MassARRAY: The Powerful Technology for genomic Analysis

Present By : MissChutipat Sriaimsaard



Адела

28 February 2018



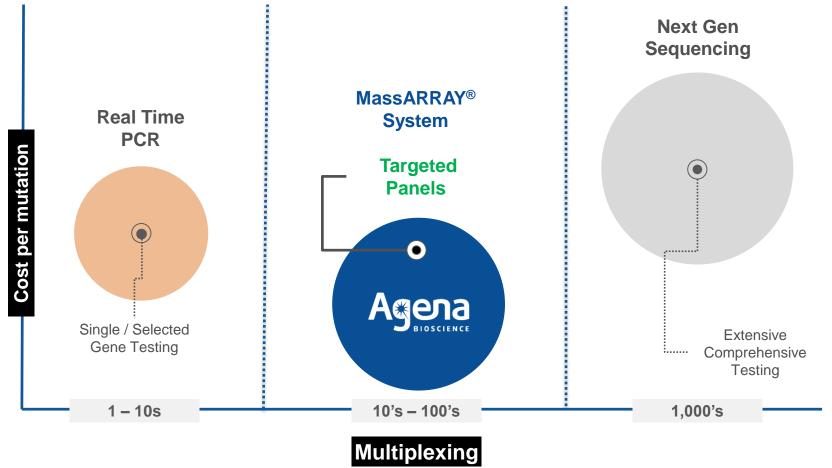


- Introduction to MassARRAY
 - -Principal
 - -Data analysis
 - -Assay design
- -Application of MassARRAY®
 - Agricultural Genomics





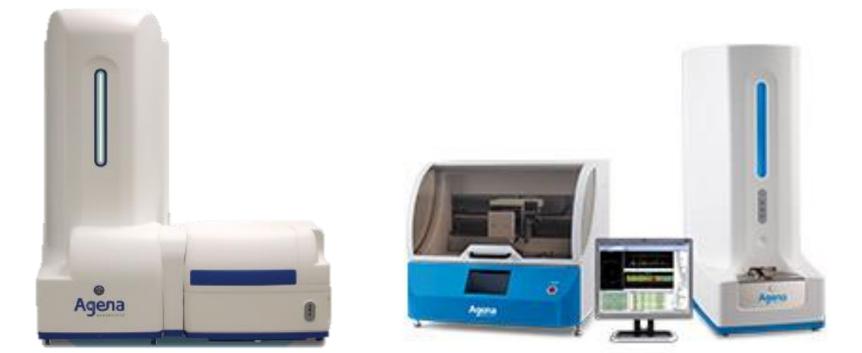
Cost-Effective, Targeted Genetic Analysis Robust, Flexible, and High Throughput







MassARRAY[®] System



Chip prep module 96 well plate

384-well plate platform





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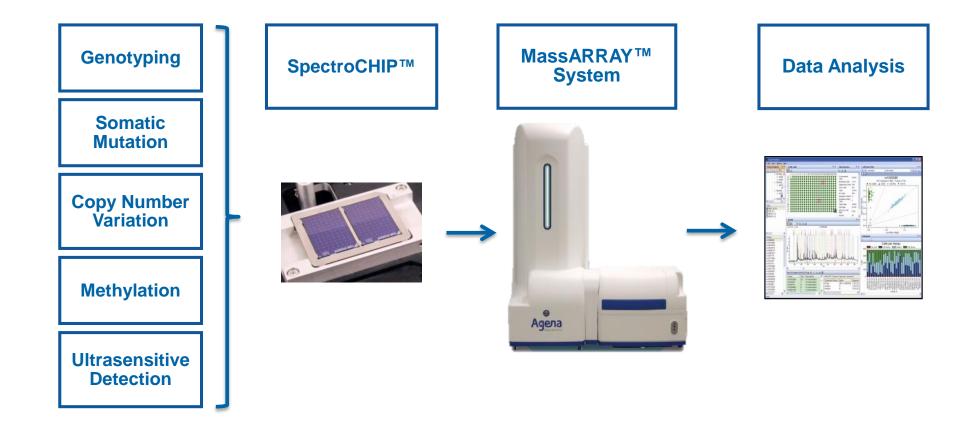
Proven Versatility of the MassARRAY[®] System



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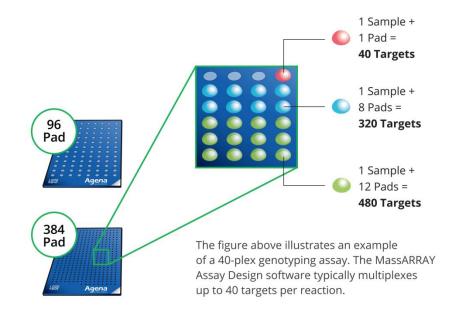
MassARRAY[™] Applications







