ Cl	KED
	⁴⁰ Ca ³⁵ Cl	
Isobaric	¹⁵⁰ Nd ⁺⁺	O ₂
	¹⁵⁰ Sm ⁺⁺	

Type	⁷⁸ Se	Method to remove
Polyatomic	⁴⁰ Ar ³⁸ Ar	KED, H ₂
Isobaric	¹⁵⁶ Gd ⁺⁺	O ₂
	¹⁵⁶ Dy ⁺⁺	



Interference removal capability



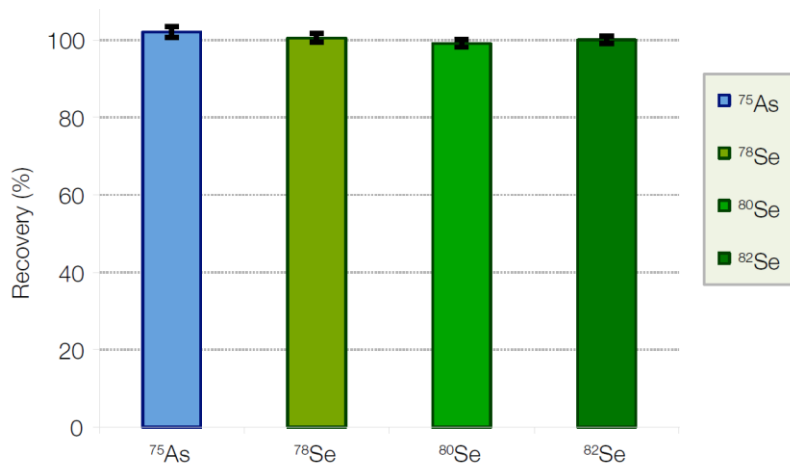
- Increased BECs observed for all SQ-modes due to unresolved doubly charged REE interferences
- Hydrogen is suitable for removing Ar based polyatomics, but is not capable of fully removing REE²⁺ interferences
- TQ-O₂ mode shows dramatically lower BEC values for both As and Se

Sample analysis results

AGV-1	Content in original sample ($\mu\text{g}\cdot\text{g}^{-1}$)	Certified content ($\mu\text{g}\cdot\text{g}^{-1}$)
^{75}As	0.892	0.88
^{78}Se	< LOQ	-
Deep Sea Sediment		
^{75}As	1.303	-
^{78}Se	0.109	-

Spike recovery in REE matrix solution

(1 ppb As and Se)



Spike recovery results in samples (1 ppb As and Se)

Analyte	AGV-1	Sediment
Arsenic	94.6 %	97.6 %
Selenium	93.4 %	97.6 %

Focus on the development of a robust method for the analysis of **titanium and other trace elements** in **human serum** reference materials using TQ ICP-MS.



Total elemental analysis in clinical research using the Thermo Scientific iCAP TQ ICP-MS

Authors

Tomoko Vincent, Applications Specialist
Thermo Fisher Scientific, Bremen, Germany

Keywords

Clinical research, isobaric interferences, serum, titanium, trace elemental analysis, urine

Introduction

Trace element analysis of biological samples provides significant information to support clinical research and forensic toxicology. An interesting example of trace elemental analysis for clinical research purposes is exploring the degradation of titanium based orthopedic and dental implants in humans. Following recent research on the possible carcinogenic effects of titanium dioxide the fate of titanium in the human body has become a growing area of clinical research focus. To support this there is a need for the development of robust analytical methods for the identification and quantification of titanium in a range of samples such as human body fluids and organs.

However, the development of such a method is challenging due to the low concentration of titanium in these types of samples and the potential isobaric interferences which single quadrupole ICP-MS cannot remove.

Advancements in ICP-MS technology have led to the development of triple quadrupole (TQ) ICP-MS instruments, which have the required sensitivity as well as the capability to resolve isobaric interferences resulting from polyatomic and isotopic species.



