



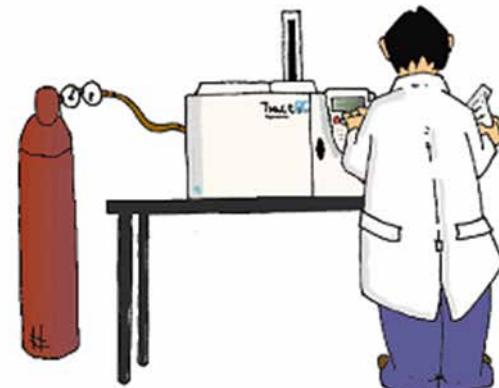
Fundamental Gas Chromatograph



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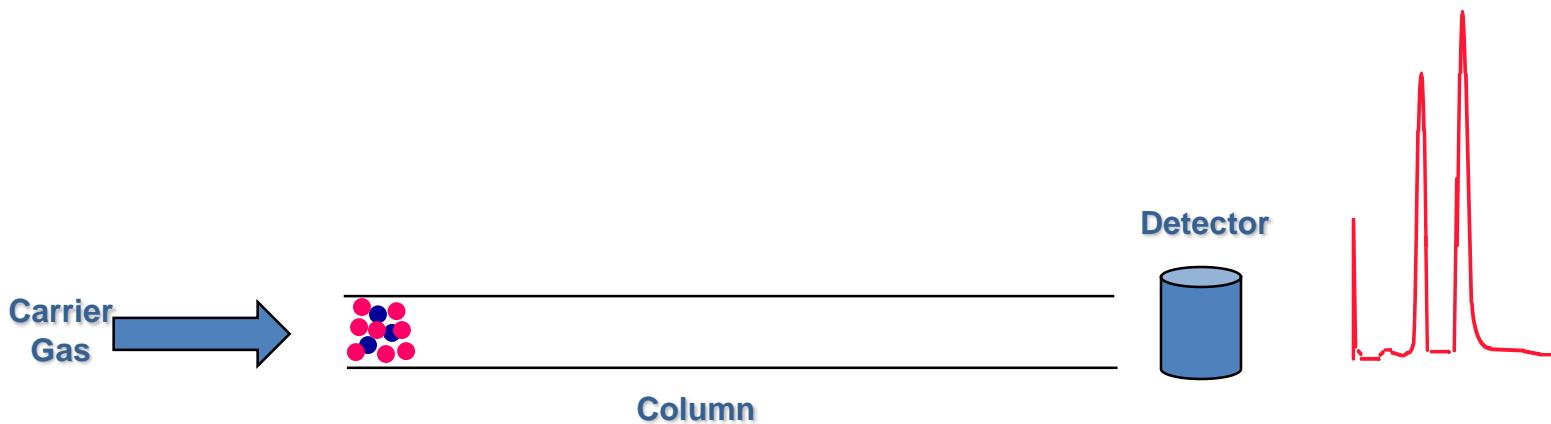
Chromatography

- Chromatography : Analytical technique that depends on separation of components in sample
- Sample components are separated and detected
- Separation : Between two phases
 - Stationary Phase
 - Mobile phase

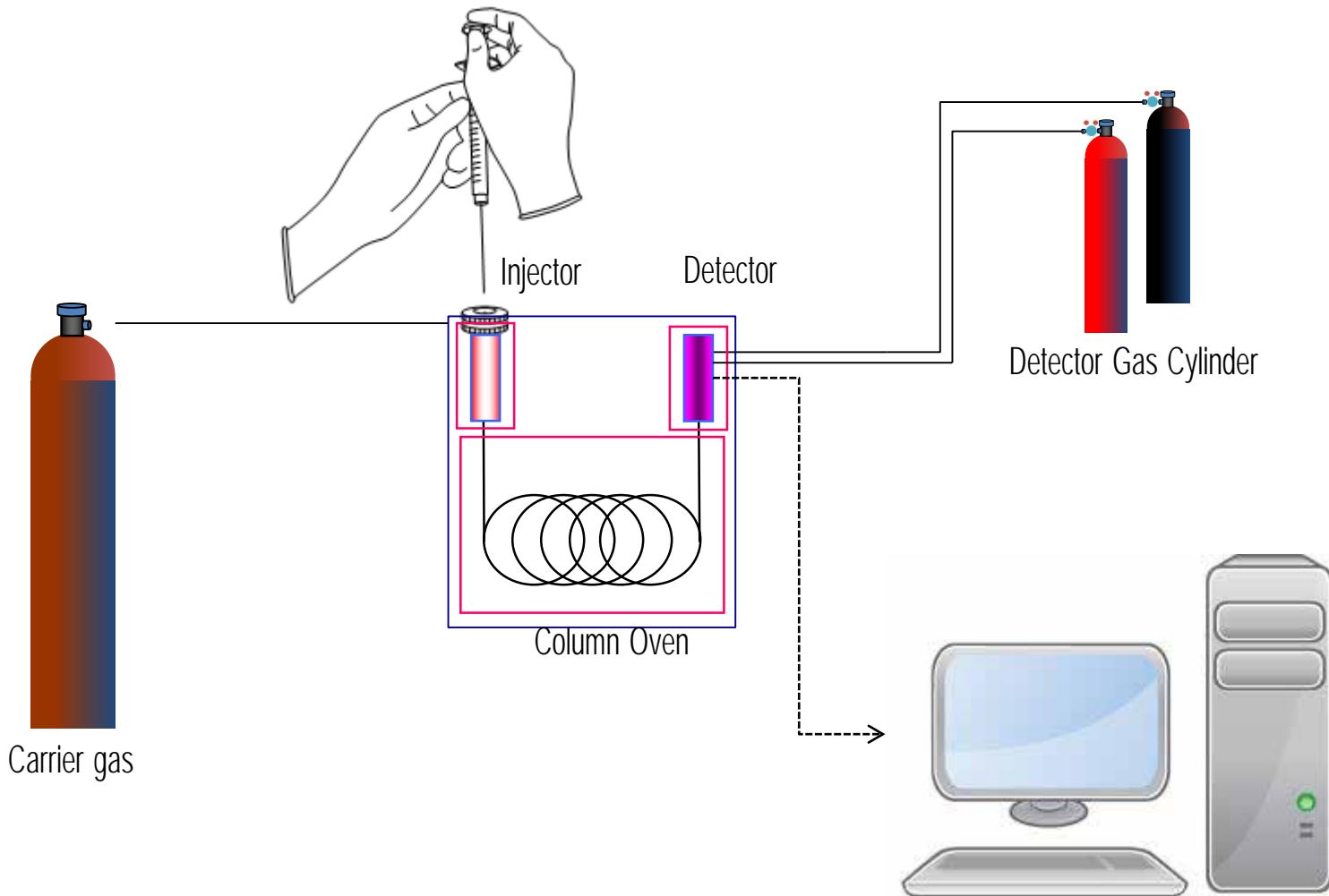


Gas Chromatography

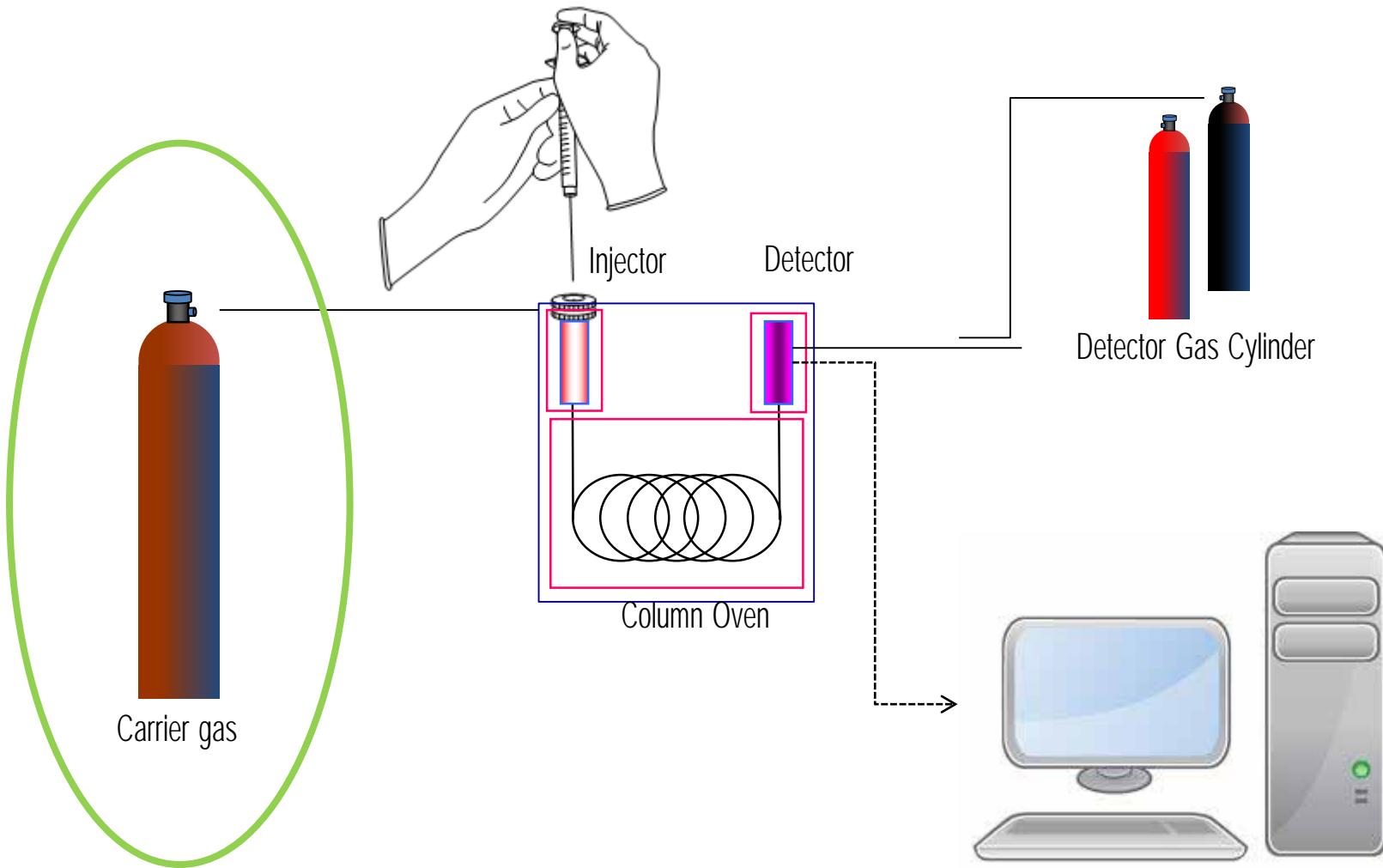
- Gas Chromatography (GC) : Chromatography technique which gas is used as mobile phase
- Sample will be injected into the system, Injection port where all components are vaporized and swept into the column
- Sample components will then be separated according to the interaction with stationary phase and eluted to detector.



GC System Components

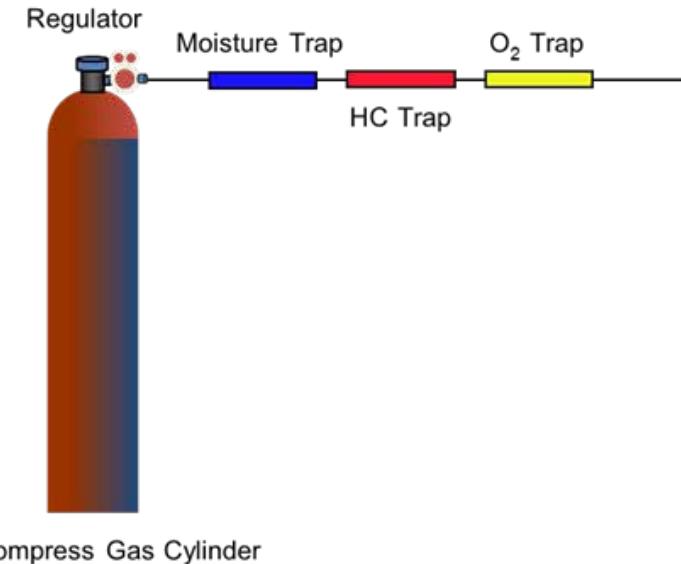


GC System Components : Carrier gas



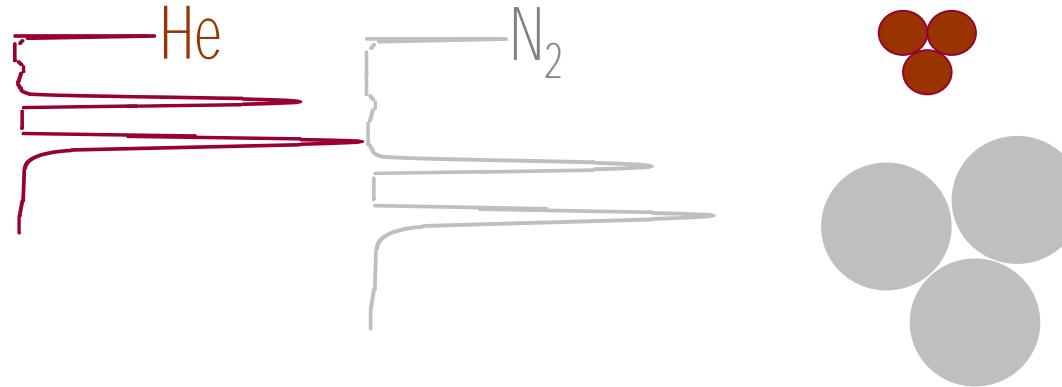
Carrier Gas Selection : Gas Purity (impurity)

- Impurities can alter stationary phase in column and cause high background (noise), contamination
 - Free from moisture, organic hydrocarbons and oxygen
 - Free from components those associate or interfere the analysis
 - Recommended at least 99.995%
 - Purified traps must be installed