A Dynamic and Flexible Platform A Single Platform with Modular Formats



Highly scalable throughput

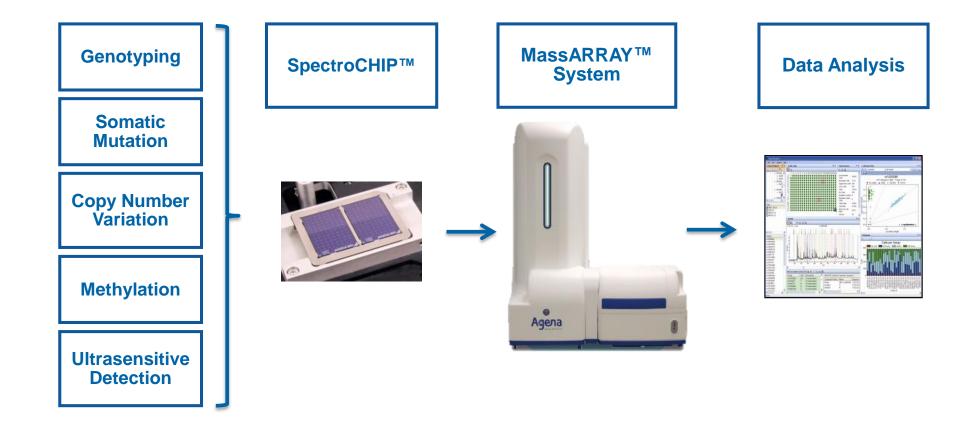
- 96 or 384-pad SpectroCHIP[®] Arrays*
- From 1 to 10 arrays per day
- 10s 100s 1000s samples per day

Marker x Sample flexibility

- Multiplex 5 40 targets per well
- Run 1 to 24 reactions per sample
- Screen 10s to 100s mutations per sample



MassARRAY[™] Applications







The MassARRAY[®] System for Genotyping Flexibility and Cost Efficiency in a Single Workflow

- Minimal input DNA -> 5-10 ng required per well
- Flexible sample types

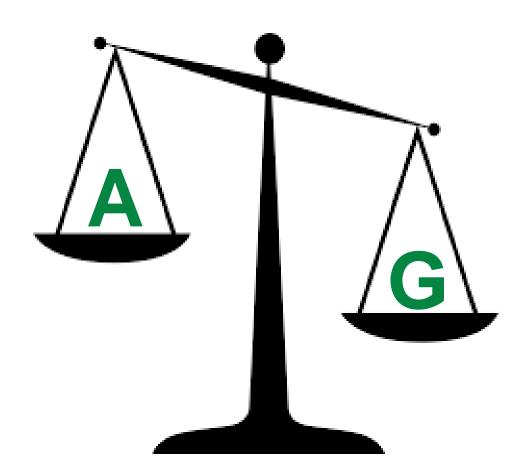


- root, seeds, leaf, etc
- Efficient multiplexing -> 5 40 targets per well





DNA Analysis Based on MassARRAY



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DNA consists of a fourletter alphabet: A, C, G, T

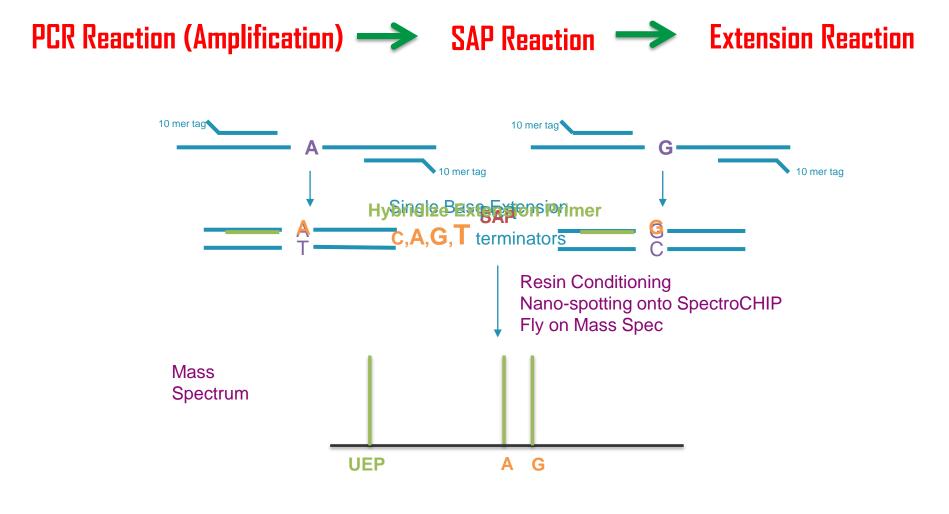
Each base within a DNA strand has defined molecular mass:

dCMP = 289.2 Da dTMP = 304.2 Da dAMP = 313.2 Da dGMP = 329.2 Da



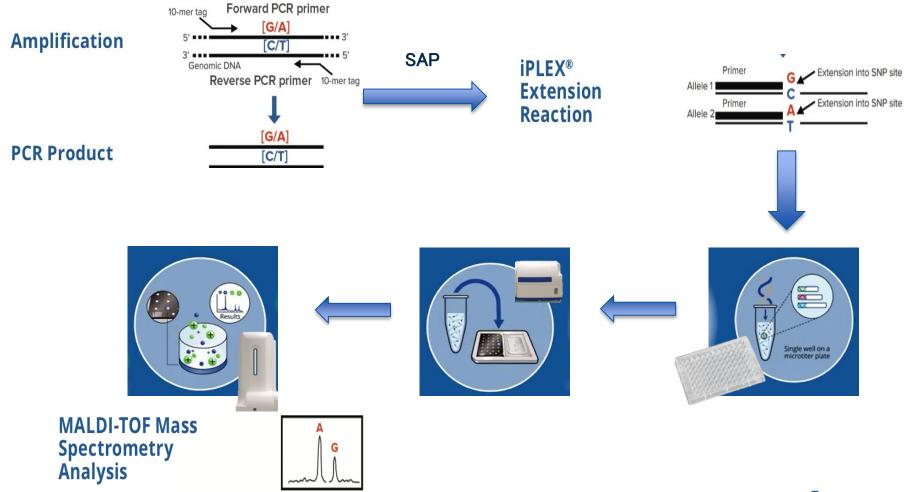
GENOTYPING Example of an [A/G] SNP

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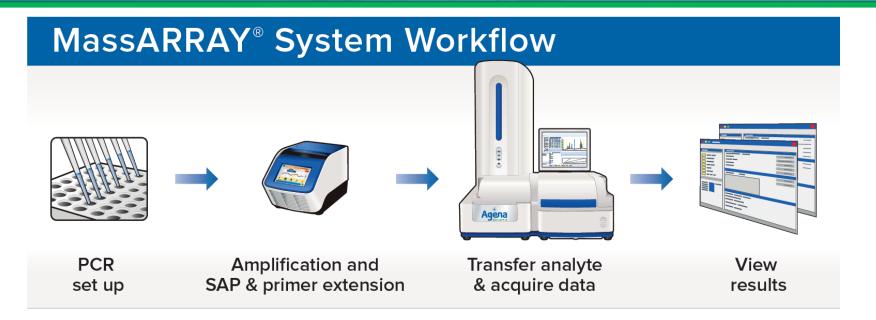


MassARRAY[®] Process



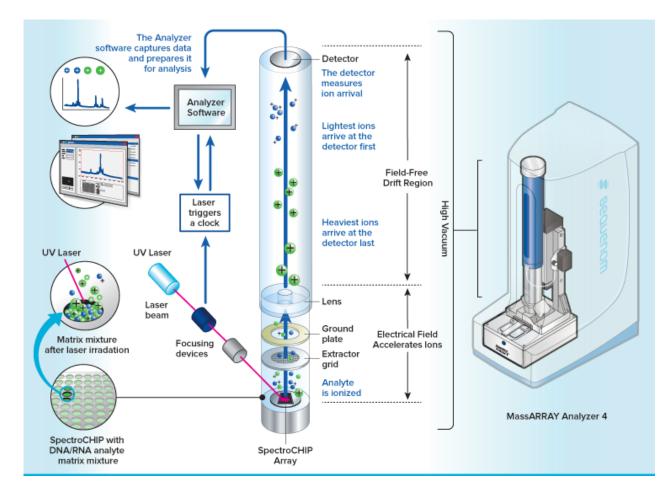


Easy Workflow From DNA to Results



- Flexible sample & assay set-up
- ~ 8 hours from DNA to results
- Simple data output on 10's- 100's actionable variants
- 96 or 384 well formats



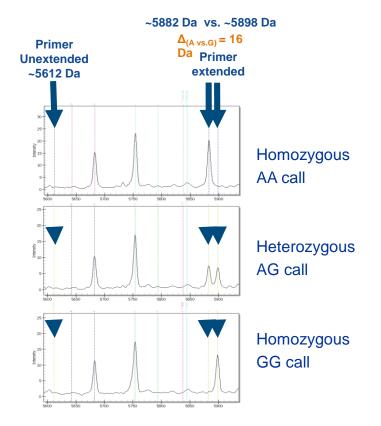


- Molecular mass is a unique intrinsic property.
- The mass of a nucleic acid string is determined by its length and composition (ATCG).