Sample preparation

- Certified reference materials (Seronorm™ Trace Elements in Serum L-1 and L-2 and volunteered human urine
- Samples were diluted 10 fold in 0.5% nitric acid and 2% tetramethylammonium hydroxide (TMAH) and ultra-pure water
- Calibration blank, a series of standards and a Quality Control (QC) were prepared using the same procedure
- All samples and standards were spiked with an internal standard mix (10 µg/L Ge, Y, Rh, Te and Bi)

Unit: mg·L⁻¹

	Major STD1	Major STD2	Major STD3	Major STD4	QC CCVs
Ca	5	10	25	50	10
Fe	0.1	0.2	0.5	1	0.2
Mg	5	10	25	50	10
P	5	10	25	50	10
K	5	10	25	50	10
S	50	250	500	1000	100
Na	50	100	250	500	100

Unit: µg·L⁻¹

	Minor STD1	Minor STD2	Minor STD3	Minor STD4	QC CCVs
Sb	0.5	1	2.5	5	1
As	0.1	0.2	0.5	1	0.2
Ba	5	10	25	50	10
Cd	0.1	0.2	0.5	1	0.2
B	5	10	25	50	10
I	5	10	25	50	10
Pb	0.1	0.2	0.5	1	0.2
Li	500	1000	2500	5000	1000
Mo	0.1	0.2	0.5	1	0.2
Rb	0.5	1	2.5	5	1
Sr	5	10	25	50	10
Ti	0.5	1	2.5	5	1
U	0.005	0.01	0.025	0.05	0.01
V	0.1	0.2	0.5	1	0.2
Zn	50	100	250	500	100
Se	0.1	0.2	0.5	1	0.2
Al	50	100	250	500	100
Cr	0.5	1	2.5	5	1
Mn	5	10	25	50	10
Ni	5	10	25	50	10
Co	0.1	0.2	0.5	1	0.2

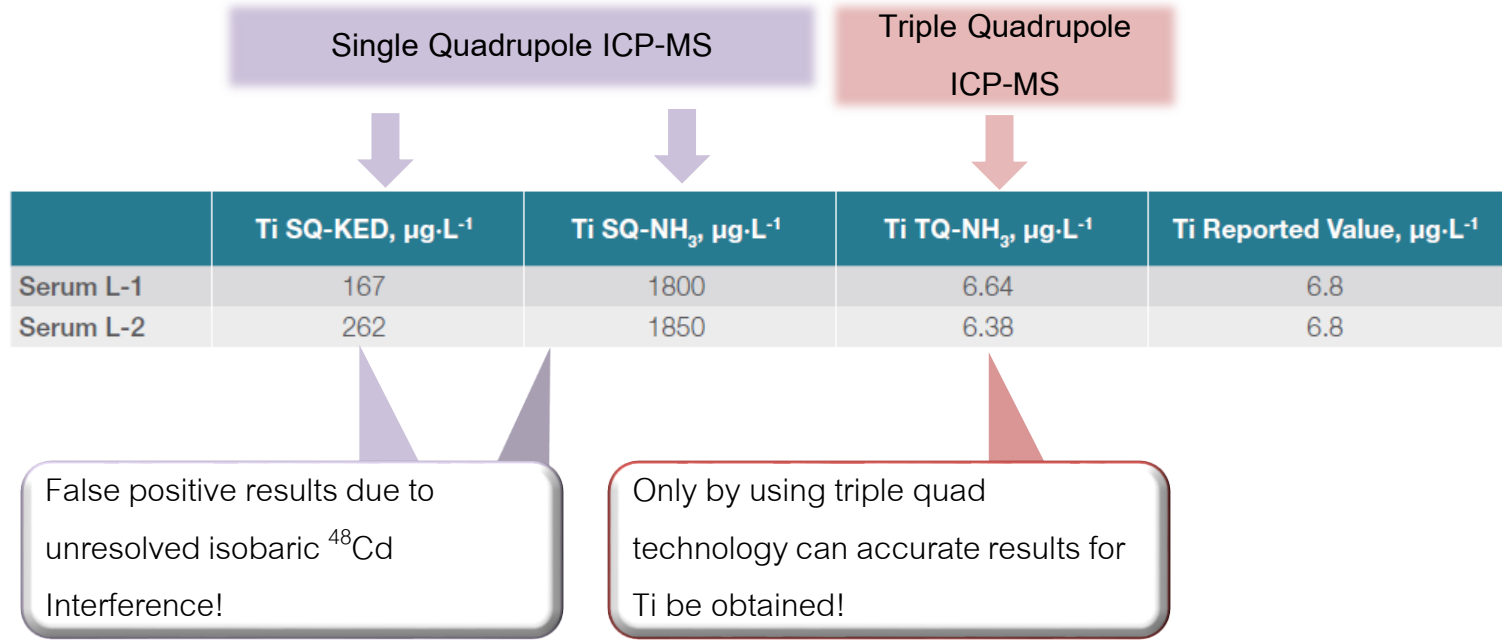
Recovery of Ti in Serum using Different Modes