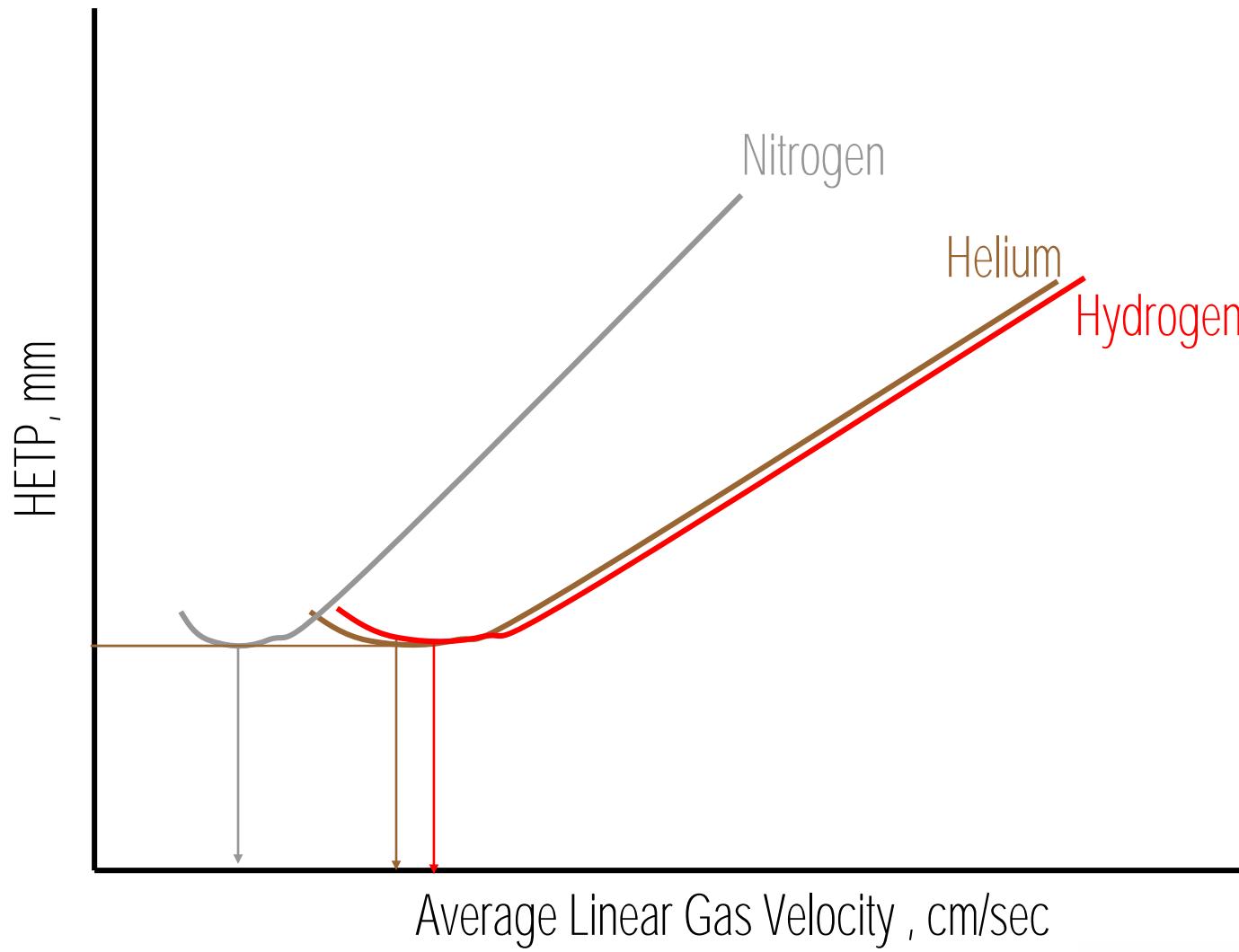


Compress Gas Cylinder

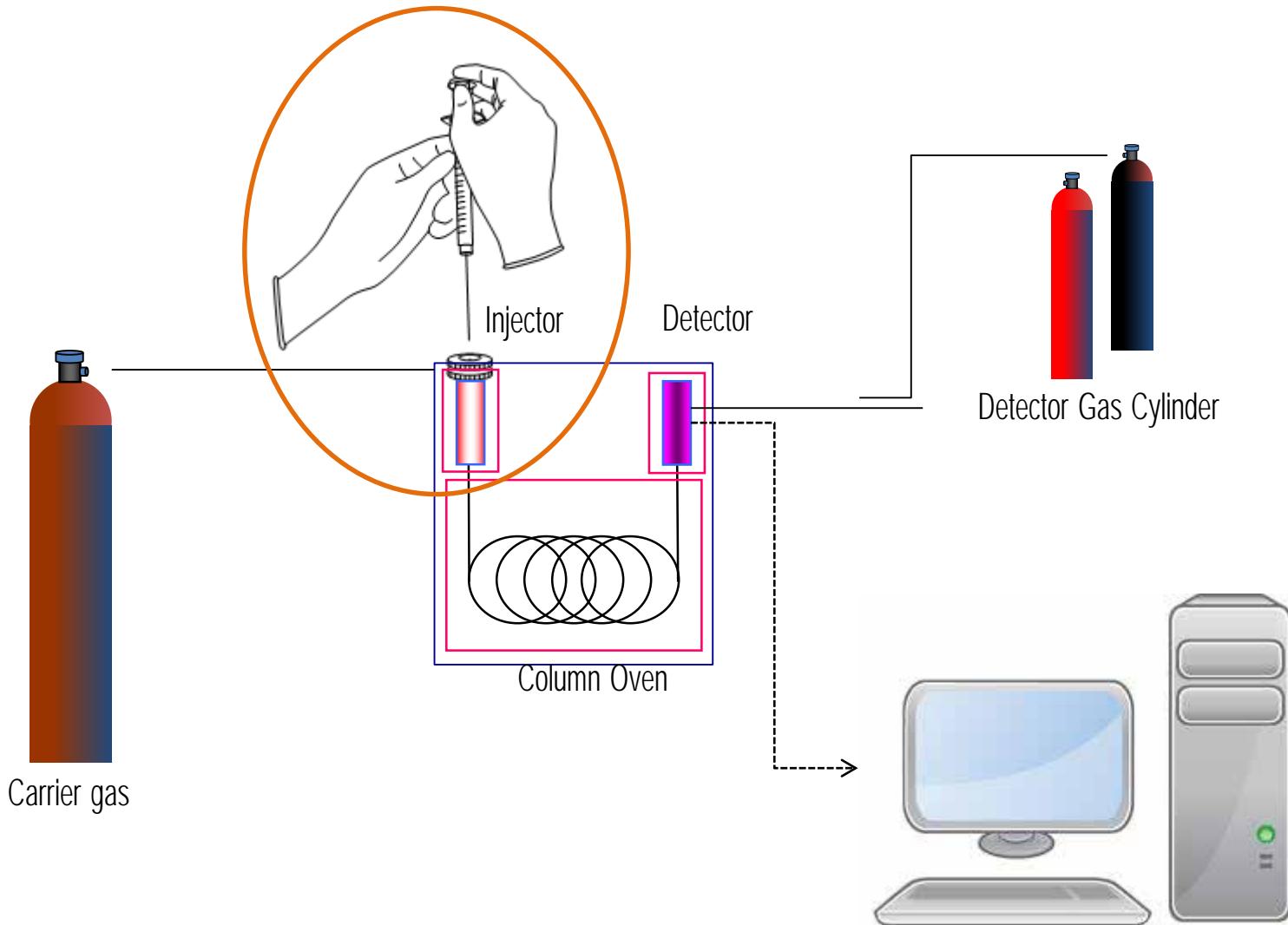
- Speed of analysis : The lighter carrier gas, the faster analysis time.
 - With the same resolution (separation performance), Helium provides shorter analysis time than Nitrogen
 - Helium is lighter than Nitrogen so it travels through column faster than Nitrogen
 - At the same supplied pressure, Helium has more density than Nitrogen so Helium will provide better peak shape (resolution).



Carrier Gas & Speed of Analysis

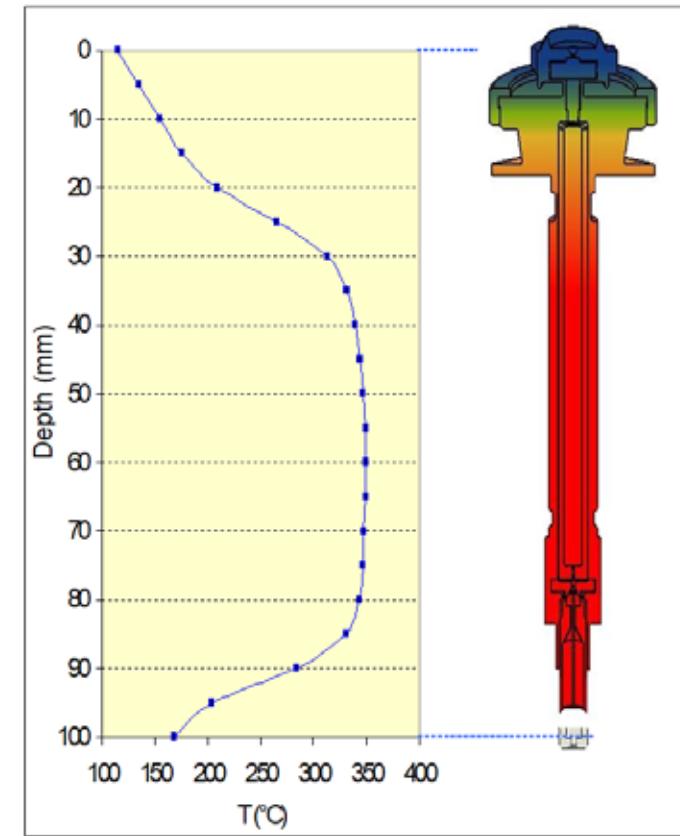


GC System Components : Injector



Injector

- Injector : The area in which the sample is introduced, evaporated instantaneously & carried to the column with a minimum of band spreading.
- Concerned parameters :
 - Sample size
 - Temperature
 - Carrier gas pressure/flow control

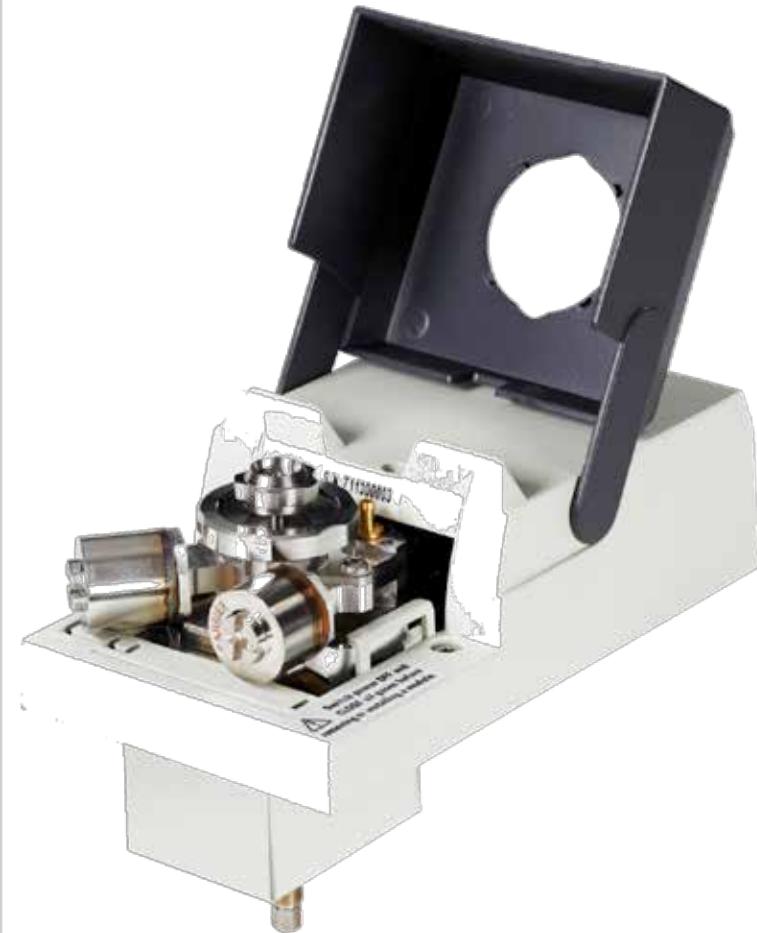


Types of Injection

- Packed Column Injector
- Split/Splitless Injector (Capillary Injector)
- On-Column Injector
 - Packed
 - Capillary
 - Cold On-Column
- PTV : Pressure Temperature Vaporizing Injector
- Injection Valve
 - Gas Sampling Valve (GSV)
 - Liquid Sampling Valve (LSV)

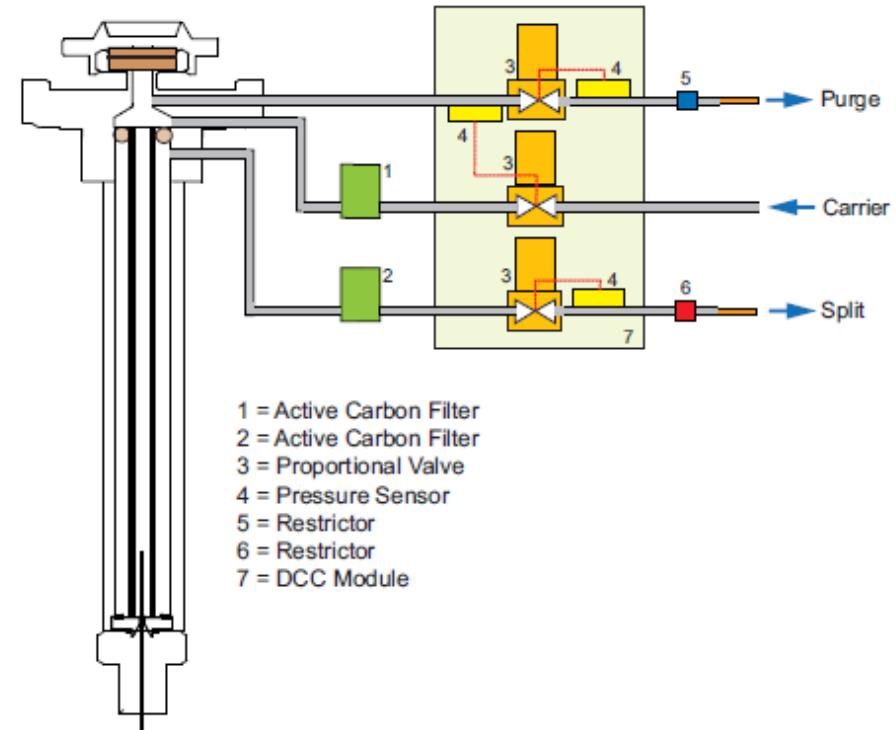
Split/Splitless Injector

- Can be used for
 - Capillary column 0.1, 0.25, 0.32 mm ID
 - Wide bore column (0.53 mm.ID)
 - Packed column (requires conversion kit)
- Can be operated in two modes
 - Split
 - Splitless



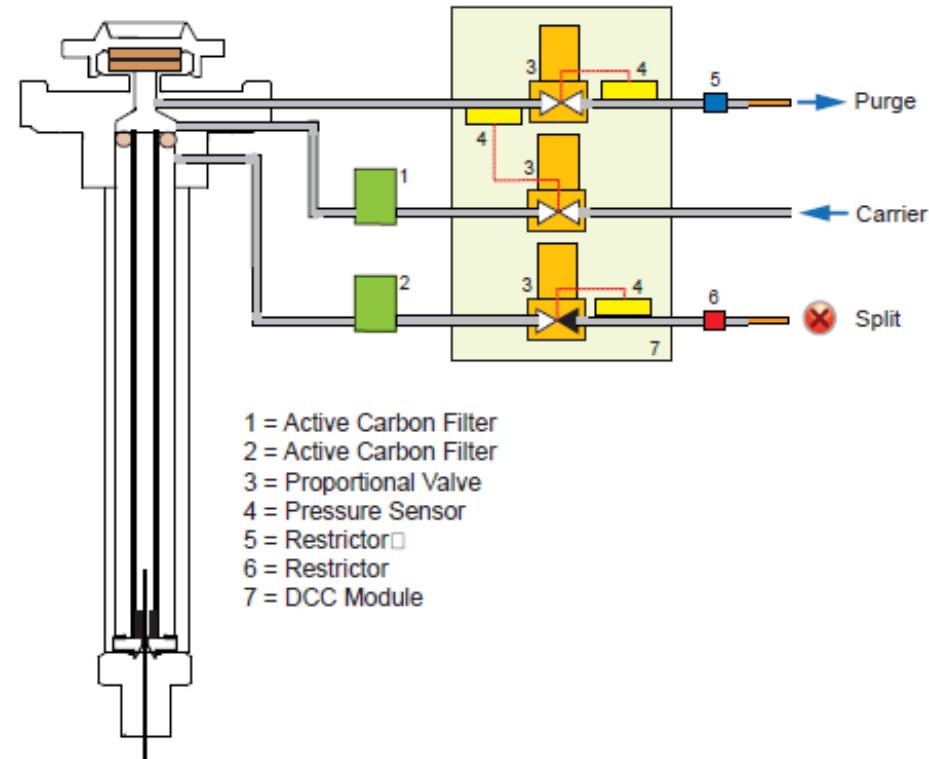
Split injection technique

- Split Injection
 - Only a part of the sample transfers into the column. The rest discharges through the split vent
 - The ratio of the split flow to the column flow so called "split ratio" determines the amount of sample that enter the column



Splitless injection technique

- Splitless injection is suitable for
 - The analysis of compounds present in very low concentration with relatively dirty matrices.
 - Allows a portion of entire sample to enter the column without splitting
 - Split vent is closed during sample injection and transfer to the column, Once the transfer is over, the split vent is reopened to flush the vaporizing chamber for any remaining sample vapors.

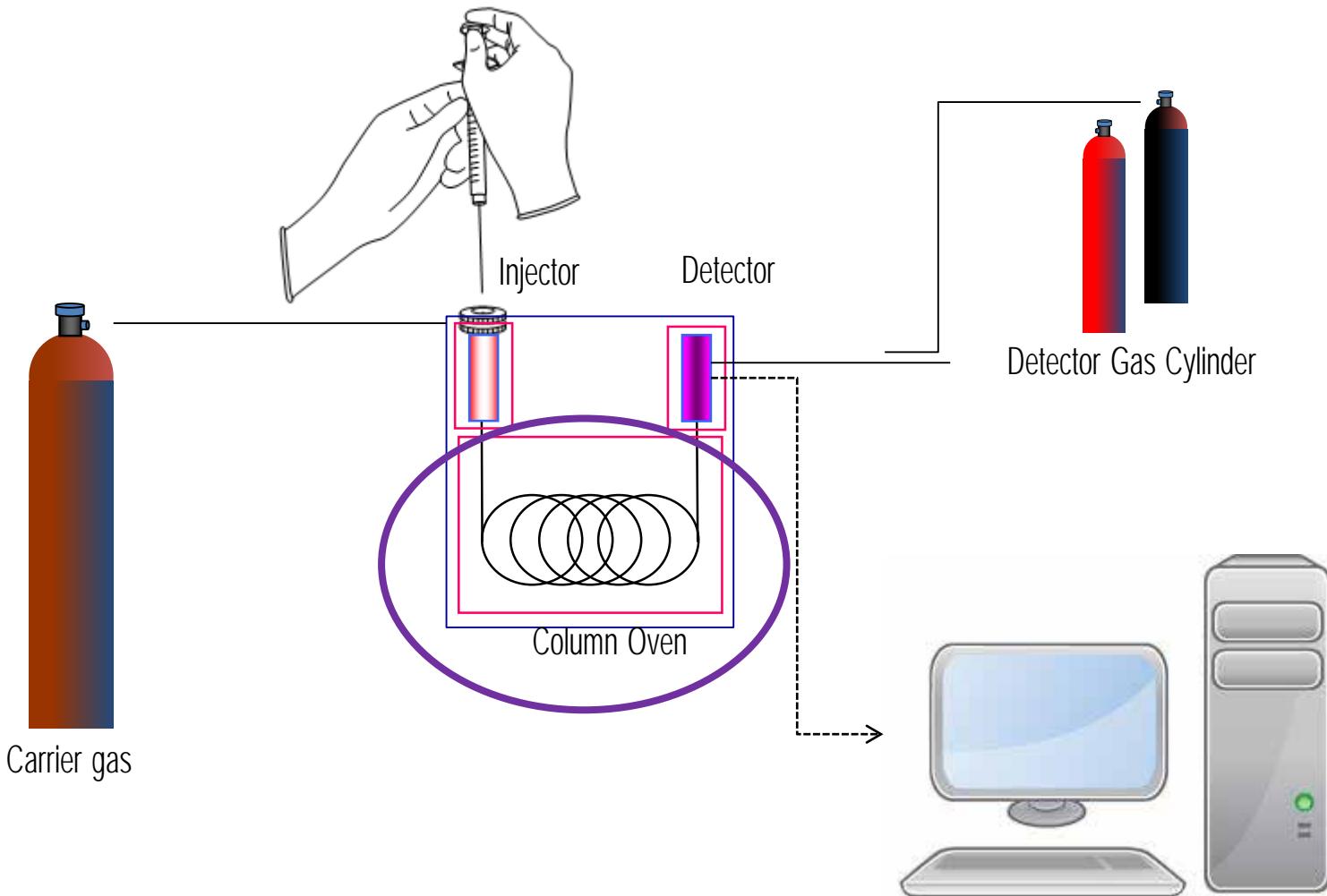


Injector : general maintenance for user

- Monitor contamination
- Set optimum injection temperature (provide complete sample vaporization)
- Inject clean sample, appropriate sample size
- Clean liner, Change liner
- Change liner seal or liner o-ring
- Change septum



GC System Components : Column and Oven



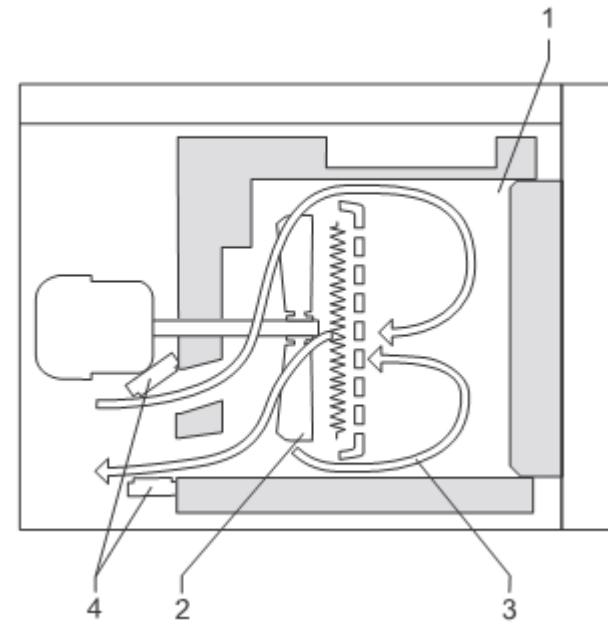
Selection of stationary phase

- The rule :
 - A non-polar component is dissolved in a non-polar liquid phases
 - A polar component is dissolved in a polar liquid phase.
- Elution Order of interested components vs. matrix
- Resolution : Separation Capability
- Temperature limitation of the stationary phase