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Mass spectra of 3 different SNP [A/G] samples



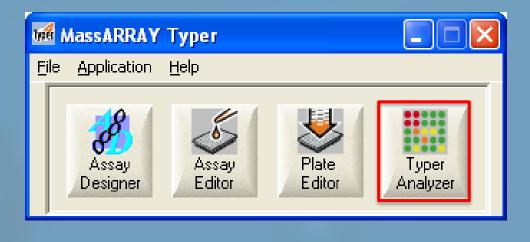
- MALDI-TOF MS of iPLEX reaction products shows peaks for the unextended primer (5612 Da) and extended primers (1 nucleotide)
- Heterozygous sample show two peaks, one at 5882 Da and the other at 5898 Da
- First two peaks are from other SNP assays in the same multiplex reaction.



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Data Analyzer

Software application suite for Genotyping & Somatic Mutation Analysis

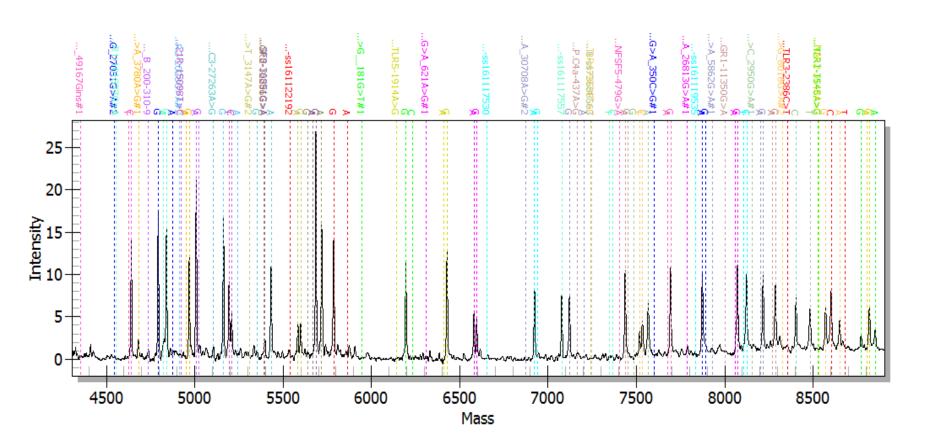




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Typical SNP Panel (31-plex)

The Power of Multiplexing Multiple Markers in a Single Well







Generating Report

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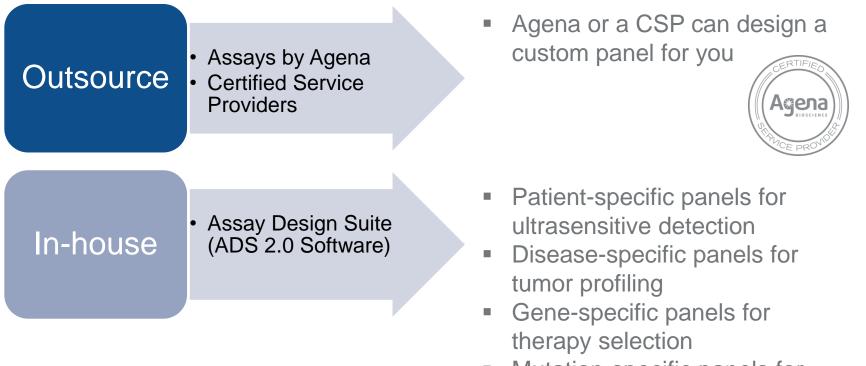
Assay Design



Assay Design Services

Fully customized options

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 Mutation-specific panels for validation



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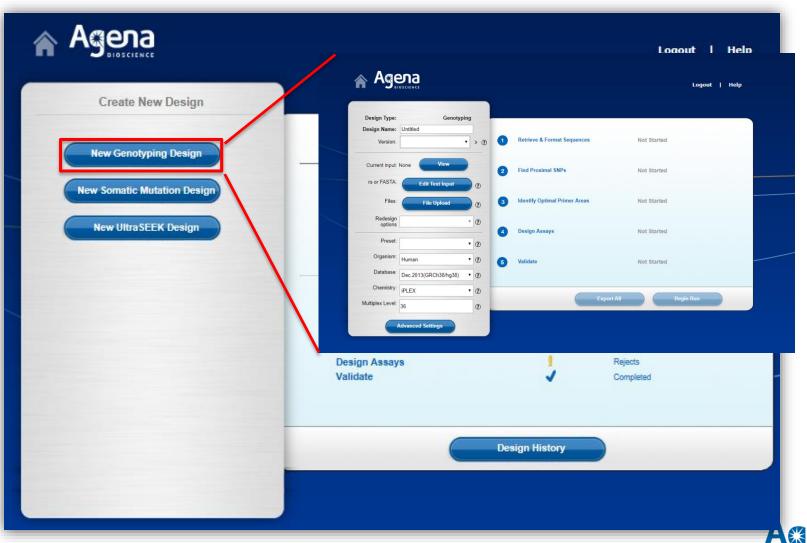