Single Quadrupole ICP-MS

Triple Quadrupole
ICP-MS

	Ti SQ-KED, $\mu\text{g}\cdot\text{L}^{-1}$	Ti SQ-NH ₃ , $\mu\text{g}\cdot\text{L}^{-1}$	Ti TQ-NH ₃ , $\mu\text{g}\cdot\text{L}^{-1}$	Ti Reported Value, $\mu\text{g}\cdot\text{L}^{-1}$
Serum L-1	167	1800	6.64	6.8
Serum L-2	262	1850	6.38	6.8

Recovery of Ti in Serum using Different Modes



- Excellent agreement with certified value obtained for Ti
- Attainable detection sensitivity allowed to detect low concentrations of Ti in the prepared sample solution

Recovery of Ti in Serum using Different Modes

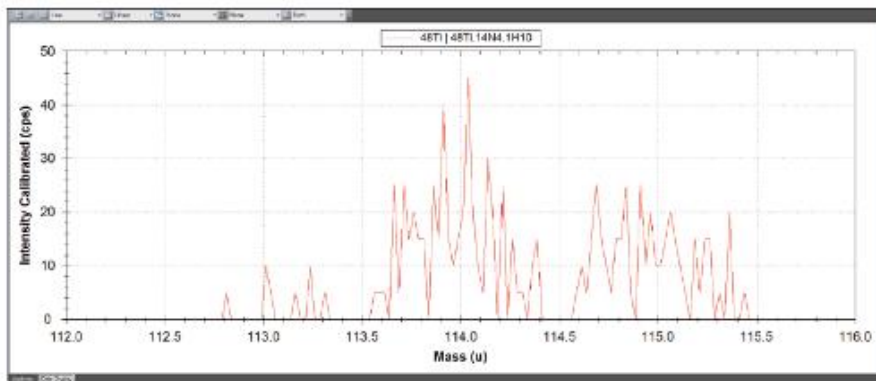
Single Quadrupole ICP-MS

Triple Quadrupole ICP-MS

	Ti SQ-KED, $\mu\text{g}\cdot\text{L}^{-1}$	Ti SQ-NH ₃ , $\mu\text{g}\cdot\text{L}^{-1}$	Ti TQ-NH ₃ , $\mu\text{g}\cdot\text{L}^{-1}$	Ti Reported Value, $\mu\text{g}\cdot\text{L}^{-1}$
Serum L-1	167	1800	6.64	6.8
Serum L-2	262	1850	6.38	6.8

False positive results due to unresolved isobaric ⁴⁸Ca Interference!

Only by using triple quad technology can accurate results for Ti be obtained!



Background signal on ⁴⁸Ti¹⁴N₄H₁₀ for a solution containing 10 mg·L⁻¹ of Cd

This would be a problem using single quadrupole instruments with NH₃!

Results – Multi-elemental analysis in urine and serum

- Excellent agreement to elements
- Full multi-elemental analysis run

	LOD	MDL	Serum L-1		Serum L-2		Urine
			Measured	Reference or reported value	Measured	Reference or reported value	Measured
Na*	0.0027	0.027	2743	2330-3504	3255	2820-4241	2977
Mg*	0.0001	0.0010	21.0	13.4-20.1	39.7	27.1-40.7	85.6
P*	0.0008	0.08	52.3	43.3-65.1	120	88-132	710
S*	0.145	1.3800	1100	1008	1495	1335	476
K*	0.0021	0.02	150	101-153	260	176-265	1946
Ca*	0.002	0.0200	90.1	69-104	124	95-143	99.8
Fe*	0.00002	0.00023	1.64	1.17-1.77	2.18	1.72-2.58	0.005
Li	1.13	11.2920	5778	4202-6320	10806	7739-11639	22.4
B	0.67	6.746	70.1	79.4	87	82.1	1548
Al	0.20	1.9670	54.2	25.2-75.7	122	96-144	13.7
V	0.002	0.022	1.04	1.10	1.26	1.10	0.229
Cr	0.008	0.0800	1.70	1.30-3.05	5.20	4.00-7.50	0.838
Mn	0.008	0.084	10.7	7.9-11.9	14.2	11.6-17.4	0.914
Co	0.0001	0.0010	1.38	0.67-1.57	2.16	2.13-3.97	0.027
Ni	0.006	0.055	6.26	3.38-7.9	9.41	7.9-11.9	1.45
Zn	0.051	0.5130	1052	844-1269	1527	1404-1831	359
As	0.002	0.018	0.383	0.400	0.374	0.380	1.31
Se	0.010	0.1000	80.8	51-120	124	95-176	7.31
Rb	0.004	0.035	4.20	4.40	8.70	8.70	812
Sr	0.006	0.0570	95.7	95.0	106	110	89.2
Mo	0.005	0.048	0.710	0.760	1.20	1.21	7.62
Cd	0.001	0.0100	0.130	0.130	0.140	0.140	0.229
Ti	0.002	0.02	6.64	6.80	6.38	6.80	0.151
Sb	0.006	0.0600	11.6	10.4	16.1	15.0	0.040
I	0.022	0.219	75.5	71.8	69.9	60.9	82.8
Ba	0.003	0.0300	172	190	133	139	2.09
Pb	0.0007	0.007	0.370	0.400	0.666	0.660	0.446
U	0.0001	0.0010	0.288	0.302	0.357	0.359	0.020