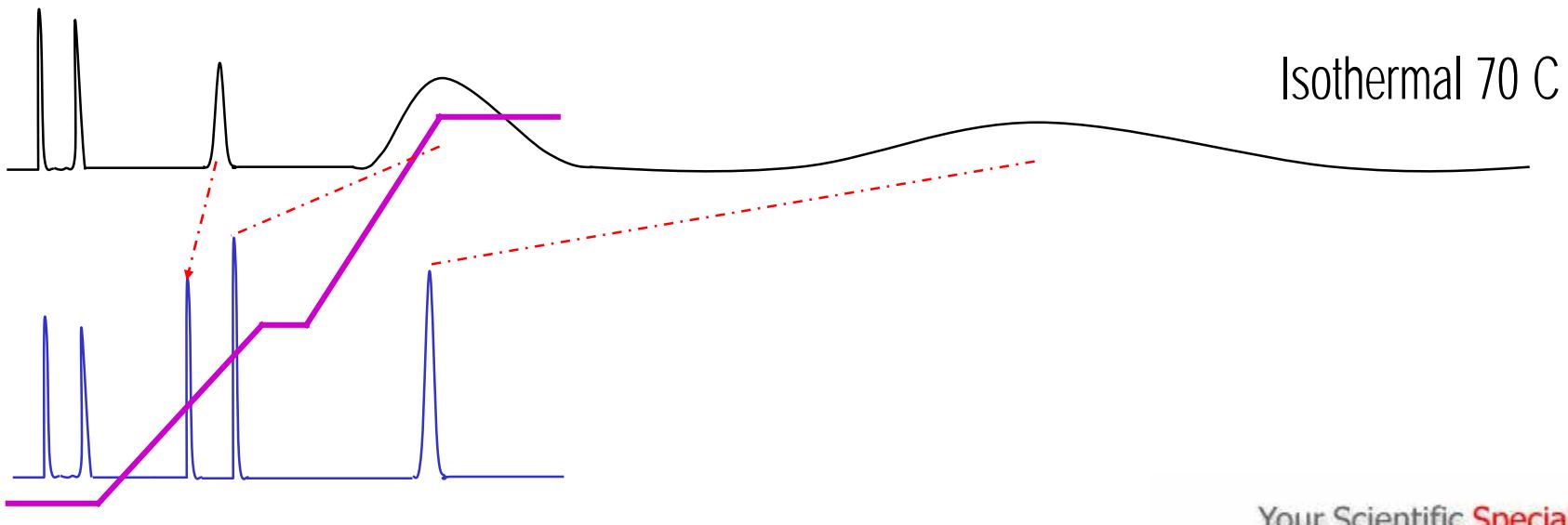
Column Oven

- Provides a stable heating environment for the analytical column.
- Must heats and cools quickly with efficient air circulation to ensures a high degree of thermal stability

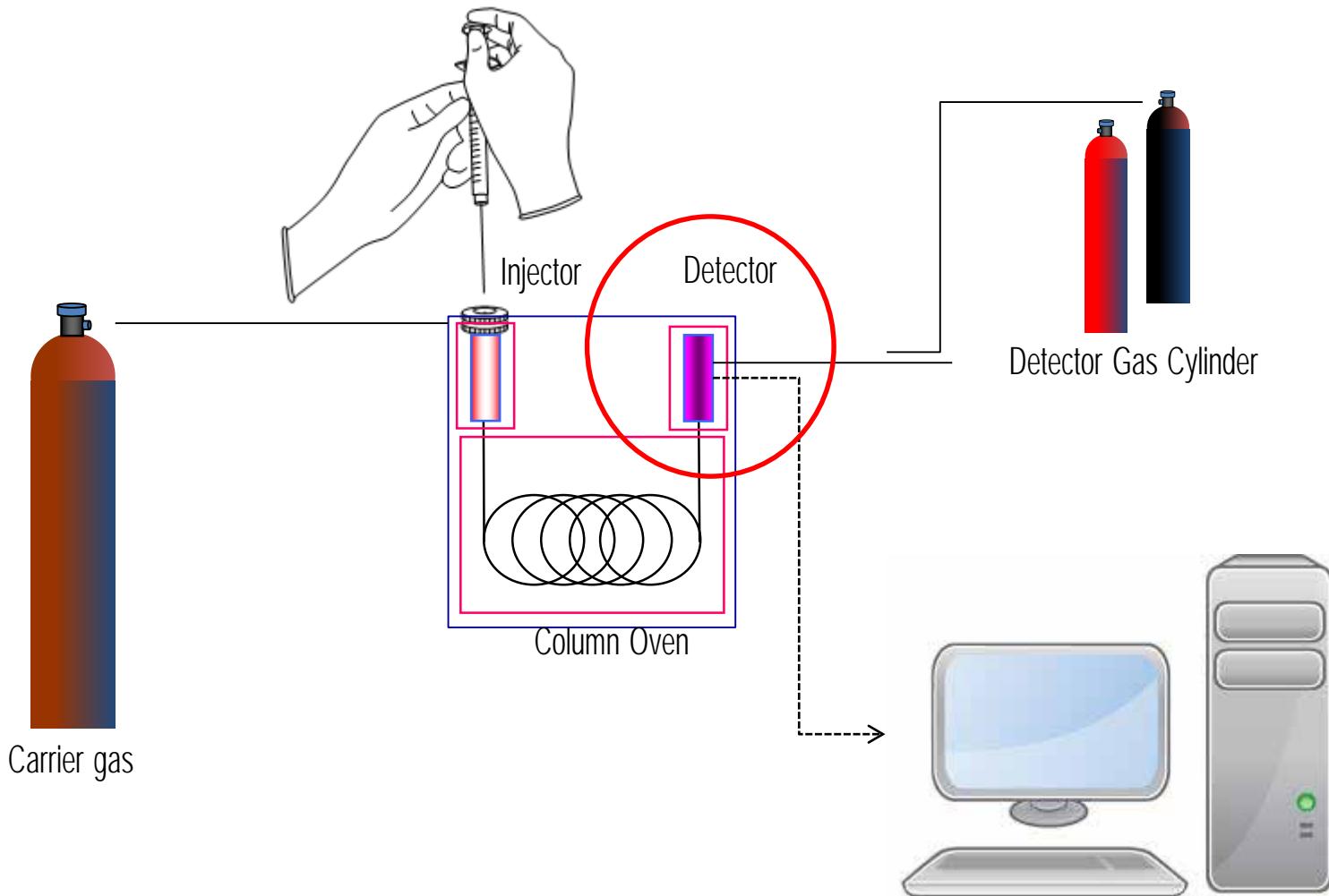


Oven Temperature vs. Resolution

- Components in the sample will be separated under optimum column temperature
- Increases oven temperature trend to reduce in resolution
- Ultimate Goal is “all components are separated with the shortest analysis time”



GC System Components : Detector



§ Miniaturized instant connect detectors

- Ø Available: FID, ECD, TCD, NPD, PDD and FPD (also dual flame)
- Ø Single bodies including cells, heater and gas feeding
- Ø Reduced volumes for increased sensitivity
- Ø Up to four can be mounted and operated at the same time
- Ø Fast acquisition speed: up to 300 Hz
- Ø Enhanced Linearity
- Ø Easy access to removable parts for maintenance



§ Front-end to Mass Spectrometers for increased selectivity and sensitivity

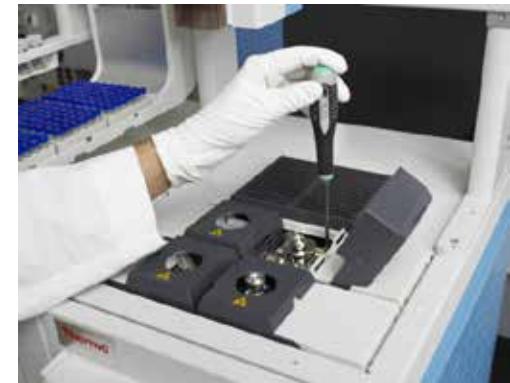
TRACE 1300 Series GC "Detector" modules



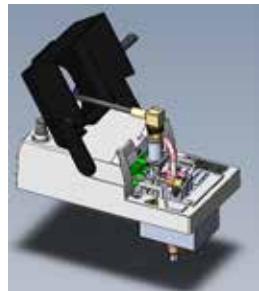
FID



TCD



PDD



NPD



ECD





Fundamental of Mass Spectrometer



Why GC/MS?

- Universal and specific
 - Full scan for unknown sample
 - SIM, MIM for specific (interested) mass
- High Sensitivity
 - ppt level
- Provides identification with standard or library spectrum
- Interference-free quantitation (SIM or MIM)
- Isotopic information
- Confirmation of other conventional detectors

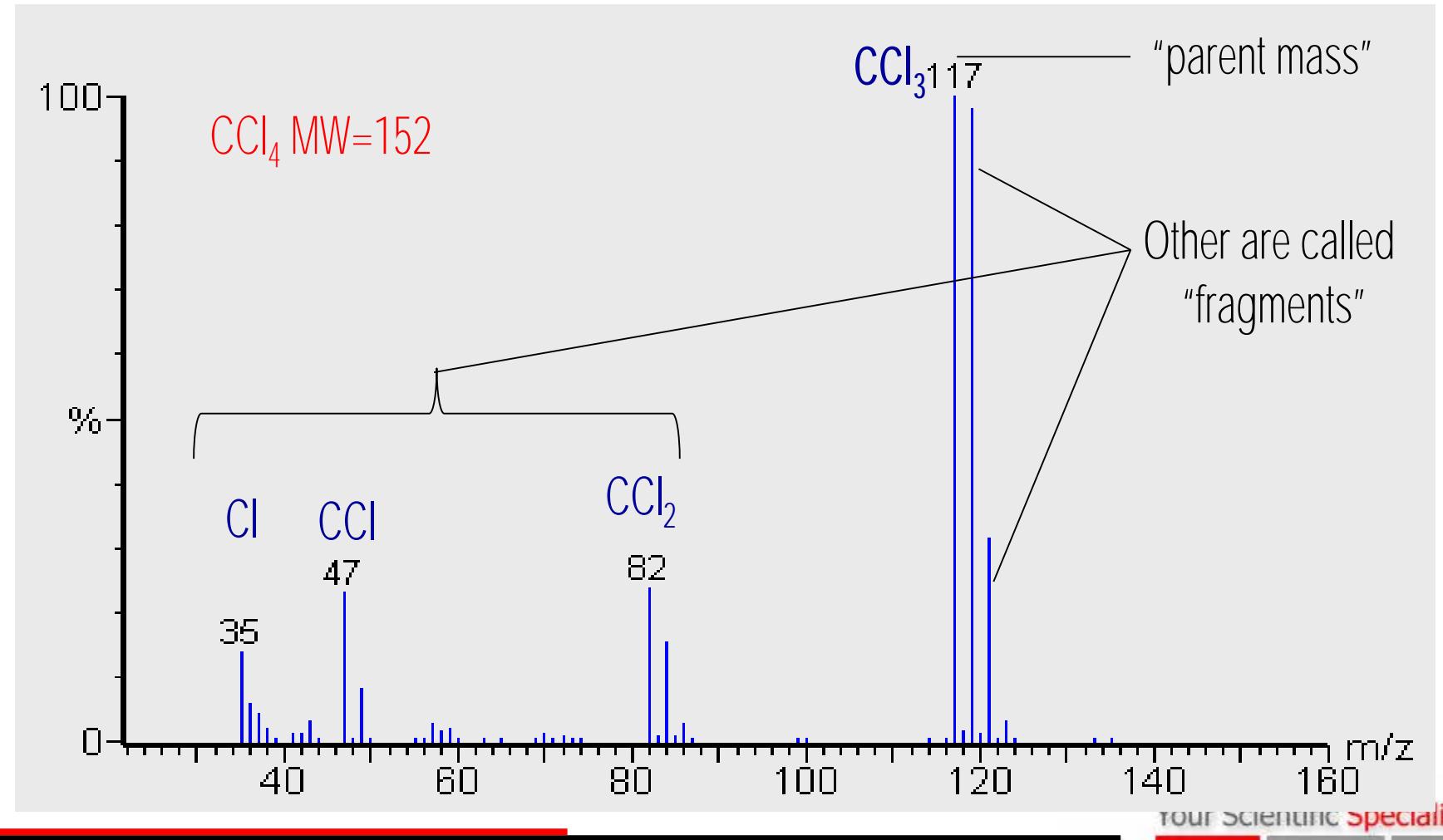


What is Mass Spectrometry?

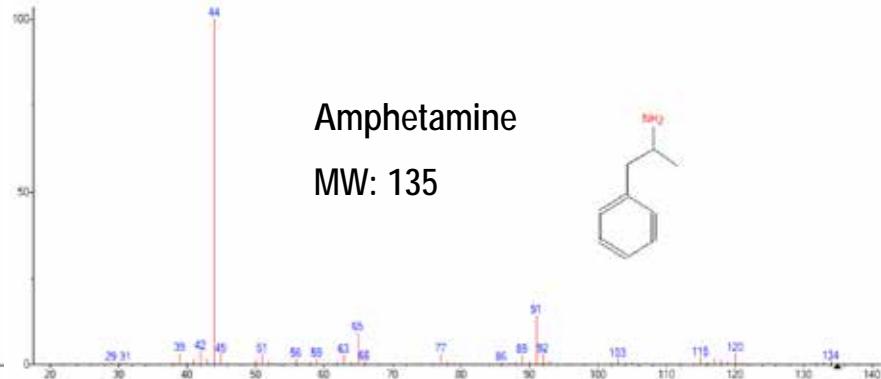
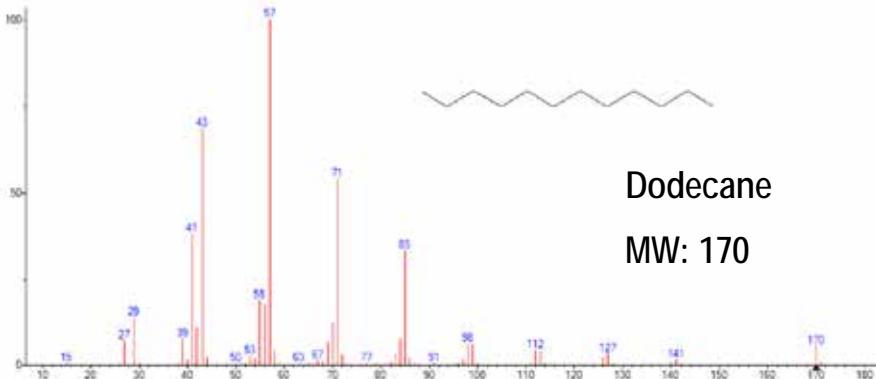
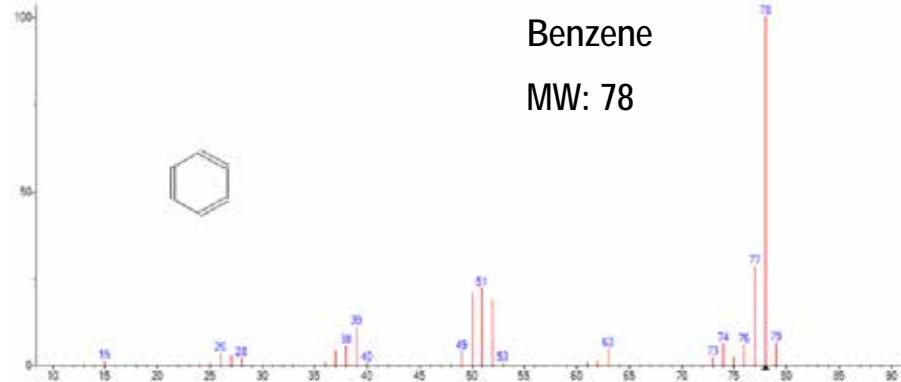
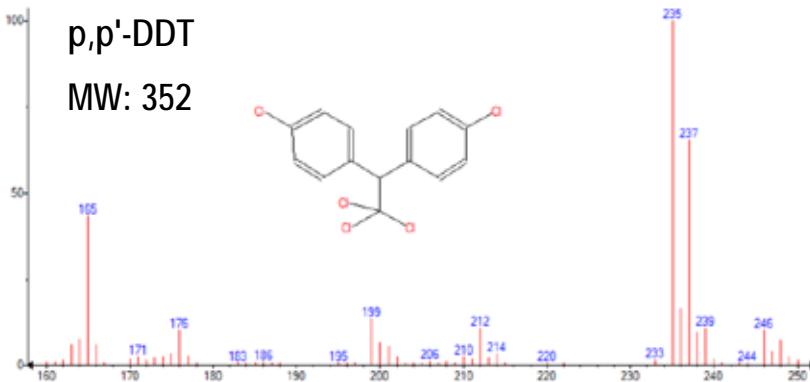
- The production of ions that are subsequently separated or filtered according to their mass-to-charge (m/z) ratio and detected.
- The resulting mass spectrum is a plot of the (relative) abundance of the produced ions as a function of the m/z ratio."

What is "Mass Spectrum" ?