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	LAUNCH Assay Design Suite is a comprehensive and powerful tool for designing genotyping, somatic mutation, and ultra sensitive assays. The user- friendly interface integrates the design steps of importing RS numbers or sequences, retrieving and formatting sequences, finding proximal SNPs, identifying optimal areas for PCR primers, designing, and verifying multiplexed assays into a single software package. Assay Design Suite also enables users to design control SNPs into the panel, specify high priority SNPs, and use multiple design iterations for increased multiplexing efficiency. The enhanced exporting functionality generates output files for easy oligo ordering.	Image: And the second secon



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Oligo order excel file

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3 rs2_W1	rs2_W1_F	ACGTTGGATGGCTGGAAAGTTTCAAACTCG	rs2_W1_R	ACGTTGGATGCTCCTTCTATCACCACAATG	rs2_W1_E	ACGTCACCGACAAGT		
4 rs3_W1	rs3_W1_F	ACGTTGGATGTAACGCCTCATAGATCCCAG	rs3_W1_R	ACGTTGGATGGGTTGATTTGGTTACCGGTG	rs3_W1_E	CGAACCGCCGATCGGTA		
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Distinct advantages Of MassARRAY for Nucleic Acid Analysis

1. Don't use fluorescence

- Mass of the actual bioanalyte is detected 4 decimal place accuracy
- No non-specific background issues background is a different mass

2. System is quantitative

- Many biological phenomena need to be accurately quantified
- Allele ratios, gene copy number, methylation

3. Multiplexed assays

- Provides high throughput
- Cost effectiveness

4. Very sensitive and flexible

- High accuracy published 99.7%
- High genotyping call rates (+98%)
- Numbers of samples and markers are easily scaled
- Simple and flexible assay design with little optimization required
- Comprehensive Genetic Analysis >> Somatic mutation, Rare mutation, SNP, Epigenetic



Application of MassARRAY[®] in Agricultural Genomics



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Applications in Agricultural Genomics

- Parentage verification for plant breeding
- Crop strain validation
- Marker assisted breeding
- Candidate genetic marker evaluation
- Genetic Mapping and QTL analysis
- Phenotype selection
- SNP validation
- Etc.





MassARRAY- SNPs detection in Oil palm

Ting et al. BMC Genomics (2016) 17:289 DOI 10.1186/s12864-016-2607-4

BMC Genomics

RESEARCH ARTICLE





Fine-mapping and cross-validation of QTLs linked to fatty acid composition in multiple independent interspecific crosses of oil palm

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Candidate SNP markers (designated SNPE) flanking various genes associated with FA and oil biosynthesis were mined from the P5 genome build. The oil palm SNP assay design and genotyping were performed by a service provider, Agena Bioscience, Inc. (San Deigo, California) using the iPLEX [®] biochemistry on MassArray® system [23]. A custom two-multiplexed genotyping assay was designed and optimized for a panel of 40 SNPs using the Assay Design Suite 1.0 software (Agena Bioscience, Inc. San Deigo, California)





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MassARRAY- SNPs detection in Oil palm



HHS Public Access

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The oil palm Shell gene controls oil yield and encodes a homologue of SEEDSTICK

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These 80 SNP markers (designated as SNPE) were genotyped in the T128 selfed population using the Sequenom MassArray® iPlex platform.





Rice Genotyping - Cornell Rice Panel



Accessions

Donor parent

- O. rufipogon IRGC 105491
- Aus-like ancestor
- No agronomic traits of interest Crosses well with both indica and



in Beaumont, Texas

- **Recurrent parent** • O. sativa - Jefferson
- Tropical japonica
- High-yield
- Semi-dwarf plant type
- High level of disease resistance
- Early maturity

Fig 1. Parent accessions: O. rufipogon inflorescences (a) and seed (b), Jefferson inflorescences (c) and seed (d)

Imai, I., McCouch, S. R., and McClung, A. M. (2011) Plant and Animal Genomics, San Diego, California.