Use as a good example of TQ multi-element data...

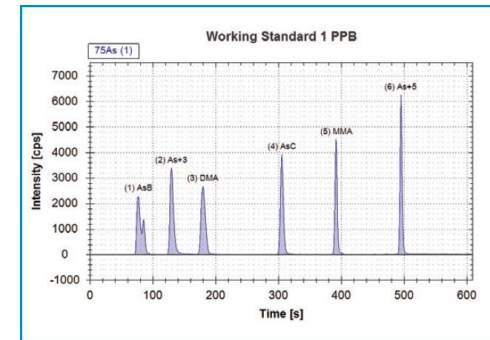
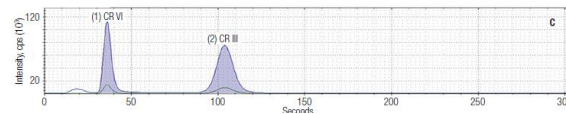
Hardware: IC and LC

IC is entirely metal free and thus ideally suited

Control over hardware: Chromeleon plug-in for Qtegra



- Data evaluation: tQuant



- Compound specific calibration
- Peak integration algorithms



AN43255 Determination of Inorganic **Arsenic** in Rice using IC-ICP-MS

AN43099 IC-ICP-MS speciation analysis of **As** in apple juice using the Thermo Scientific iCAP Q ICP-MS

AN43141 Total and speciation analysis of **Mercury** in contact lens solutions by ICP-MS

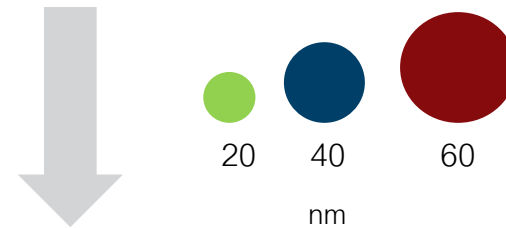
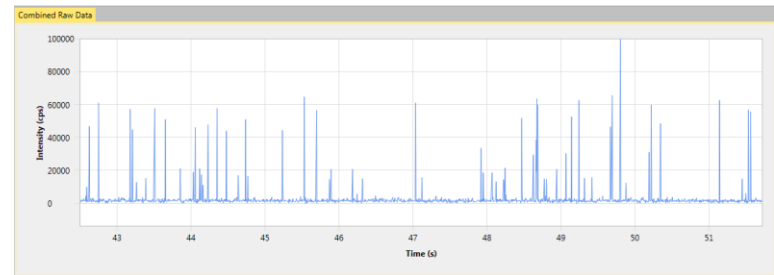
AN43227 Speciation of **Bromine** Compounds in Ozonated Drinking Water using Ion Chromatography and Inductively Coupled Plasma Mass Spectrometry

AN43098 Speciation Analysis of **Cr (III)** and **Cr (VI)** in Drinking Waters Using Anion Exchange