- Graph of Relative Ion Intensity vs. m/z
- Ion Fragments detail structure and molecular weight of compound

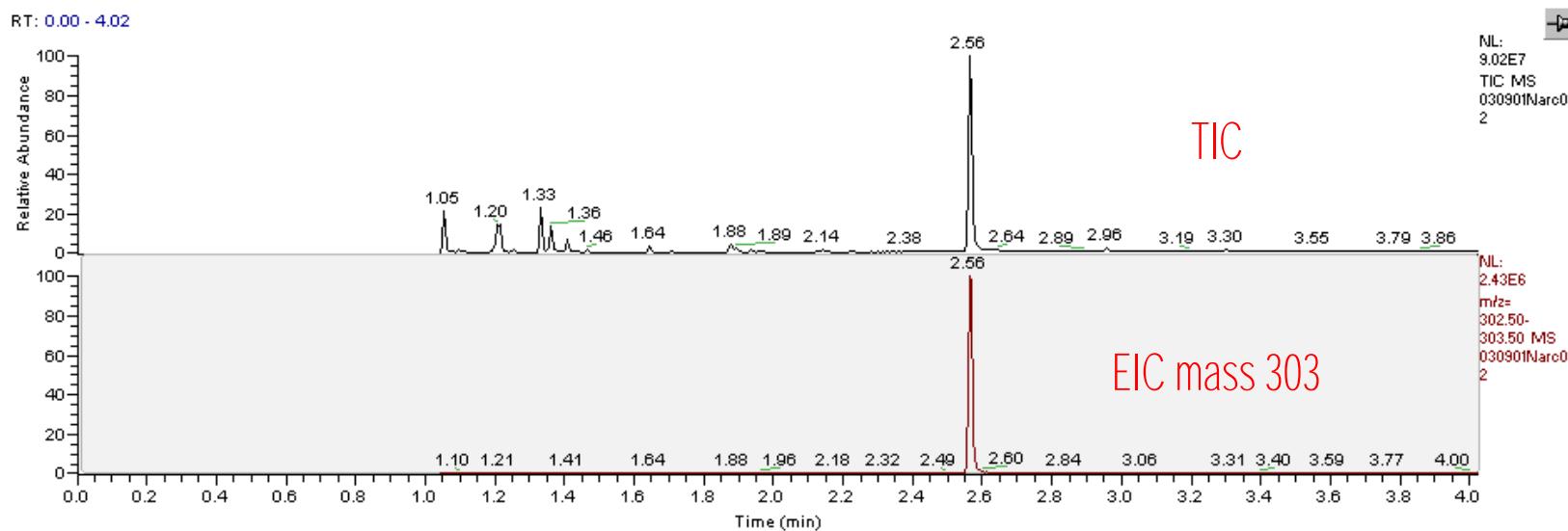


Mass Spectrum



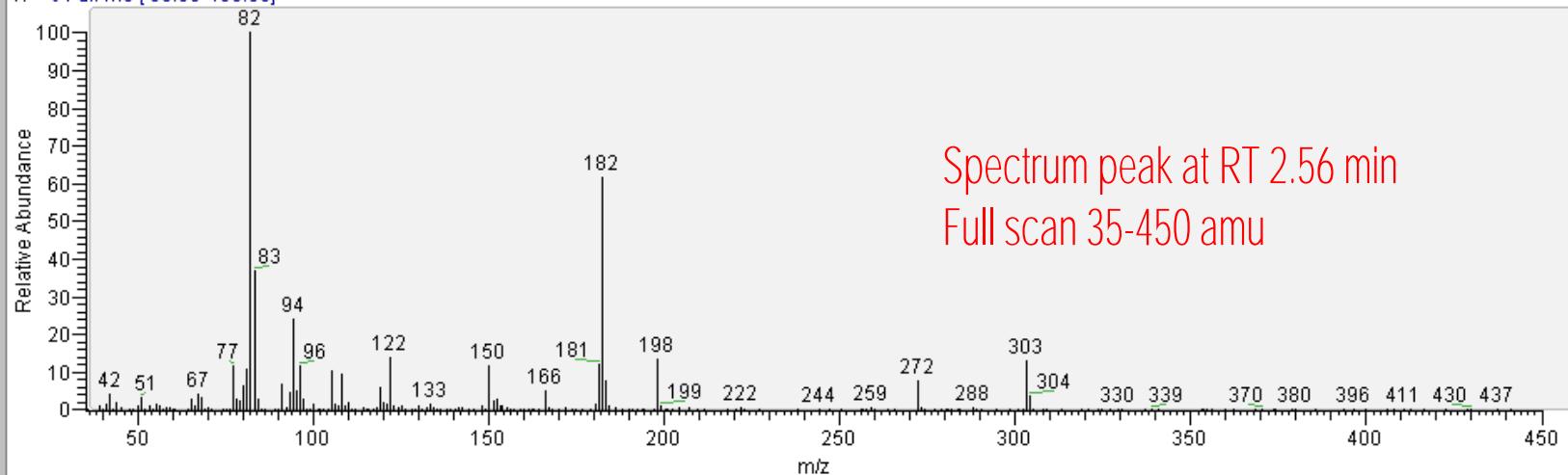
Sci Spec

Total Ion Chromatogram (TIC), Extracted Ion Chromatogram (EIC), and Mass Spectrum

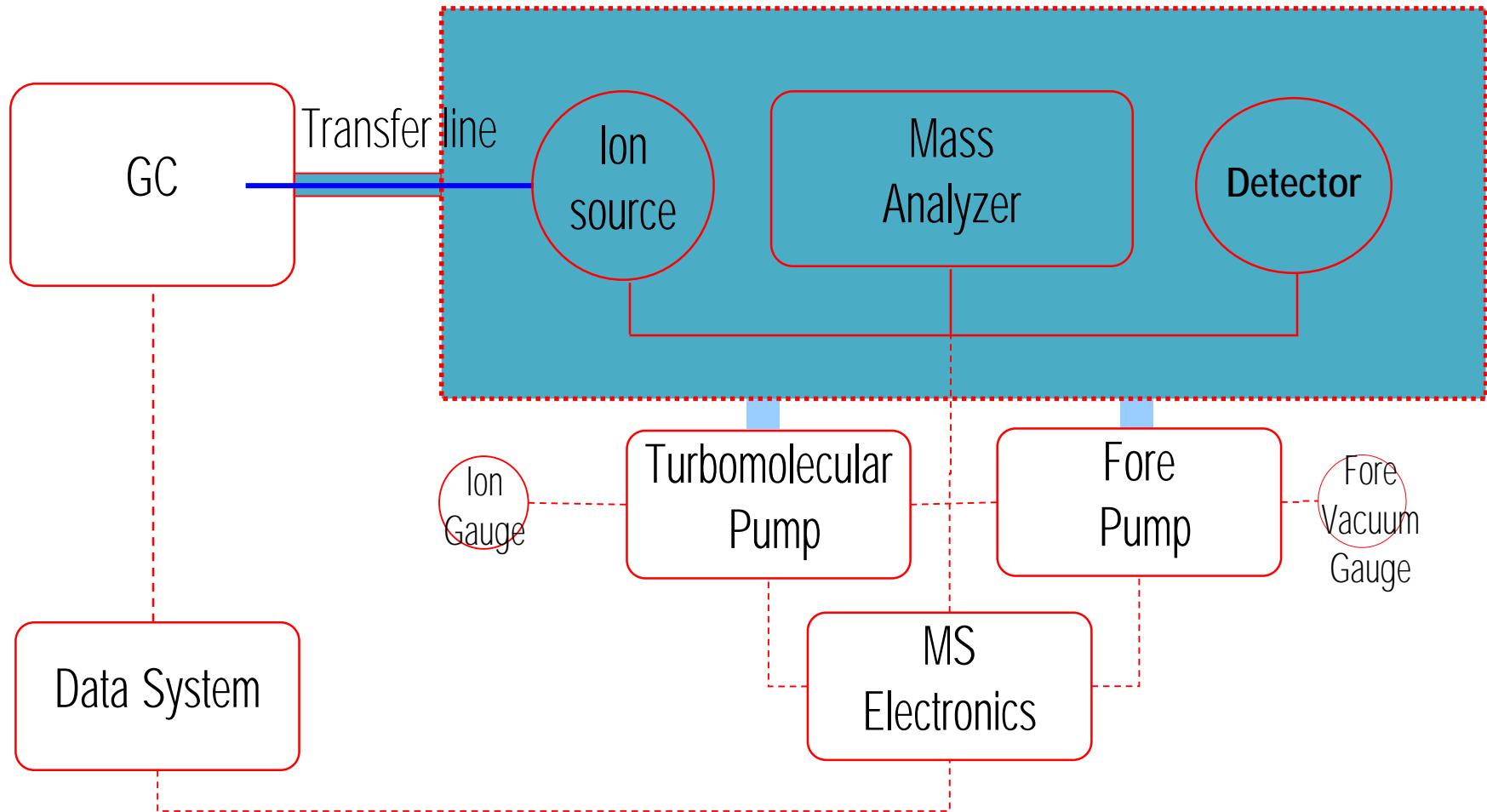


030901Narc02 #295 RT: 2.56 AV: 1 NL: 1.84E7

T: + c Full ms [35.00-450.00]

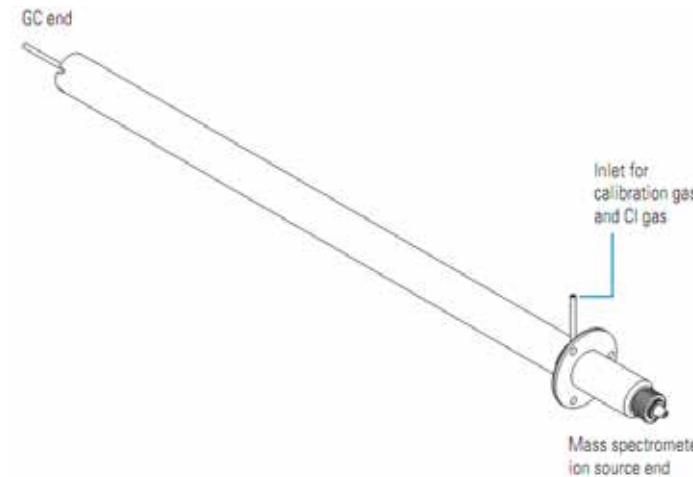


Components in GC/MS



Transfer line

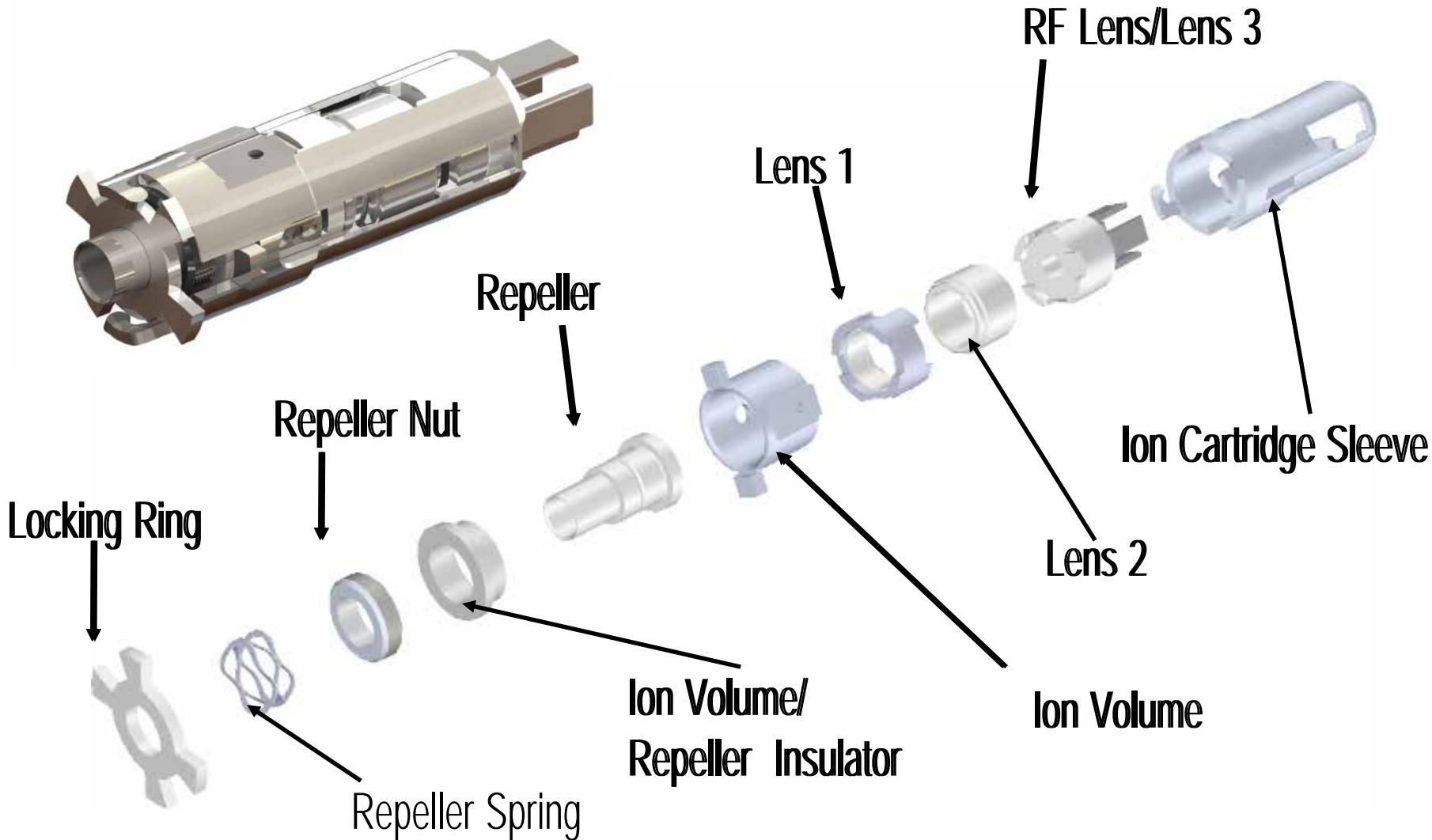
- "Bridge" between GC and MS's Ion Source
- Vacuum tube with have heater coil on the internal tube.
- GC column is inserted inside the internal tube.
- High temperature (200-350 C) is set to protect sample condensation.
- Type
 - Direct capillary transfer line (most widely used) – GC column connect directly to ion source
 - Open/Split transfer line
 - Splitter transfer line
 - Jet separator



Ion source

- Ion Source convert sample molecules (neutral) into charged molecules or molecular ions.
- Charged molecules (Molecular ions) can be easily manipulated with electrical and magnetic fields
- Process in mass spectrometer are using DC, RF to
 - Focusing : arrange the molecular ion to travel in a straight direction
 - Diverting : turn the direction of molecular ion
 - Filtering : get rid of unwanted molecular ion
 - Detecting : detect those interested molecular ion

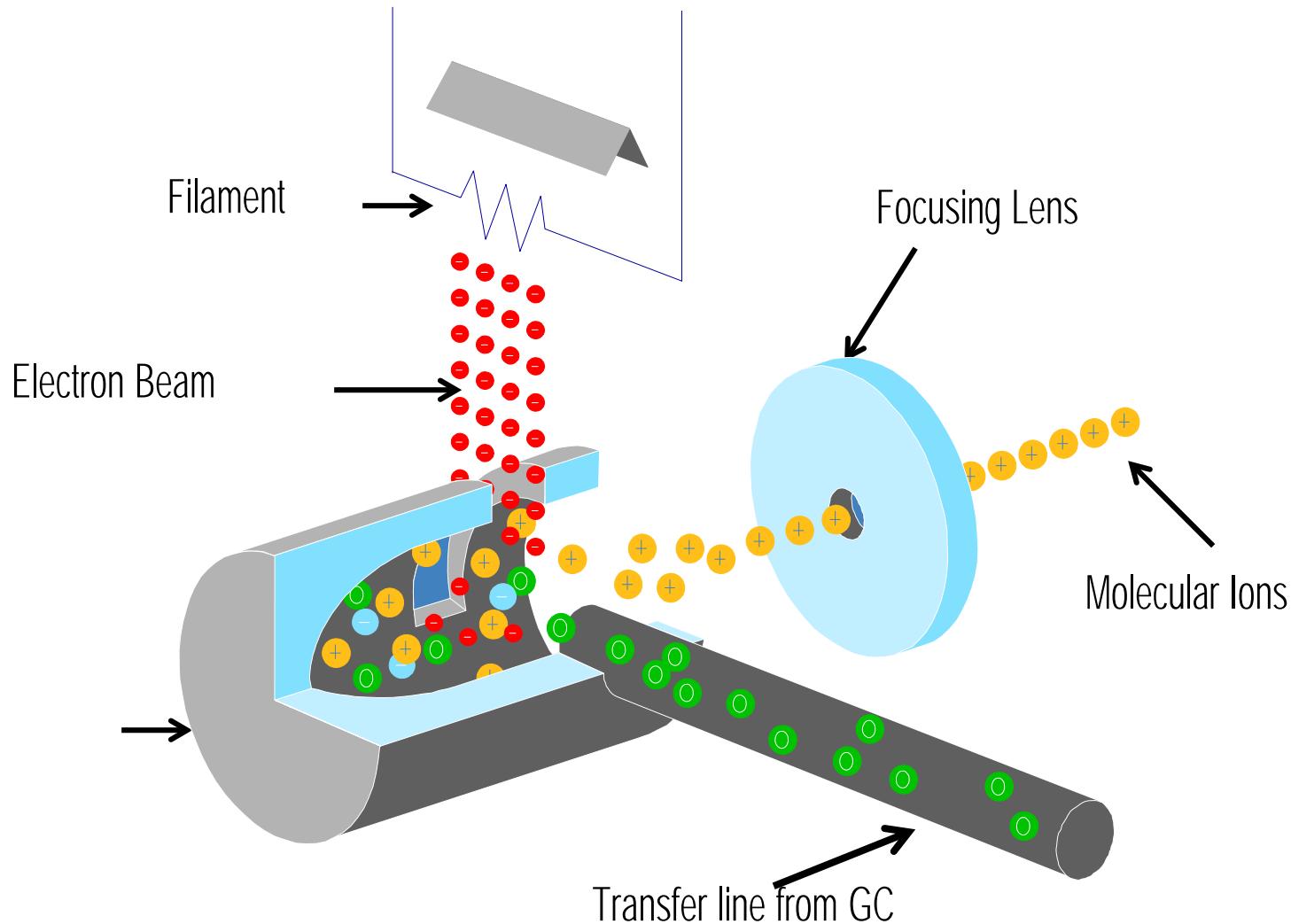
Ion Source Cartridge (iSQ)



Ionization Methods in GCMS

- Electron Ionization
- Chemical Ionization
 - Positive Ion Chemical Ionization
 - Negative Ion Chemical Ionization