- Collaborative project among IRRI, JIRCAS, U. of Calcutta, U. of Aberbeen, Arkansas, Duke
- 18,760 SNPs discovered on Affymetrix 44K rice chip
- 84 SNPs for higher resolutions of key traits for MassARRAY panel
- Currently designing MassARRAY panels arranged by chromosome





8 SNP Marker Analysis for Quality and Agronomic Traits (rice)

Loci	Functions	Mutation and traits
sd-1	gibberellin 20-oxidase	C/T mutation reduces plant height and increases yield
sd-del		380-383bp deletion increases yield
Pi-ta	928-aa polypeptide	G: blast resistant; T: susceptible
waxyIN1	granule-bound starch	Starch quality. A: low amylose; G: high amylose
waxyEX 6	synthase	Startch quality. A: low amylose; T: high amylose
alk3	Starch synthase	Cooking and eating quality.
alk4		alk3 'G'+alk4"GC": high gelatinization temperature and low alkali spreading
fgr	BADH2	8bp deletion results in fragrant rice

Conclusion: MassARRAY is a powerful tool for SNP detection and confirmation in rice and can be applied to segregating populations in breeding programs for traits selection.

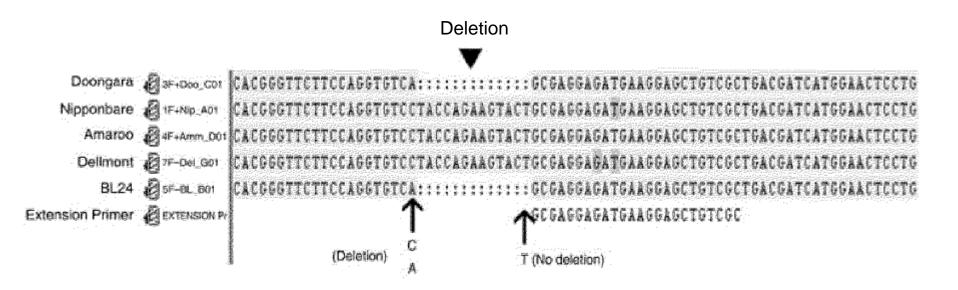
Masouleh, A. K., et al. Plant Biotech J. 2009. 7:355-63





Rice SNP Marker Analysis: Ability to Detect Deletions

MassARRAY iPLEX Gold assays are capable to detect large or small deletions.



Masouleh, A. K., et al. Plant Biotech J. 2009. 7:355-63



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SNP Identification and Panel Development for *BADH1* Haplotype Association Study

Haplotypes of the BADH1 Gene in 92 Diverse Rice Varieties

Variety	S-1	S-2	S-3	S-4	S-5	S-6	S-8	S-9	S-11	S-14	S-15	S-16	S-17	S-18	S-19	SNP Haplotype	Protein Haplotype	Frequency
Jaya	G	С	G	Т	Т	Α	Α	С	С	Т	Т	Т	Т	Α	Т	SH1	PH1	38
ADT43	Α	С	G	Т	Т	Α	Α	G	С	Т	Т	С	Т	Α	Т	SH2	PH1	19
Basmati 370	G	С	Α	Α	С	Т	G	G	Т	Т	С	С	С	С	Т	SH3	PH2	17
Taraori Basmati	G	Α	Α	Α	Т	Т	G	G	Т	Т	С	С	С	С	Т	SH4	PH2	6
Kalanamak 3119	G	С	Α	Т	Т	Α	Α	G	С	Α	Т	С	С	Α	С	SH5	PH3	2
Taipai 309	G	С	Α	Α	Т	Т	G	G	Т	Т	С	С	С	С	Т	SH6	PH2	1
Jyothi	G	С	G	Т	С	Α	G	С	С	Т	Т	Т	Т	Α	Т	SH7	PH1	1
Pusa 44	G	С	G	Т	Т	А	Α	G	С	Т	Т	Т	Т	Α	Т	SH8	PH1	1
SKR 126	G	С	G	Т	Т	А	Α	G	С	Т	Т	С	Т	Α	Т	SH9	PH1	1
CSR 10	G	С	G	Т	Т	Т	G	G	Т	Т	Т	С	Т	Α	Т	SH10	PH4	1
IR 64	G	С	G	т	Т	Α	А	С	С	Т	Т	Т	С	Α	Т	SH11	PH1	1
Pusa 1266	G	С	Α	Α	С	Т	G	С	Т	Т	С	С	С	С	Т	SH12	PH2	1
Kasturi	G	С	Α	Т	С	Т	G	G	Т	Т	С	С	С	С	Т	SH13	PH2	1
Pusa 1121	Α	С	G	т	Т	А	А	С	С	Т	Т	С	Т	Α	Т	SH14	PH1	1
Pant Dhan 4	G	С	G	Т	Т	Α	Α	С	С	Т	Т	Т	т	С	Т	SH15	PH5	1