Chromatography Coupled to the Thermo Scientific iCAP Q ICP-MS

Concentrations	No.	Time	Sample Type	Label	AsB [ppb]	DMA [ppb]	AsIII [ppb]	AsC [ppb]	MMA [ppb]
1	3/14/2012 9:23:12 AM	BLK			N/A	N/A	0.001	-0.002	0.064
2	3/14/2012 9:45:30 AM	STD							
2	3/14/2012 9:45:30 AM	STD	STD 1		0.067 (0.072)	0.083 (0.088)	0.073 (0.074)	0.068 (0.073)	0.073 (0.076)
3	3/14/2012 10:07:42 AM	STD	STD 1		0.068 (0.072)	0.088 (0.088)	0.072 (0.074)	0.069 (0.073)	0.068 (0.076)
4	3/14/2012 10:29:49 AM	STD	STD 2		0.398 (0.401)	0.445 (0.438)	0.426 (0.414)	0.400 (0.398)	0.138 (0.427)
5	3/14/2012 10:51:55 AM	STD	STD 2		0.397 (0.401)	0.441 (0.438)	0.421 (0.414)	0.400 (0.398)	0.454 (0.427)
6	3/14/2012 11:14:03 AM	STD	STD 3		0.808 (0.801)	0.876 (0.898)	0.814 (0.823)	0.811 (0.808)	0.781 (0.842)
7	3/14/2012 11:36:15 AM	STD	STD 3		0.819 (0.801)	0.889 (0.899)	0.828 (0.823)	0.819 (0.808)	1.016 (0.842)
8	3/14/2012 11:58:16 AM	STD	STD 4		1.617 (1.618)	1.738 (1.793)	1.648 (1.658)	1.592 (1.608)	1.681 (1.683)
9	3/14/2012 12:20:27 PM	STD	STD 4		1.614 (1.618)	1.756 (1.793)	1.658 (1.658)	1.591 (1.608)	1.851 (1.683)
10	3/14/2012 12:42:36 PM	STD	STD 5		3.280 (3.282)	3.585 (3.563)	3.406 (3.401)	3.243 (3.261)	3.351 (3.413)
11	3/14/2012 1:04:36 PM	STD	STD 5		3.283 (3.282)	3.553 (3.563)	3.401 (3.401)	3.250 (3.261)	3.385 (3.413)
Calibrations									
12	3/14/2012 1:26:48 PM	UNKNOW	BLK		-0.002	0.012	0.001	-0.002	0.064
13	3/14/2012 1:40:01 PM	UNKNOW	Sample 1 Digest 1		0.001	0.144	0.104	-0.002	N/A
14	3/14/2012 2:11:07 PM	UNKNOW	Sample 1 Digest 1		0.000	0.148	0.107	-0.002	N/A
15	3/14/2012 2:33:07 PM	UNKNOW	Sample 2 Digest 2		0.000	0.149	0.149	-0.001	N/A
16	3/14/2012 2:56:22 PM	UNKNOW	Sample 2 Digest 2		0.000	0.151	0.107	-0.002	N/A
17	3/14/2012 3:17:19 PM	UNKNOW	Sample 2 Digest 3		0.002	0.154	1.225	-0.002	N/A
18	3/14/2012 3:39:34 PM	UNKNOW	Sample 2 Digest 3		-0.002	0.178	1.263	-0.002	N/A
19	3/14/2012 4:01:35 PM	UNKNOW	BLK		N/A	N/A	0.001	-0.002	N/A
20	3/14/2012 4:23:45 PM	UNKNOW	STD 3		0.040	1.020	0.162	0.161	1.058
21	3/14/2012 4:45:57 PM	UNKNOW	BLK		-0.002	0.012	N/A	-0.002	N/A
22	3/14/2012 5:08:02 PM	UNKNOW	Sample 2 Digest 1		-0.002	0.164	1.158	-0.001	N/A
23	3/14/2012 5:30:01 PM	UNKNOW	Sample 2 Digest 2		0.001	0.160	1.126	-0.002	N/A
24	3/14/2012 5:52:14 PM	UNKNOW	Sample 2 Digest 3		0.001	0.174	1.178	-0.001	N/A

- Nanoparticle analysis by sp-ICP-MS becoming a part of routine measurements
- spICP-MS data handling and processing for consistent and accurate interpretation of the data; to give the right result
- Hardware: High Sensitivity Interface and 100 uL/min nebulizer
- Software: npQuant plug-in