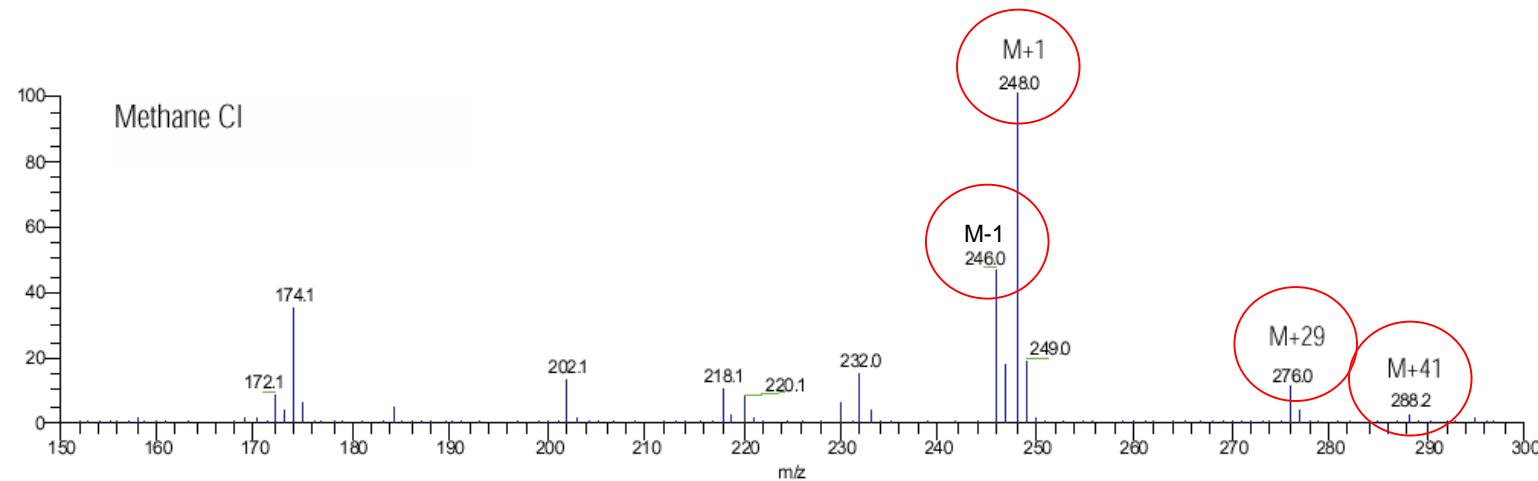
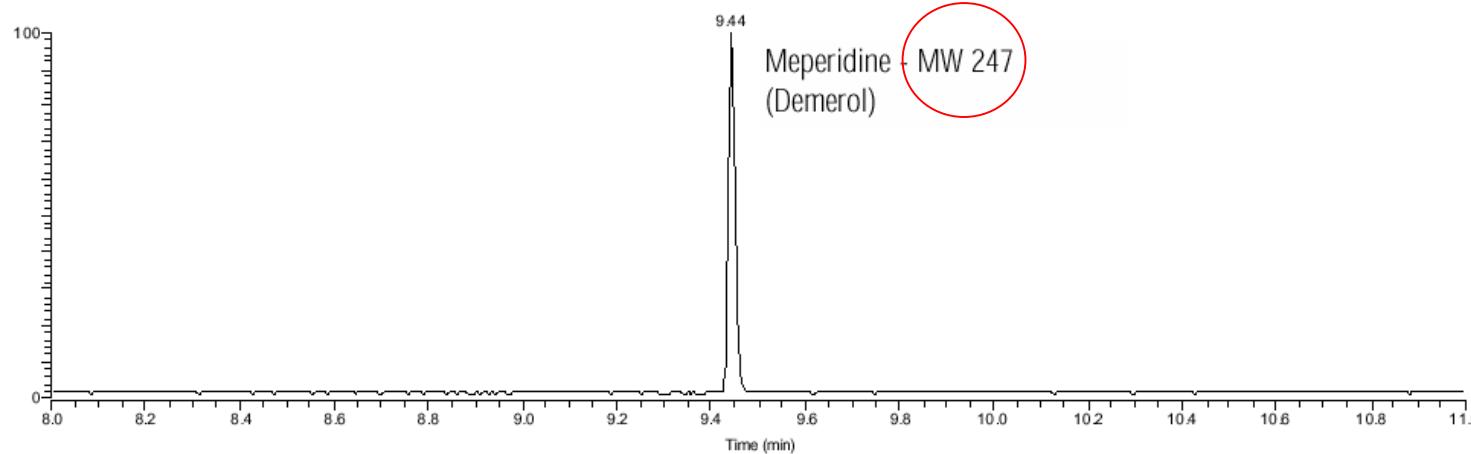
Electron Ionization



PCI : Positive Ion Chemical Ionization

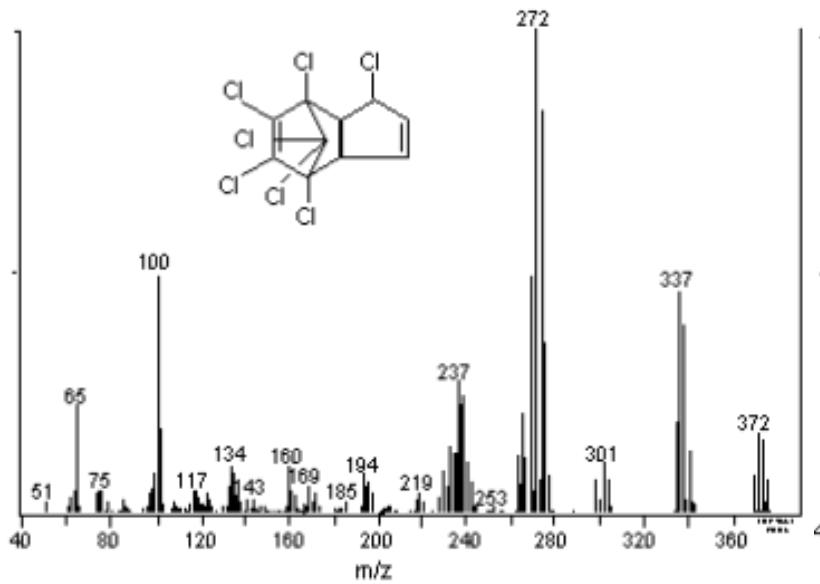
- Reagent gas reacts with electrons to form primary ions
- Primary ions react with CH_4 and form collided ions
- Collided ions react with sample molecules (soft ionization) and form molecular ions
- Molecular ions present in form of $[\text{M}+\text{H}]^+$, $[\text{M}-\text{H}]^+$, $[\text{M}+17]^+$, $[\text{M}+29]^+$, $[\text{M}+41]^+$
- Main use is molecular weight confirmation (clean spectra)
- Example of reagent gas : CH_4 , Isobutane

Adduct Formation in PICI



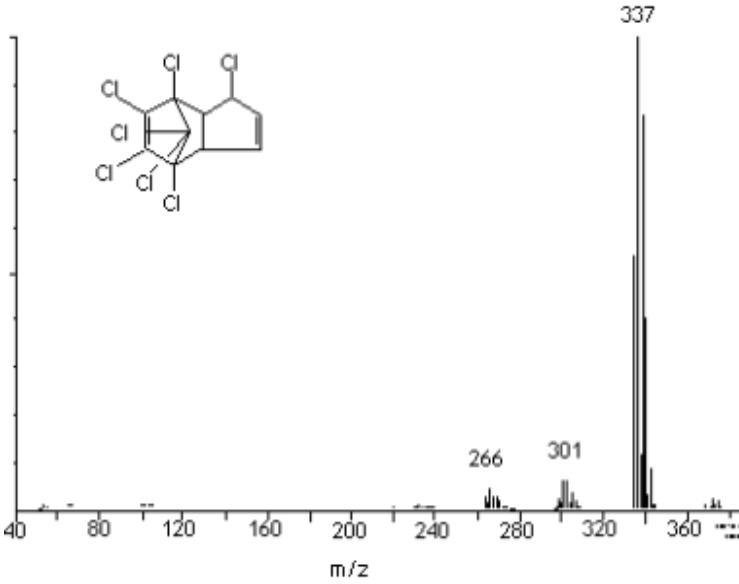
El versus PCI for Pesticides (heptachlor MW 336)

El Spectrum of Heptachlor



Intensity is low for any single m/z ion.

PCI Spectrum of Heptachlor

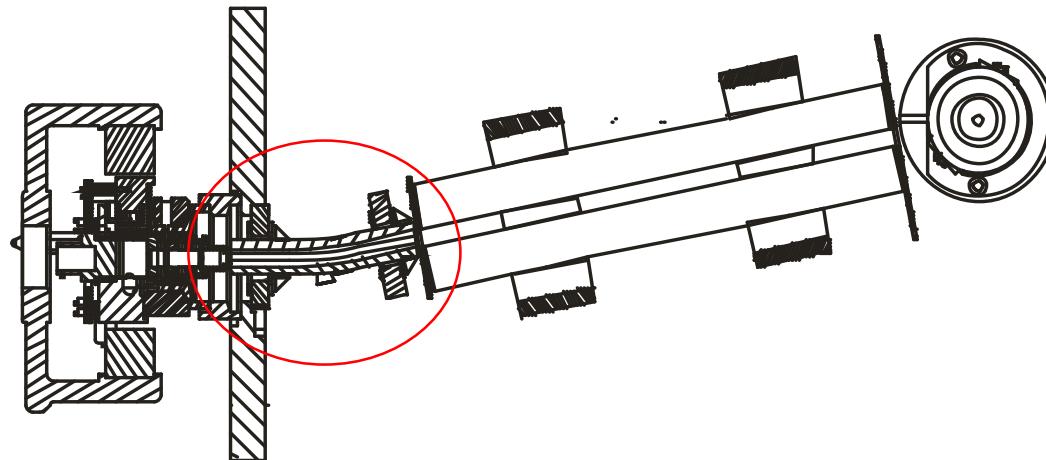
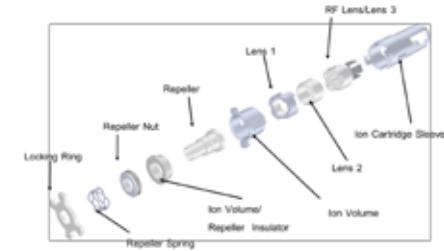


Intensity is concentrated in $[M+H]^+$ ion.
Spectrum is simpler.

In PCI, sample is not fragmented. Therefore, PCI will provide higher ion intensity
Which means better detection limit when compares with EI

Ion Transmission

- Lens :
 - Applying appropriate voltage to lens can be used to induced molecular ions into certain distance and direction
- Multi-pole rods :
 - quadrupoles , hexapoles, octapoles are widely used to transmit ions for longer distance

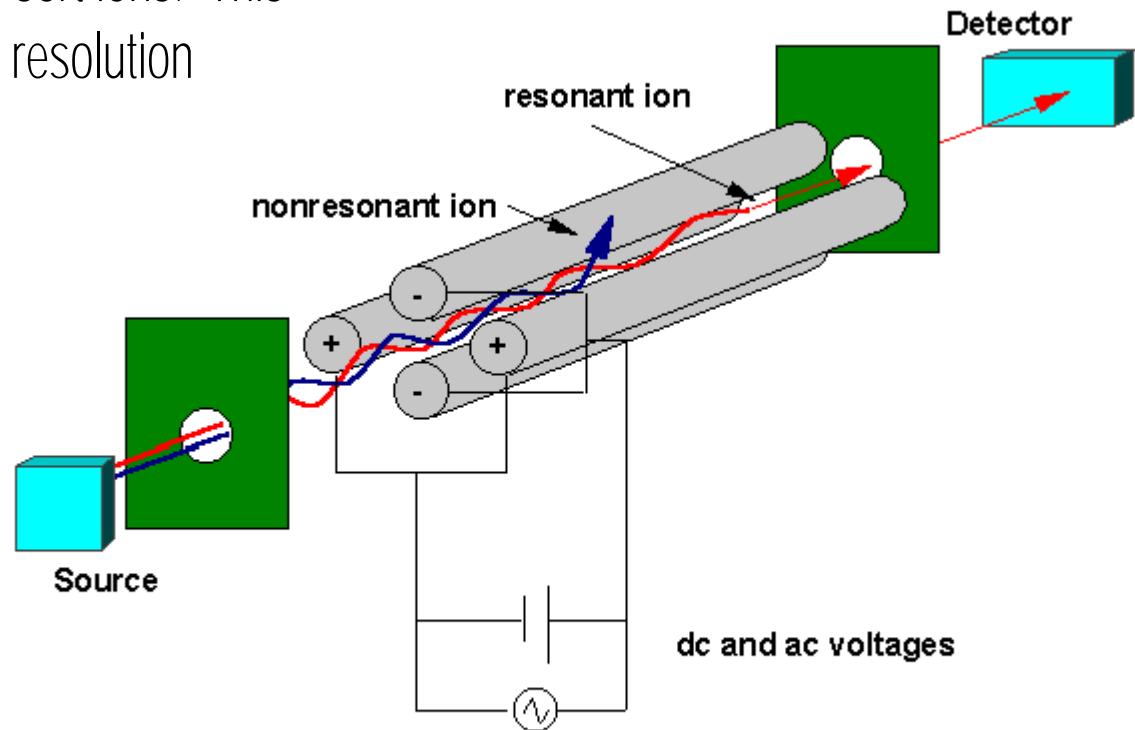


Mass Analyzers

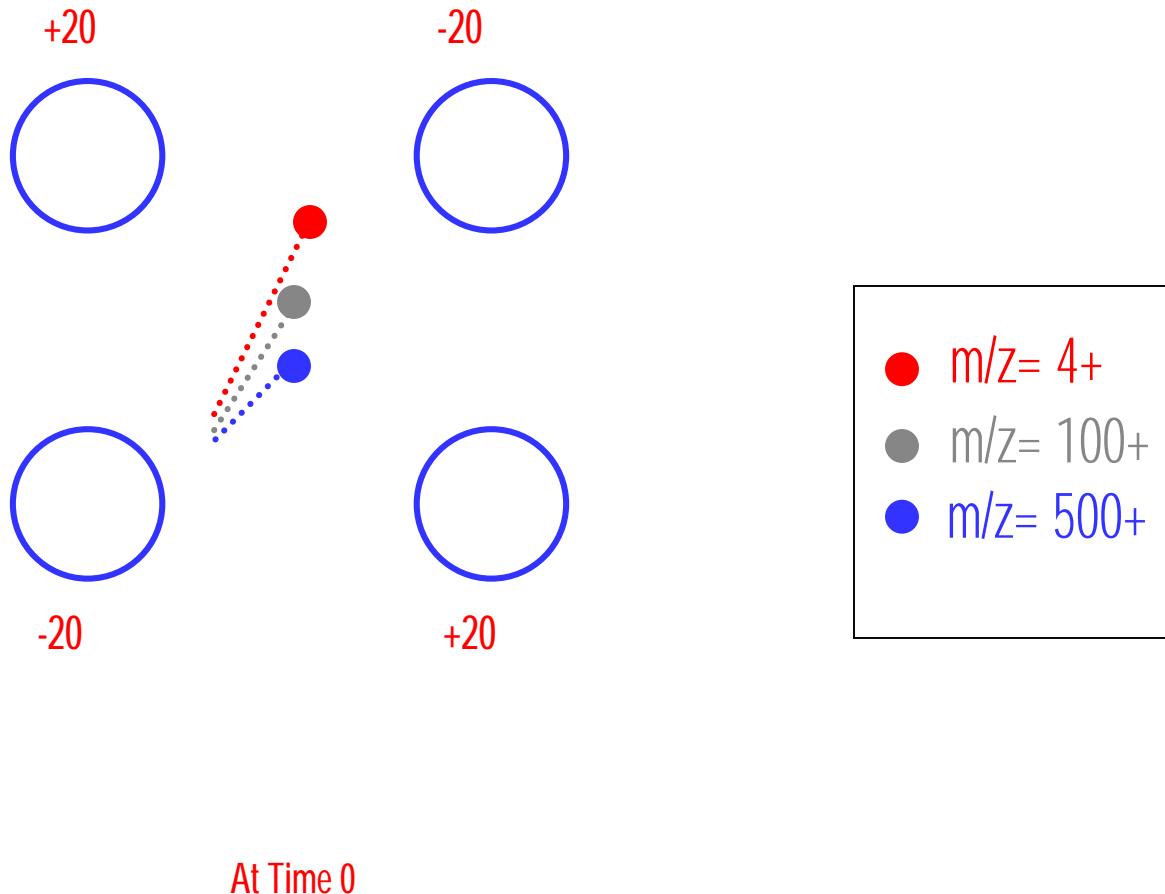
1. Quadrupole *or* Single Quadrupole
2. Triple Quadrupole
3. Time of Flight (TOF)
4. Magnetic Sector
5. Orbitrap

Single Quadrupole Mass Analyzer

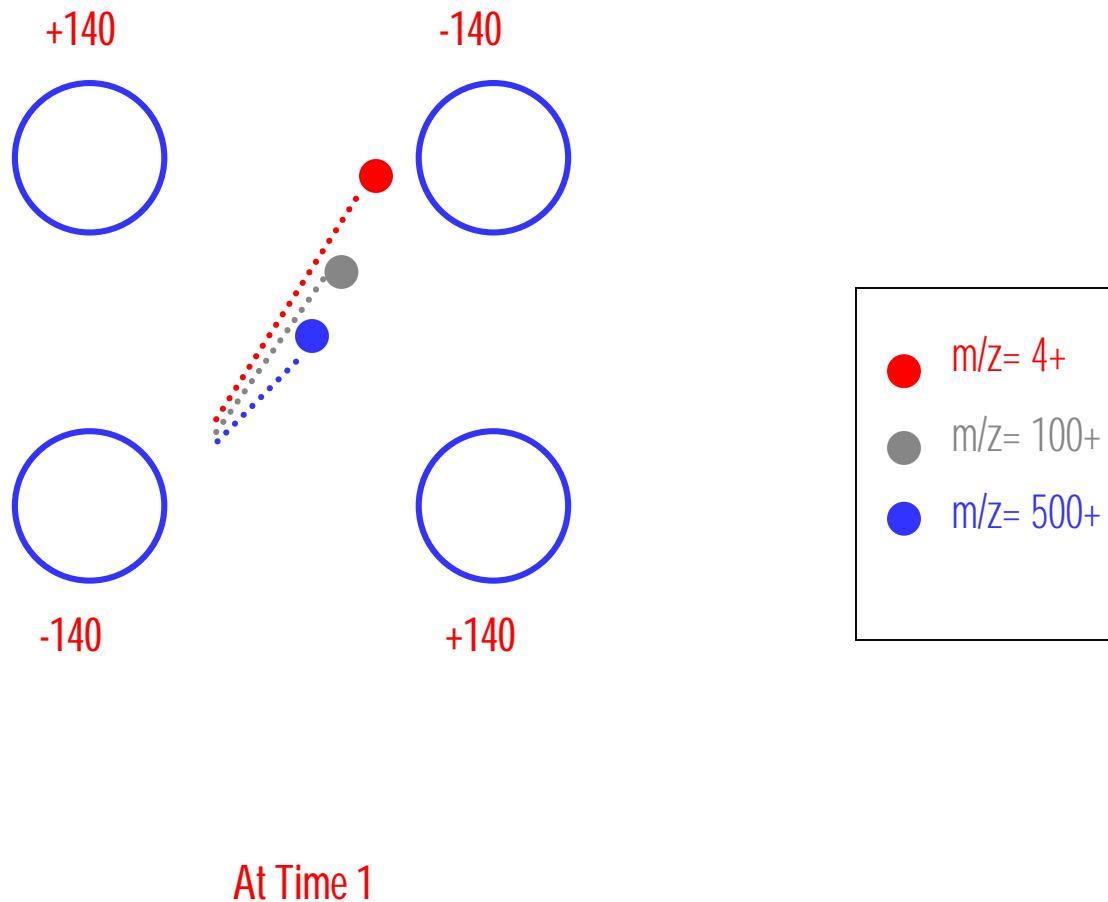
Quadrupole - consists of two sets on opposing rods. This mass analyzer uses a combination of RF(AC) and DC modulation to sort ions. This analyzer provides nominal mass resolution