Conclusion: There is no association between salt tolerance and the *BADH1* haplotypes

Singh, A. et al. 2010 Mol. Breeding 26,(2): 325-338,



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Plant Biotechnology Journal (2009) 7, pp. 355-363

A high-throughput assay for rapid and simultaneous analysis of perfect markers for important quality and agronomic traits in rice using multiplexed MALDI-TOF mass spectrometry

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Received 19 December 2008; revised 3 February 2009; accepted 4 February 2009 * Correspondence (fax +61 266222080; e-mail robert.henry@scu.edu.au)

Keywords: matrix-assisted laser desorption ionization-time of flight (MALDI-TOF), multiplex perfect markers, rice, single nucleotide polymorphism.

Summary

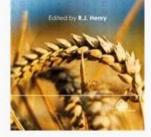
The application of single nucleotide polymorphisms (SNPs) in plant breeding involves the analysis of a large number of samples, and therefore requires rapid, inexpensive and highly automated multiplex methods to genotype the sequence variants. We have optimized a high-throughput multiplexed SNP assay for eight polymorphisms which explain two agronomic and three grain quality traits in rice. Gene fragments coding for the agronomic traits plant height (semi-dwarf, sd-1) and blast disease resistance (Pi-ta) and the quality traits amylose content (waxy), gelatinization temperature (alk) and fragrance (fgr) were amplified in a multiplex polymerase chain reaction. A single base extension reaction carried out at the polymorphism responsible for each of these phenotypes within these genes generated extension products which were quantified by a matrix-assisted laser desorption ionizationtime of flight system. The assay detects both SNPs and indels and is co-dominant, simultaneously detecting both homozygous and heterozygous samples in a multiplex system. This assay analyses eight functional polymorphisms in one 5 µL reaction, demonstrating the high-throughput and cost-effective capability of this system. At this conservative level of multiplexing, 3072 assays can be performed in a single 384-well microtitre plate, allowing the rapid production of valuable information for selection in rice breeding.



CABI Book Info

Plant genotyping II: SNP technology

Plant Genotyping II SNP Technology



Description

This book aims to describe some of the important recent developments in plant genotyping. It is based upon a second workshop held recently to review progress in this area. Recent developments focus on high- throughput methods and generally target single nucleotide polymorphism (SNP) discovery and analysis. The topics covered include: SNP discovery in plants; SNPs and their use in maize; rare SNP discovery with endonucleases; sequence polymorphisms in the flanking regions of microsatellite markers; SNP discovery by ecotilling using capillary electrophoresis; genotyping by allele-specific PCR; the MassARRAY system for plant genomics; mutation screening; nanotechnology (the future of cost-effective plant genotyping); functionally associated molecular genetic markers for temperate pasture plant improvement; genotyping for rice eating qualities; towards universal loci for plant genotyping; DNA banks as a resource for SNP genotyping; DNA extraction from plant tissue; future prospects for plant genotyping.

Available In Print 🗲

Book details

Editor(s) Henry, R. J.

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Centre for Plant Conservation Genetics Southern Cross University Lismore, New South Wales, Australia.

Year of Publication 2008

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Descriptor(s) alleles crop quality enzyme activity enzymes genetic markers genetic polymorphism genetic variation genotypes



Book Chapters

Chapter: 1 (Page no: 1)	SNP discovery in plants. Author(s): Edward, K. J. Poole, R. L. Barker, G. L.
Chapter: 2 (Page no: 30)	SNPs and their use in maize. Author(s): Rafalski, A. Tingey, S.
Chapter: 3 (Page no: 44)	Rare SNP discovery with endonucleases. Author(s): Cross, M. J.
Chapter: 4 (Page no: 68)	Sequence polymorphisms in the flanking regions of microsatellite markers. Author(s): Ablett, G. Henry, R. J.
Chapter: 5 (Page no: 78)	Snp discovery by ecotilling using capillary electrophoresis. Author(s): Eliott, F. Cordeiro, G. Bundock, P. C. Henry, R. J.
Chapter: 6 (Page no: 88)	Genotyping by allele-specific PCR. Author(s): Waters, D. L. E. Bundock, P. C. Henry, R. J.
Chapter: 7 (Page no: 98)	The MassARRAY system for plant genomics. Author(s): Irwin, D.
Chapter: 8 (Page no: 114)	Mutation screening. Author(s): Izquierdo, L.
Chapter: 9 (Page no: 133)	Nanotechnology: the future of cost-effective plant genotyping. Author(s): Pattemore, J. A. Trau, M. Henry, R. J.
Chapter: 10 (Page no: 154)	Functionally associated molecular genetic markers for temperate pasture plant improvement. Author(s): Forster, J. W. Cogan, N. O. I. Dobrowolski, M. P. Francki, M. G. Spangenberg, G. C. Smith, K. F.
Chapter: 11 (Page no: 187)	Genotyping for rice eating qualities. Author(s): Bradbury, L. M. T. Waters, D. L. E. Henry, R. J.
Chapter: 12 (Page no: 195)	Towards universal loci for plant genotyping. Author(s): Pacey-Miller, T.