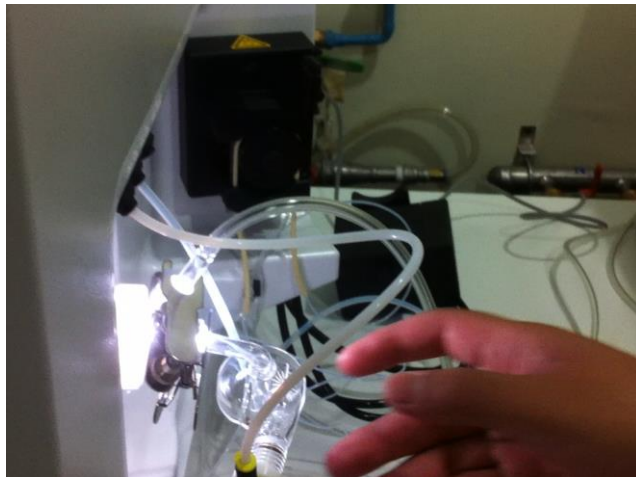
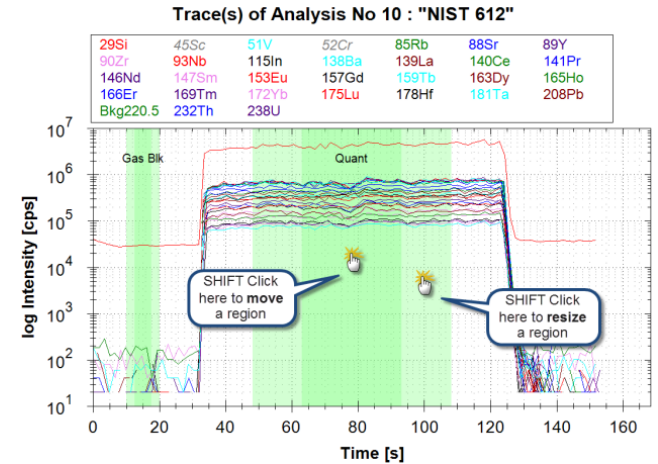
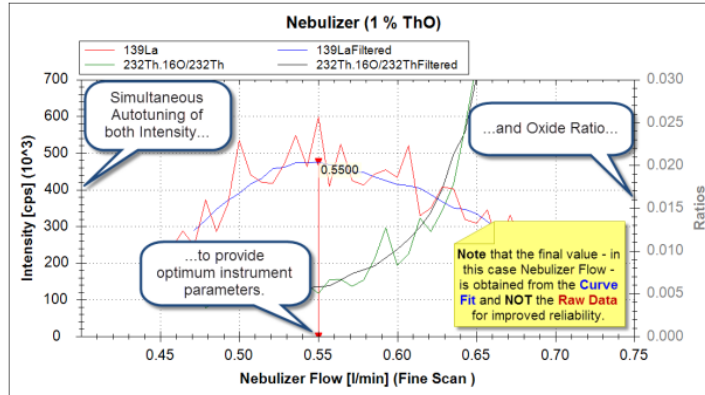
- ✓ Acquire and evaluate data for nanoparticle size characterization
- ✓ Automated determination of key input parameters
- ✓ Statistical evaluation of each data set to judge data quality

Verification of the Calculation Procedures in the npQuant Evaluation Module for Qtegra Intelligent Scientific Data Solution Software

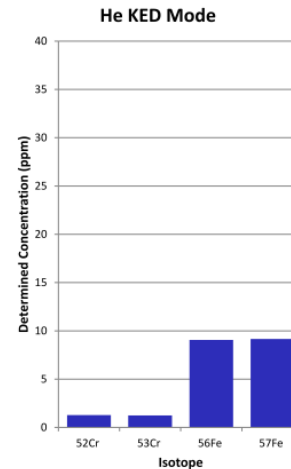
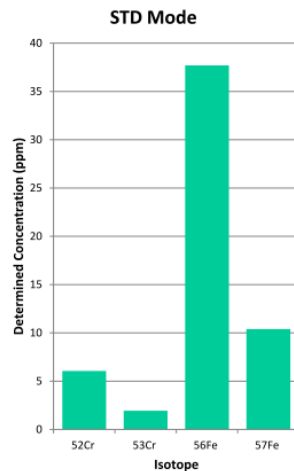
Daniel Kutscher, Shona McSheehy Ducos, Thermo Fisher Scientific, Bremen, Germany

Application Brief 43330

Fully automated optimization for LA-ICP-MS



The high base sensitivity of the iCAP RQ enables routine analysis in He KED LA-ICP-MS.



Results for Cr and Fe isotopes in LA-ICP-MS analysis of artificial sapphire.

Get Connected

www.scispec.co.th/

Follow Us

LINE

@scispec



<https://www.facebook.com/scispec/>



Questions?