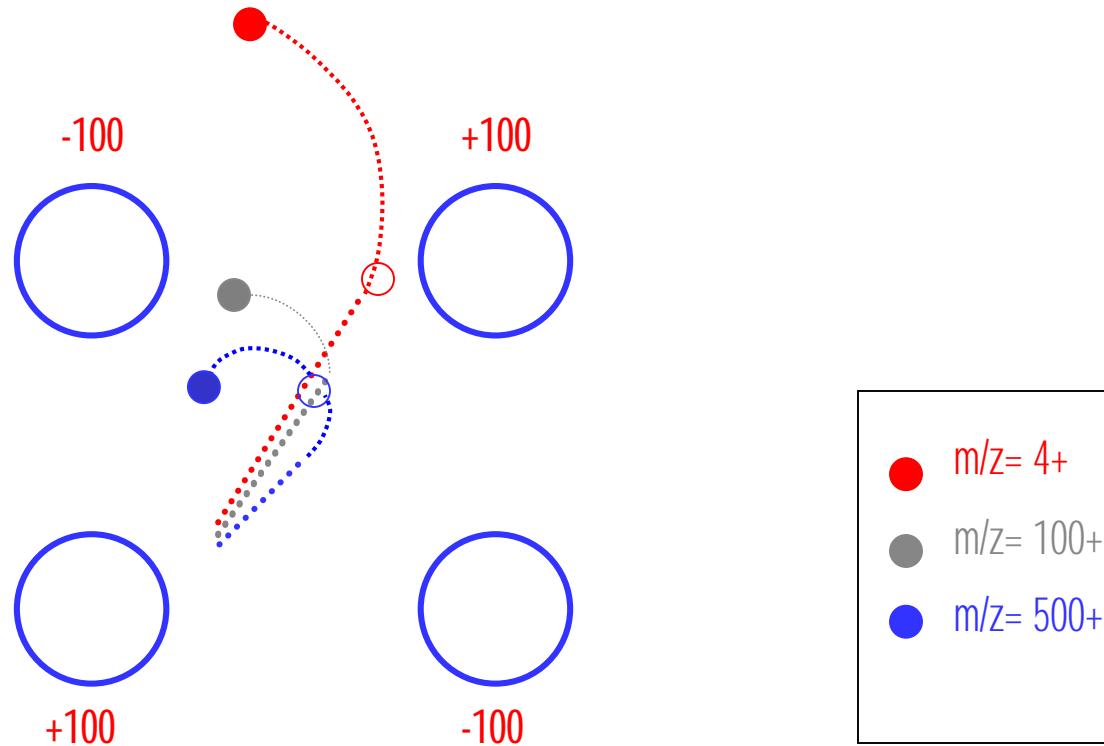
Quadupole Mass Filter Operation



Quadupole Mass Filter Operation

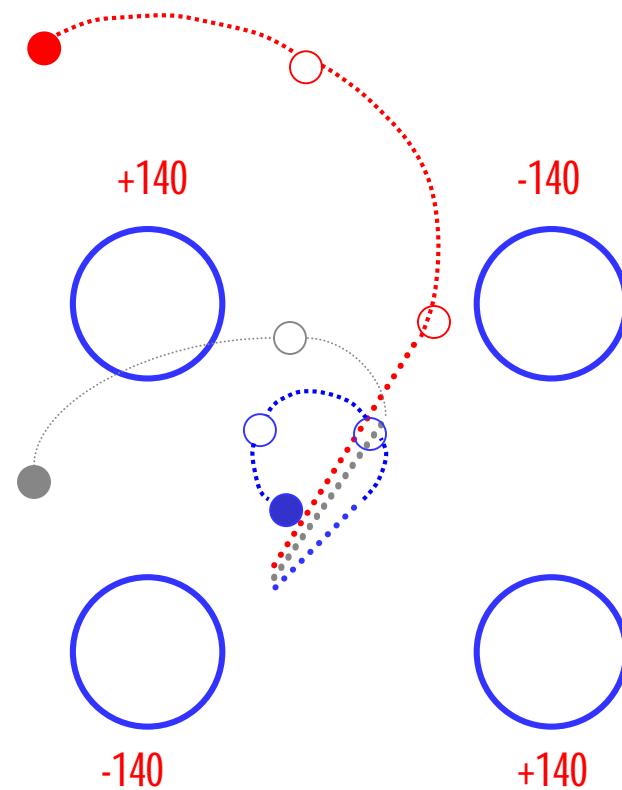


Quadupole Mass Filter Operation



At Time = 2

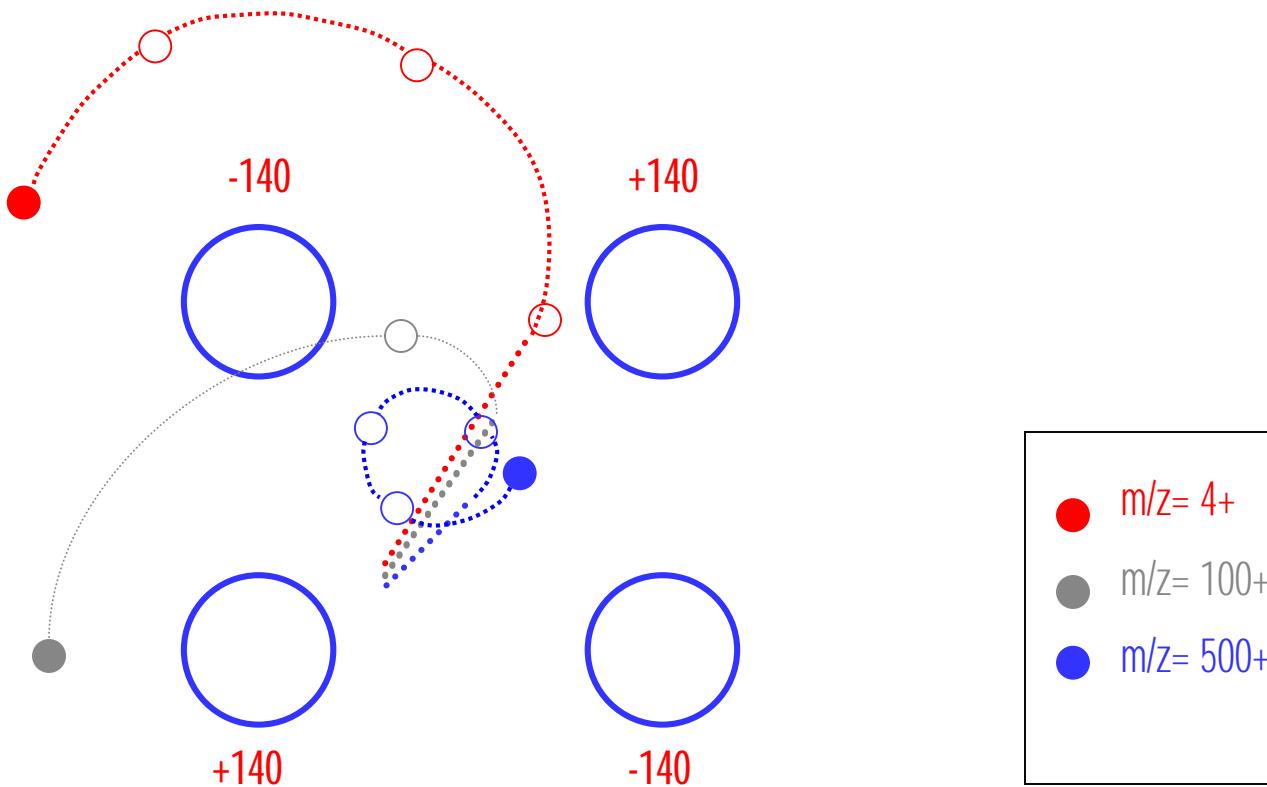
Quadupole Mass Filter Operation



At Time = 3

- $m/z = 4+$
- $m/z = 100+$
- $m/z = 500+$

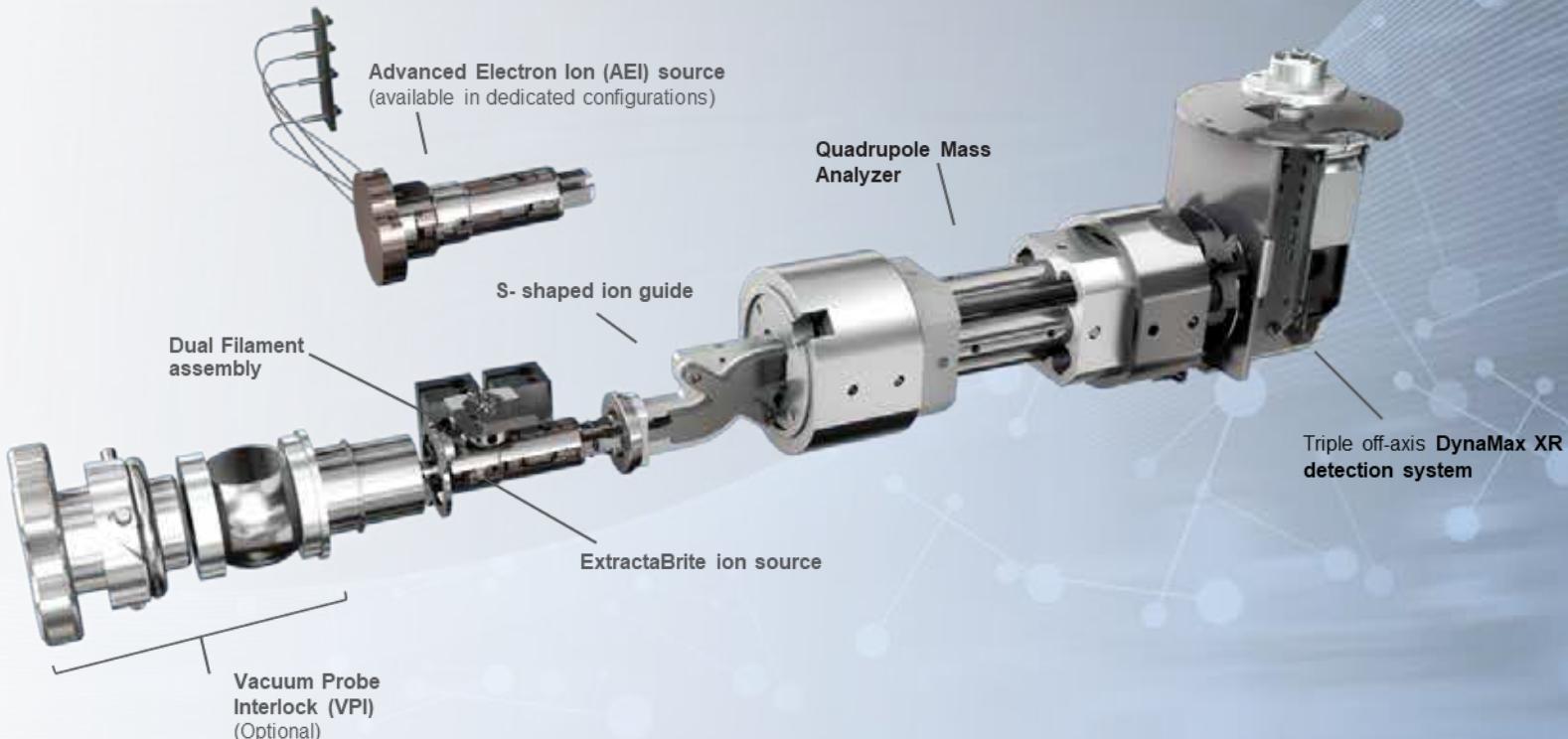
Quadupole Mass Filter Operation



At Time = 4

ISQ 7000 GCMS – Designed with Intention

ISQ 7000 GCMS – Designed with Intention

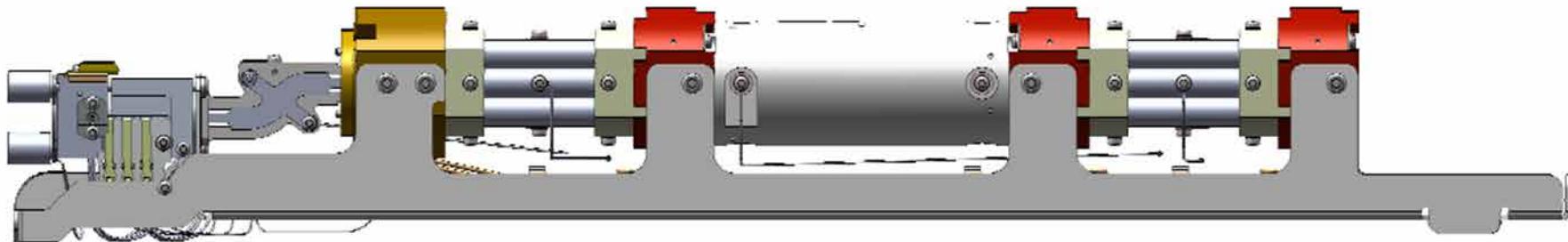


Operation modes in Single Quad MS

- Full Scan
 - Set a mass range to cover sample's molecular ions
 - Get spectrum for identification
 - Good for unknown but Low sensitivity
- Selected Ion Monitoring (SIM)
 - Select one or a few molecular ions those will be monitored
 - Lost spectrum information
 - High sensitivity but may cause false positive error

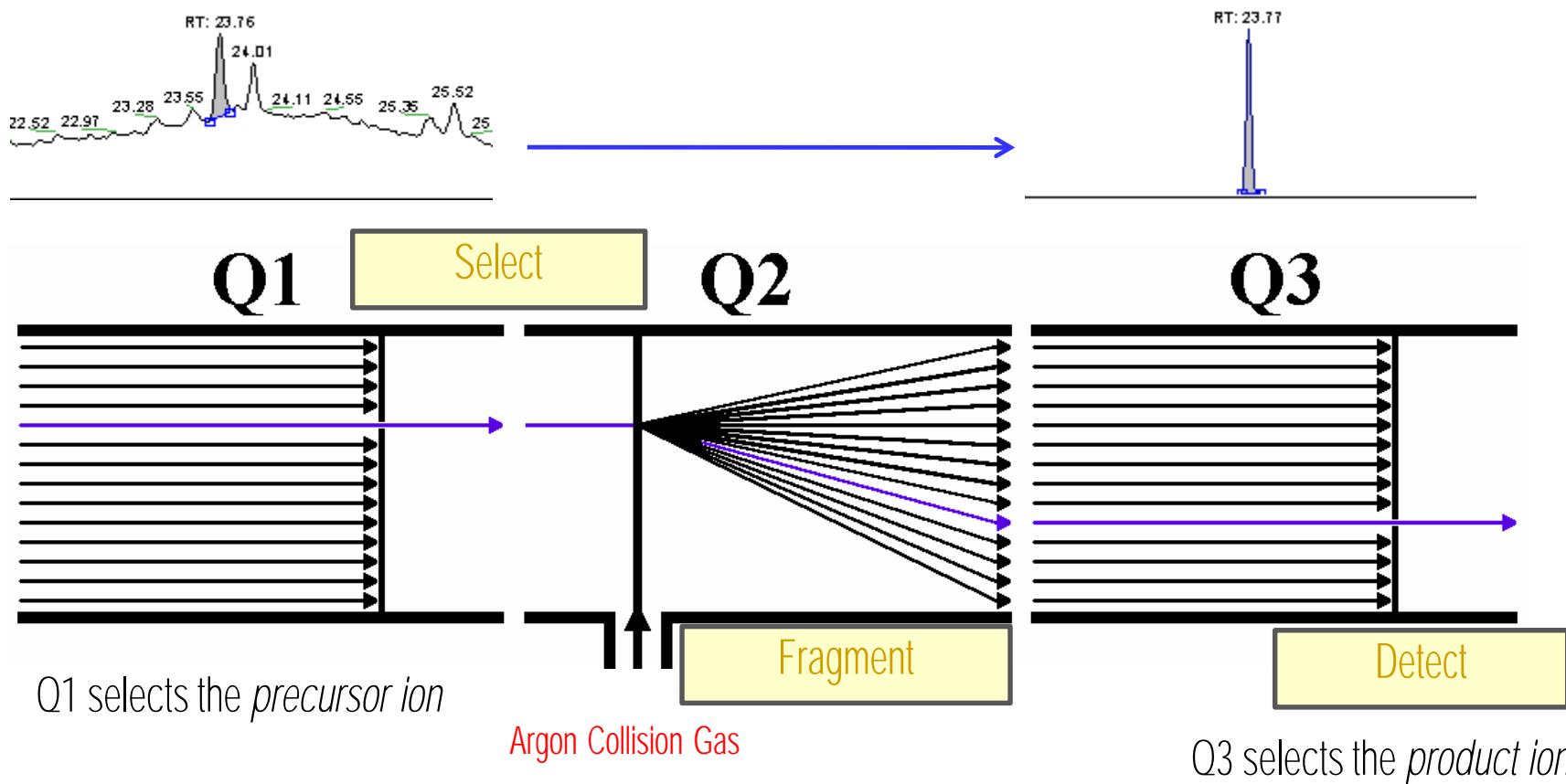
Triple Quadrupole Mass Analyzer

- Triple Quadrupole - consists of two sets of quadrupole with one collision cell in between. This mass analyzer uses a combination of RF and DC modulation to sort ions just like single quadrupole. Q1 and Q3 work like mass filter (using RF and DC) while Q2 works as a Collision cell (RF only and Collided gas). Q1 can selected any precursor (parent mass) and pass it into collision cell (Q2) where precursor are fragmented and pass through Q3 for ion sorting again. This analyzer provides high sensitivity with fast confirmation analysis.