polymerase chain reaction reviews rice single nucleotide polymorphism

Subject Code(s)

FF005 - Field Crops. (New March 2000) FF020 - Plant Breeding and Genetics ZZ360 - Molecular Biology and Molecular Genetics. (Discontinued March 2000, Reinstated and Revised June 2002)

Record Number 20083134963



Livestock Genotyping



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Assays by Agena: Bovine Panels

- U.S. Department of Agriculture (USDA) Markers selected from HapMap project
- Genome wide SNPs with high MAF >0.41 selected for discrimination of inbred diary herds
- 2 well panels for parentage verification with 100-121 SNPs per International Society for Animal Genetics (ISAG) recommendations
- Additional SNPs/wells for disease susceptibility and trait identification





Pig Identification and Parentage Exclusion



Rohrer, G. A. et al. Animal Genetics. 2007. 38 (3): 253–258

Comparison of 60 SNPs & 10 microsatellite markers

- Designed panel with 80 SNPs
 - 155 boars that represent four US purebred populations tested representative of US purebred Duroc, Hampshire, Landrace and Yorkshire populations
 - 60 SNPs w/ MAF >0.15 selected for the final panel of 60 markers
- Parentage Exclusion Probability
 - One parent: 0.9974 (all data)
 - Ranged from 0.9594 to 0.9963 within breeds.
- Sire Exclusion Probability
 - When dam's genotype is known: 0.99998 (all data)
 - Ranged from 0.99868 to 0.99997 within breeds

Parental exclusion probabilities for SNP and microsatellite marker panels were similar, but genotyping was more sensitive for individual identification.



ANIMAL GENETICS Immunogenetics, Molecular Genetics and Functional Genomics



doi:10.1111/j.1365-2052.2009.01903.x

Relationships among *calpastatin* single nucleotide polymorphisms, *calpastatin* expression and tenderness in pork longissimus¹

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Summary

Genome scans in the pig have identified a region on chromosome 2 (SSC2) associated with tenderness. Calpastatin is a likely positional candidate gene in this region because of its inhibitory role in the calpain system that is involved in postmortem tenderization. Novel single nucleotide polymorphisms (SNP) in calpastatin were identified and used to genotype a population (n = 1042) of Duroc-Landrace-Yorkshire swine for association with longissimus lumborum slice shear force (SSF) measured at days 7 and 14 postmortem. Three genetic markers residing in the calpastatin gene were significantly associated with SSF (P < 0.0005). Haplotypes constructed from markers in the *calpastatin* gene were significantly associated with SSF (F-ratio = 3.93; P-value = 0.002). The levels of normalized mRNA expression of calpastatin in the longissimus lumborum of 162 animals also were evaluated by real-time RT-PCR and were associated with the genotype of the most significant marker for SSF (P < 0.02). This evidence suggests that the causative variation alters expression of calpastatin, thus affecting tenderness. In summary, these data provide evidence of several significant, publicly available SNP markers associated with SSF that may be useful to the swine industry for marker assisted selection of animals that have more tender meat.

Keywords calpastatin, gene expression, meat quality, pig, single nucleotide polymorphism.





Predictive SNPs in Calpastatin for Pork Tenderness



Nonneman, D. et al. J. Anim. Sci. 2011: 2010-3556

- 194 SNPs in calpastatin locus on pig chr 2 were identified by re-sequencing
- SNP Genotyping on MassARRAY using 40 pigs
- 37 SNPs significantly associated with slice shear force in USMARC population
 - 4 SNPs significantly associated with tenderness, juiciness, chewiness, and other flavor scores in all 4 populations

SNP markers identified by the association study should be predictive of pork tenderness in industry populations



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MassARRAY [™] Customers List

- USDA
- CIAT
- Neogen/GeneSeek
- CaptitalBio
- Washington State University
- Kansas State University
- etc

Scientific Specialis





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Questions







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