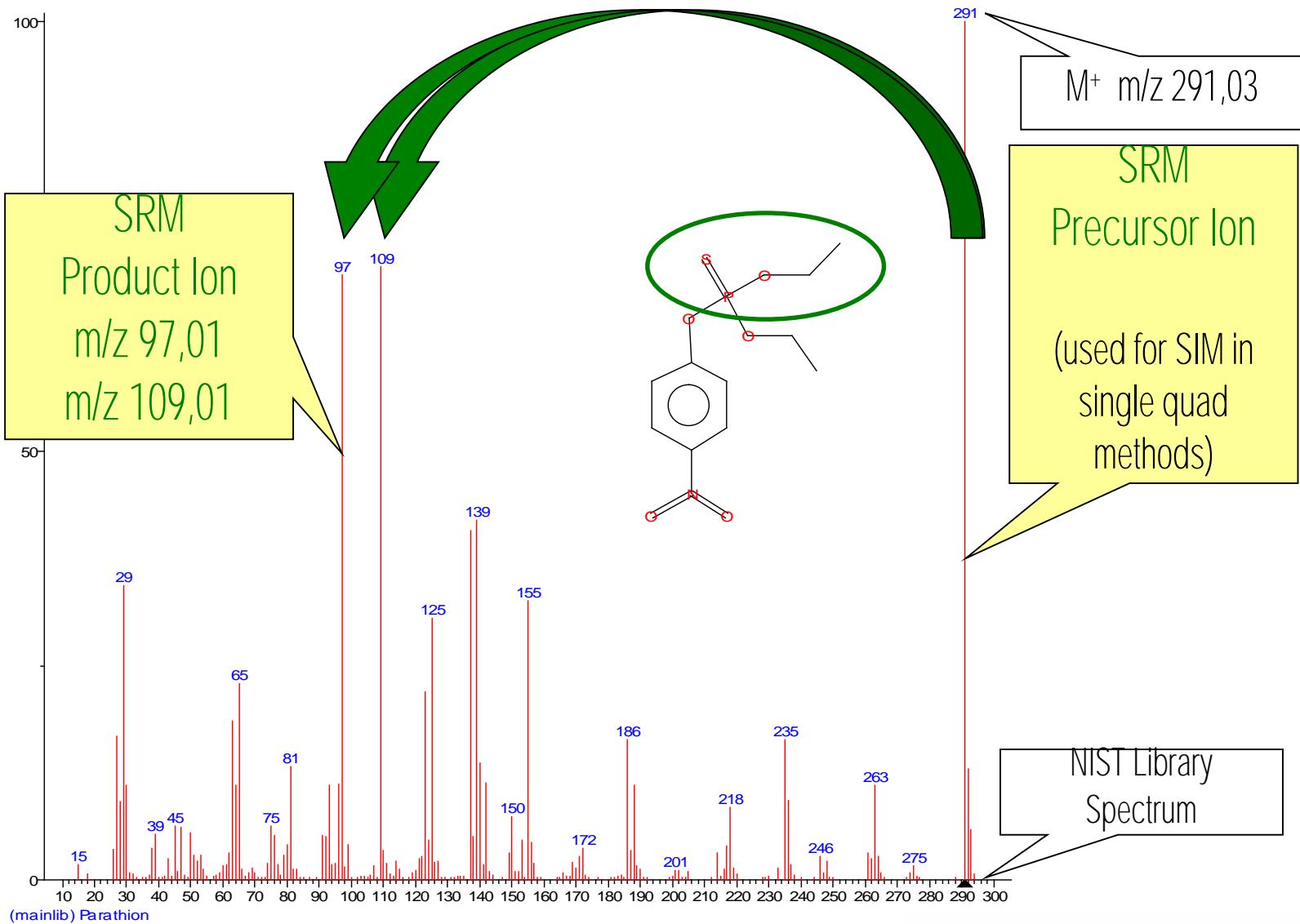


Selected Reaction Monitoring (SRM or MRM)

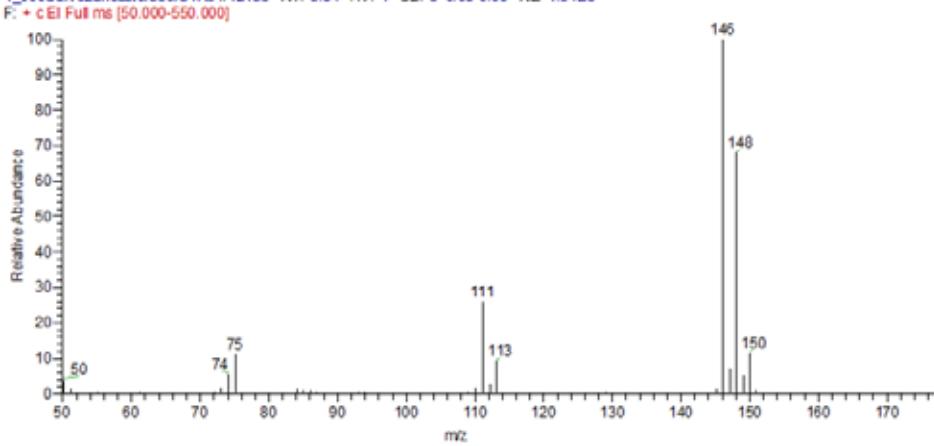
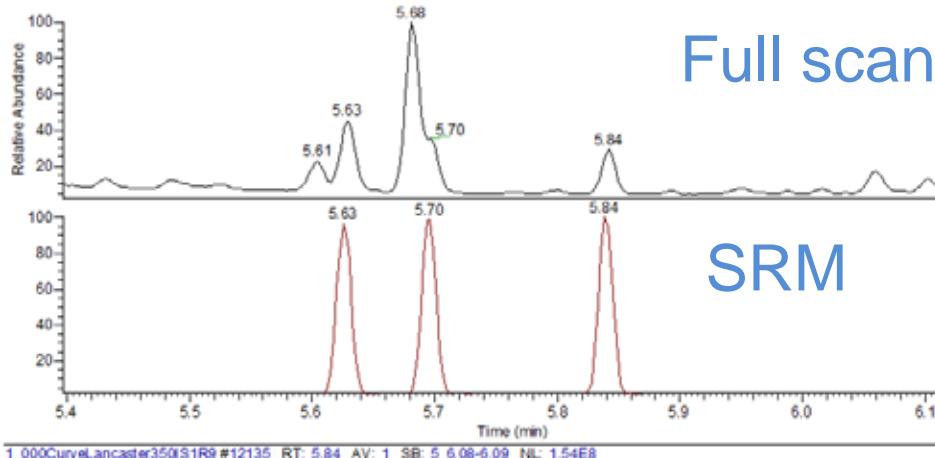
Quantitation of target compounds in matrix samples



Structure Specific Selectivity by QQQ : Parathion-Ethyl

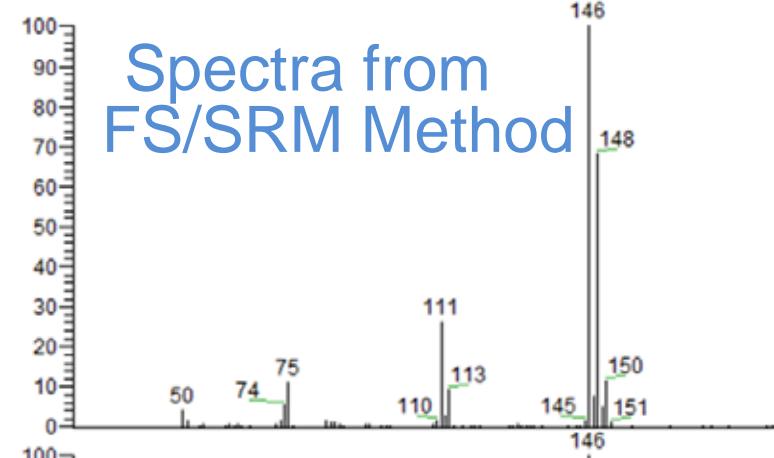


Full scan/SRM Acquisition

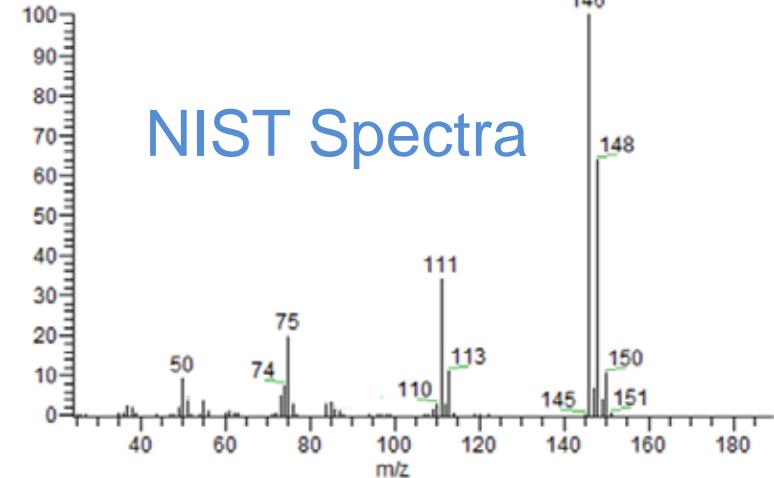


Full scan

SRM



Spectra from
FS/SRM Method



NIST Spectra

Detector : Dynode Electron Multiplier

- Dynode converses Molecular ions into electron
 - Continuous Dynode
 - Discrete Dynode
- Electron are then sent to multiplier for signal enhancing

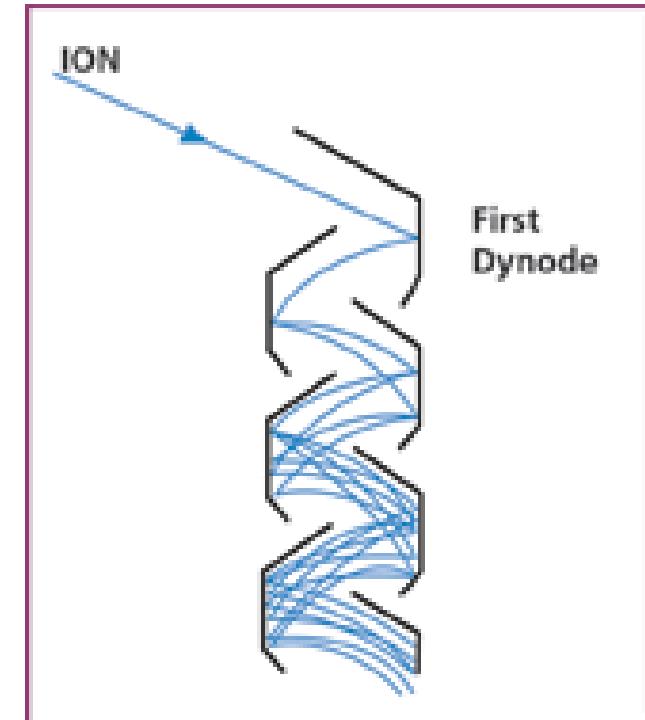
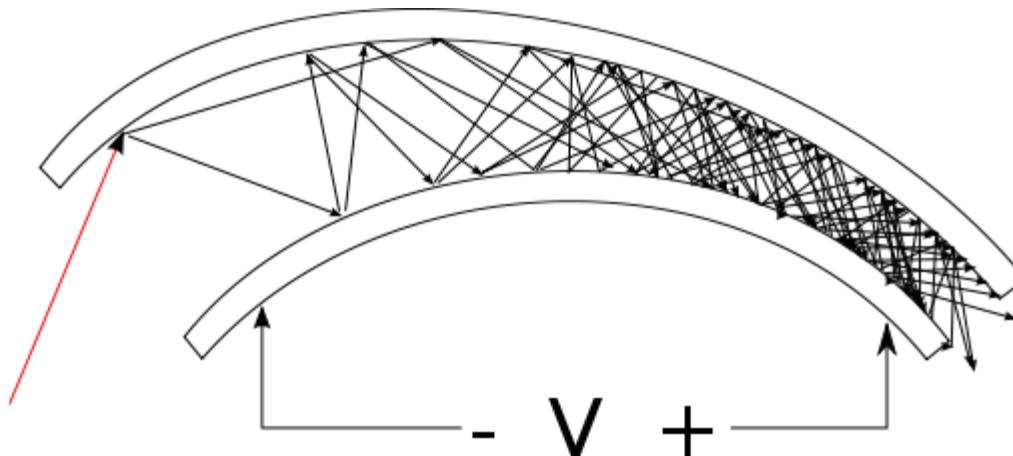
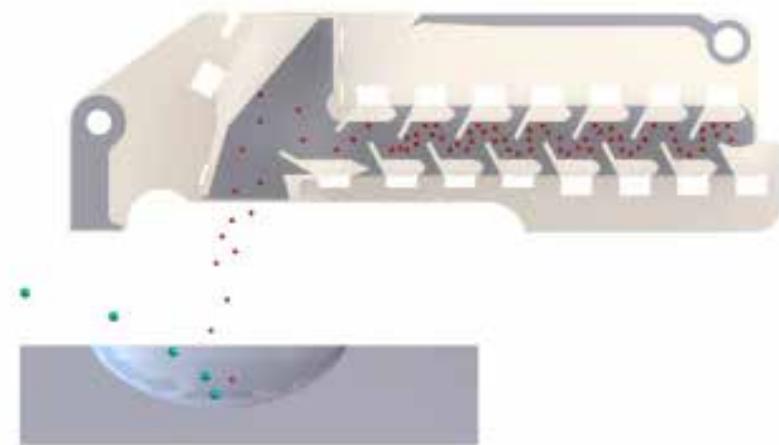


Photo courtesy from SGE & ETP, Wikipedia

Off -axis dynode and EM

- Off axis dynode
 - High voltage is applied (+/-10 KV) for high signal (accelerate ion velocity from mass analyzer to dynode)
 - Induces only molecular ions to hit dynode
- Electrons from dynode hit internal wall of EM.
- Multiplication process amplifies for more signals

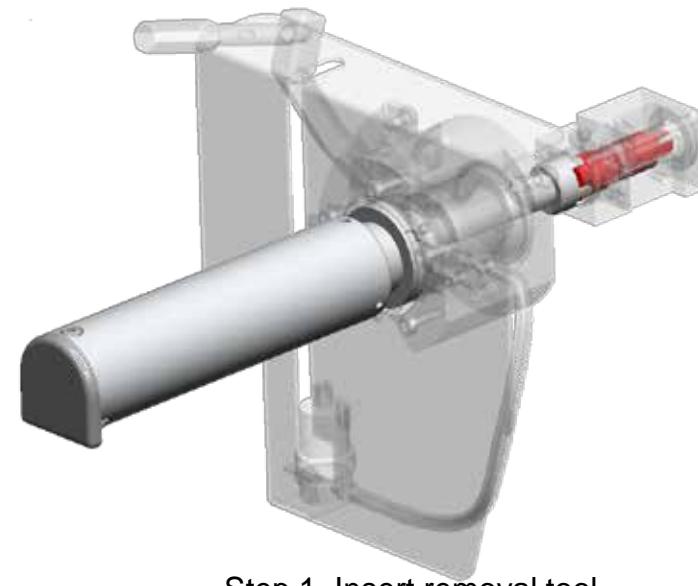
Electron Multiplier



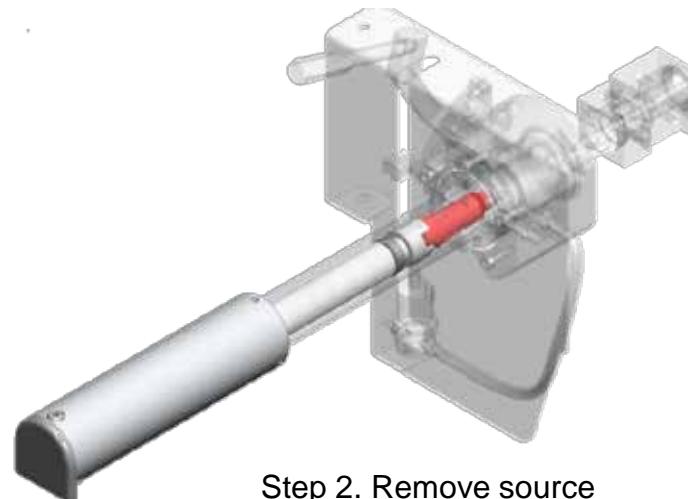
Dynode

- High Vacuum Pumps (10^{-3} to 10^{-10} Torr)
 - Oil Diffusion
 - No moving parts
 - Turbomolecular
 - Clean - no oil
- Mechanical Backing Pump, (Fore Pump) (atm. to 10^{-3} Torr)
 - Rotary vane

User maintenance : Vacuum probe interlock



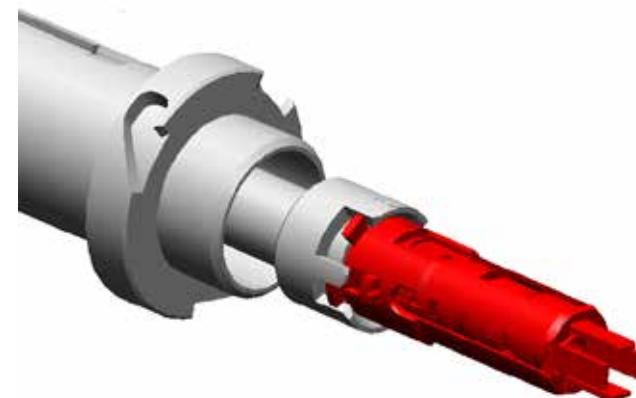
Step 1. Insert removal tool



Step 2. Remove source

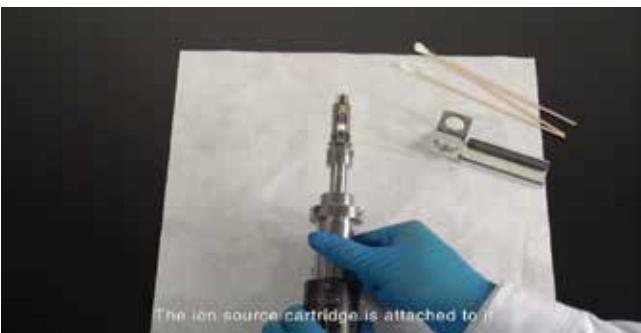


Step 3. Hot source is held in tool



Step 4. Push source out of tool

User maintenance : Vacuum probe interlock



Perfect for today, ready for tomorrow

- Fit for purpose GC-MS solution
- Grows with evolving regulatory requirements
- Base to advanced configurations
- Full field upgrade path

Affordable first entry
66L/s ExtractaBrite



Accessible high performance
300L/s ExtractaBrite



Ultra high sensitivity and robustness
ISQ 7000 AEI



High-throughput solution
ISQ 7000 NeverVent EI & CI



High-throughput solution
ISQ 7000 NeverVent EI



Perfect for today, ready for tomorrow

- Grows with laboratory requirements
- Base to advanced configurations
- Full field upgrade path

Most accessible entry from SQ>TQ
240L/s ExtractaBrite



Affordable performance
300L/s ExtractaBrite



High-throughput solution
TSQ 9000 NeverVent EI



Ultra high performance
and robustness
TSQ 9000 AEI



High-throughput solution
TSQ 9000 NeverVent EI & CI



Application.....

Multi-Residue Pesticide Analysis in Herbal Products Using Accelerated Solvent Extraction with a Triple Quadrupole GC-MS/MS System

Hans-Joachim Hildebrandt, Joachim Gummertbach, Hans-Peter Lautenbacher, Johann Kettner, Birgit Höller, Physio

Key Words:
Pesticides, Tea, Herbal products, ASE, SRM, MRM, Multi-residue at TSG 8000 GC-MS/MS

Introduction

The residue analysis of pesticides has developed over years into a comprehensive methodology for the detection of many hundreds of potential contaminating compounds. A multi-residue method for herbal products is faced with additional challenges from the possible origin of the products and the complex matrix of plant materials.

In the due control of food, plant protection treatments must be taken into account a wide variety of potential pesticide contaminants. Dried leaves, fruits or seeds and other herbal products usually contain highly complex extracts from simple preparation due to the rich content of active ingredients, essential oils and the typical high heat-stable polyphenol compounds from broken cells, in their cases. A thorough clean up of the extracted samples leads to losses of critical analysis of interest. A complete characterization of the sample, and other resubstitution is done by both LC and GC-MS/MS.

the complete range of functional groups.

This application report describes methodology used for the multi-residue analysis of herbal tea using accelerated solvent extraction (ASE) and gas chromatography-mass spectrometry (GC-MS) detection and quantification by Thermo Scientific™ TSG 8000 GC-MS system.



Thermo
SCIENTIFIC

Application Note: 10212

Determination of Total FAME and Linolenic Acid Methyl Ester in Pure Biodiesel (B100) by GC in Compliance with EN 14103

Fabrice Meunier, Daniela Catagnini, Andrea Cadoppi, Thermo Fisher Scientific, Milan, Italy

Introduction

An oil for biodiesel to be commercialized as pure biofuel or blending stock for heating and diesel fuels, must meet a set of requirements defined in ASTM D6751 and EN 14103 standard specifications.^{1,2} These specific ratios indicate the maximum allowable concentrations of unsaturation in pure biodiesel (B100) finished product, along with other chemical/physical properties necessary for a safe and satisfactory engine operation.

Gas chromatography (GC) is commonly adopted to characterize pure biodiesel (B100) according to the following standard methods:

- EN 14103: Determination of Total FAMEs (linoleic acid methyl ester) and Linolenic Methyl Ester (C18:3)
- EN 14350/ASTM D6388: Determination of FAME and Total Glycerine^{1,3,4}
- EN 14430: Determination of residual Methanol⁵

Comprehensive Thermo Scientific GC solutions have been developed in compliance with each of these methods, based on the Thermo Scientific™ TRACE GC Ultra[®] and the versatile TriPlus[®] auto-sampler (Figure 1). This application note relates to the determination of total FAME and linoleic acid methyl ester in biodiesel according to EN 14103.

This application report describes methodology used for the multi-residue analysis of herbal tea using accelerated solvent extraction (ASE) and gas chromatography-mass spectrometry (GC-MS) detection and quantification by Thermo Scientific™ TSG 8000 GC-MS system.

Figure 1: Thermo Scientific TRACE GC Ultra with TriPlus auto-sampler

Application Note: 30112

Confirmation of Low Level Dioxins and Furans in Dirty Matrix Samples using High Resolution GC/MS

Dieter Kremser, Hans-Joachim Hildebrandt, Thermo Fisher Scientific, Bremen, Germany

Introduction

Over the past 30 years, dioxin/TCDD levels and body burden levels in the general population have been on the decline and continue to decrease^{1,2}. More than 95% of human exposure to dioxins and dioxin-like substances is through food.³ With increasingly lower dioxin levels in food, feed, and tissues, more demanding limits of detection, selectivity, sensitivity and QC checks are required to confirm their presence at these ever decreasing levels.

Methods

Instrumentation

A Thermo Scientific™ PTV 1000 with liquid detector (LID), a 100 m column, 100 µl injection volume, Chem-Card data system, Thermo Scientific™ 0.25 mm ID, 0.2 µm film thickness was used for the analysis. The temperature program is shown in Table 1.

The injection was performed using the hot needle technique. The capillary needle was heated up to the injection for 2-3 seconds before injecting the sample, thus eliminating any discrimination of higher boiling components.

The DRS mass spectrometer was set up in the multiple ion detection mode (MID) at a resolution of 10,000 (10% valley definition). PCP4 was used as a reference compound to provide the inherent back and salt masses. Their reference masses are monitored scan-to-scan to insure the highest mass precision, stability and ruggedness necessary for routine target compound analysis on a high resolution mass spectrometer. For all native dioxin/PCP4 congeners, as well as for their specific ^{13C} labeled internal standards, one quantitative mass and one ratio mass were implemented in the MID set up, as shown in Table 2. The effective resolution is constantly monitored on the reference masses and documented in the data files for each MID window.

The new derivatives demonstrate the continuing need for ever more sensitive analytical instrumentation. As an example, the new methods for confirmation require limits of quantitation (LOQ) to be 10% lower than the lowest reported level in the method. This requires the instrumentation to reach even lower levels of detection, and reduce the necessary sample volumes needed for analysis.

Application Note: 52242

Identification and Quantification of Impurities in Wines by GC/MS

Brenda Gauvin-Denney, Eric Phillips, Stacy Crum, Tracy Rothery, Thermo Fisher Scientific, Austin, TX, USA
(With special thanks to members of Theologic Center of GezaG)

Introduction

While wine makers have historically used gas chromatography and mass spectrometry (GC/MS) to detect pesticides, they now more commonly use the technique to supplement quality control checks of wine taste. Without GC/MS,



that determine wine purity *in situ* rather than having to send samples for expensive, external analysis. In this report, we present the design and results of this study, including the experimental method used to detect impurities and the concentration ranges that compare GC/MS with human detection.

Methods

For this experiment, several targeted molecule types that affect wine quality were analyzed using an "ISO" Single Quadrupole GC-MS system (Figure 1). Table 1 contains a brief description of the effects on wine quality of the four target molecule types, and examples of how GC/MS analysis can provide value in quality control.

Set up & Run

Results of GC/MS Analysis

Analysed in various ways to determine the lowest concentration of furan lactones. The best FID, 4-ethylphenol and 4-ethoxyphenol are "target" GC/MS quality.

Set-up: detector from ready to run: GC/MS quality. Detecting phenols in wine matrix makes the job easier. Set up to run: 100 µl of wine + 100 µl water + dilute phenol to 1000 µl. Set up to run: 100 µl of wine + 100 µl water + dilute phenol to 1000 µl. Set up to run: 100 µl of wine + 100 µl water + dilute phenol to 1000 µl.

Run: from 100 µl to 1000 µl. Phenol detector: 4-ethylphenol and 4-ethoxyphenol are "target" GC/MS quality. Phenol detector: 4-ethylphenol and 4-ethoxyphenol are "target" GC/MS quality. Phenol detector: 4-ethylphenol and 4-ethoxyphenol are "target" GC/MS quality. Phenol detector: 4-ethylphenol and 4-ethoxyphenol are "target" GC/MS quality.

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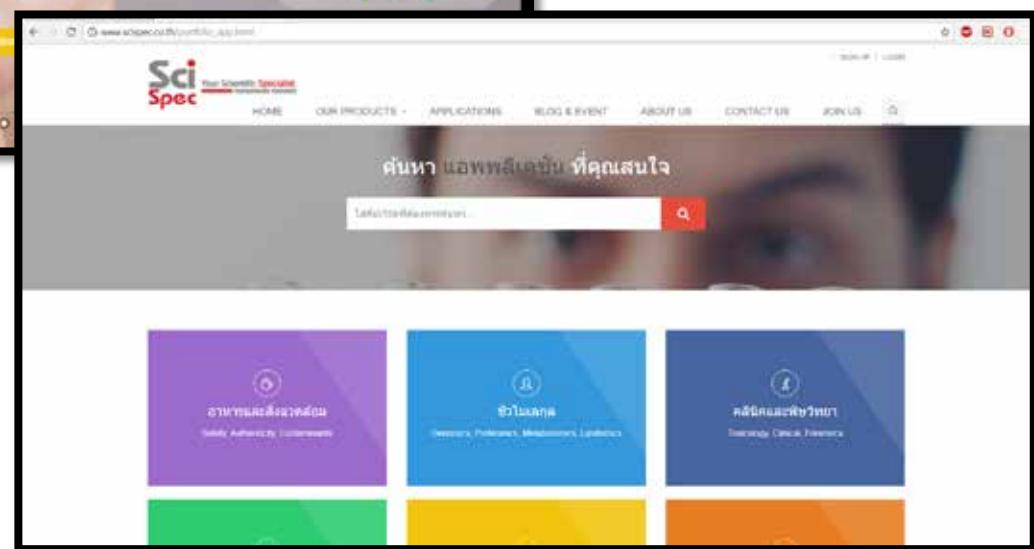
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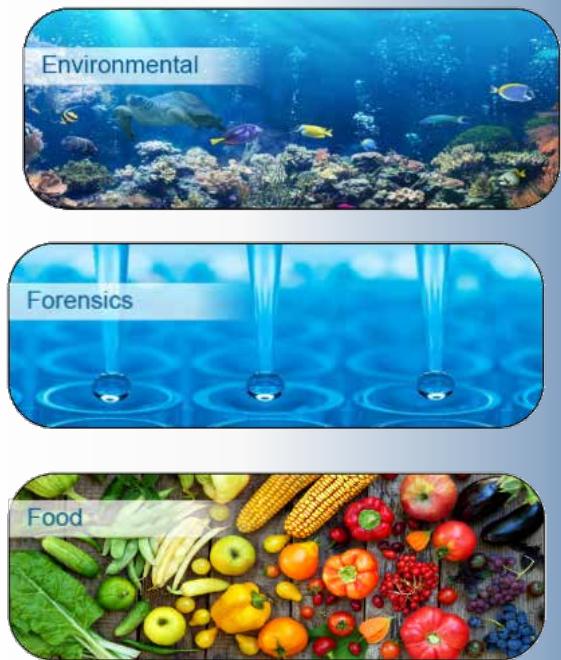
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Scispec website : Application.....

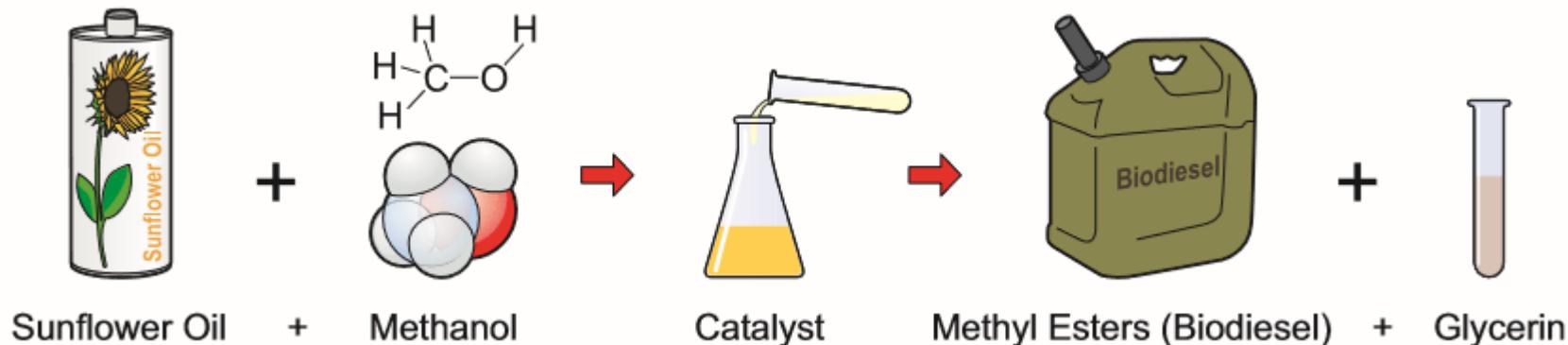
www.scispec.co.th



GC and GCMS application support.



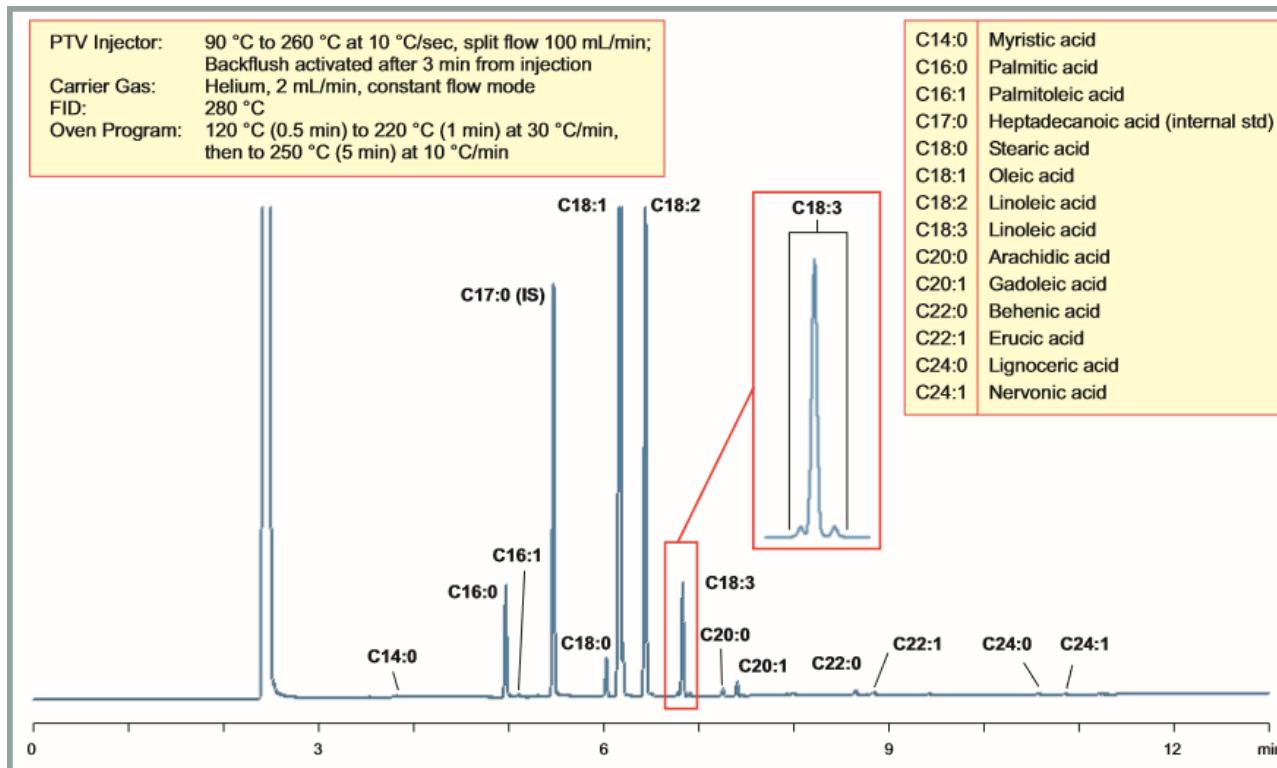
Application : Biodiesel



- Total FAME and Linolenic Acid Methyl Ester : EN 14103
 - Free and Total glycerin : ASTM D6584 / EN 14150
 - Methanol Content : EN 14110

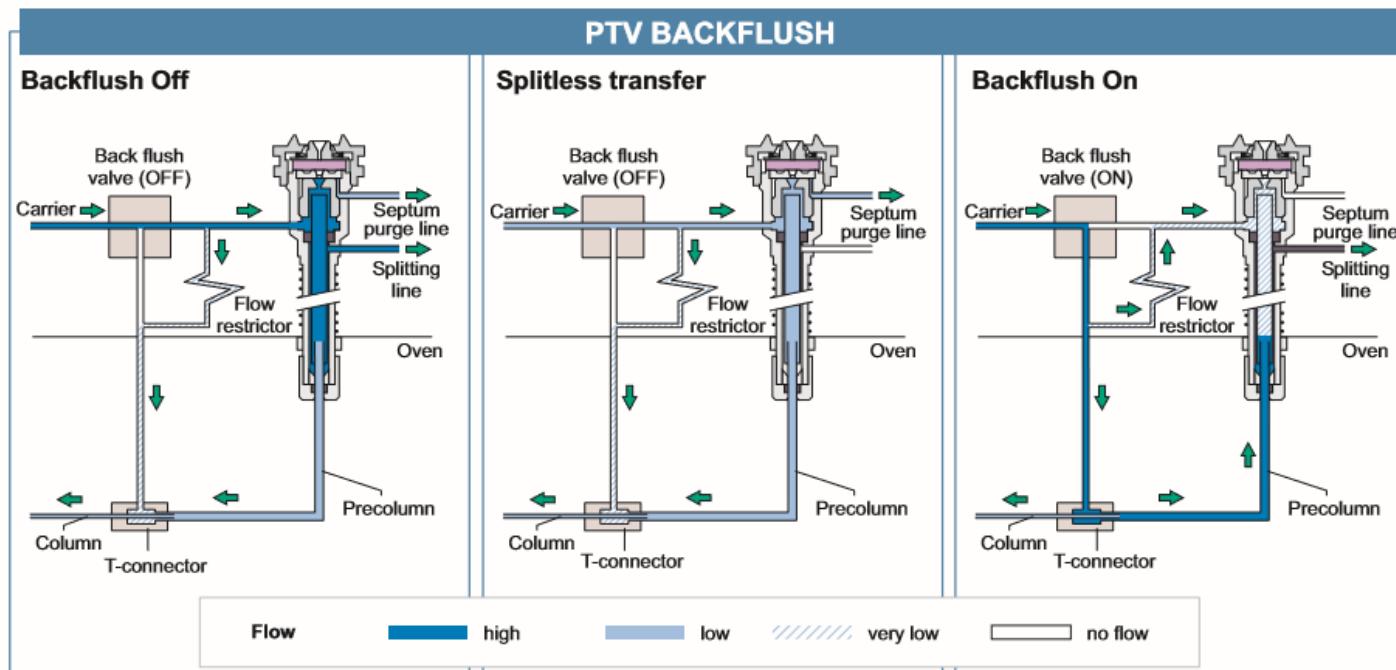
Total FAME and Linolenic Acid Methyl Ester : EN 14103

The cetane number of biodiesel depends on the distribution of fatty acids in the original oil. Thus a reliable characterization of FAME is essential for a more accurate calculation of the cetane index. EN 14103 is a standard method for determination of esters and linolenic acid methyl ester and can be applied to biodiesel analysis. EN 14103 requires GC analysis.



By incorporating the backflush option into the PTV injector, heavy compounds can be vented out of the inlet system, effectively preventing column contamination while still allowing efficient transfer of compounds of interest.

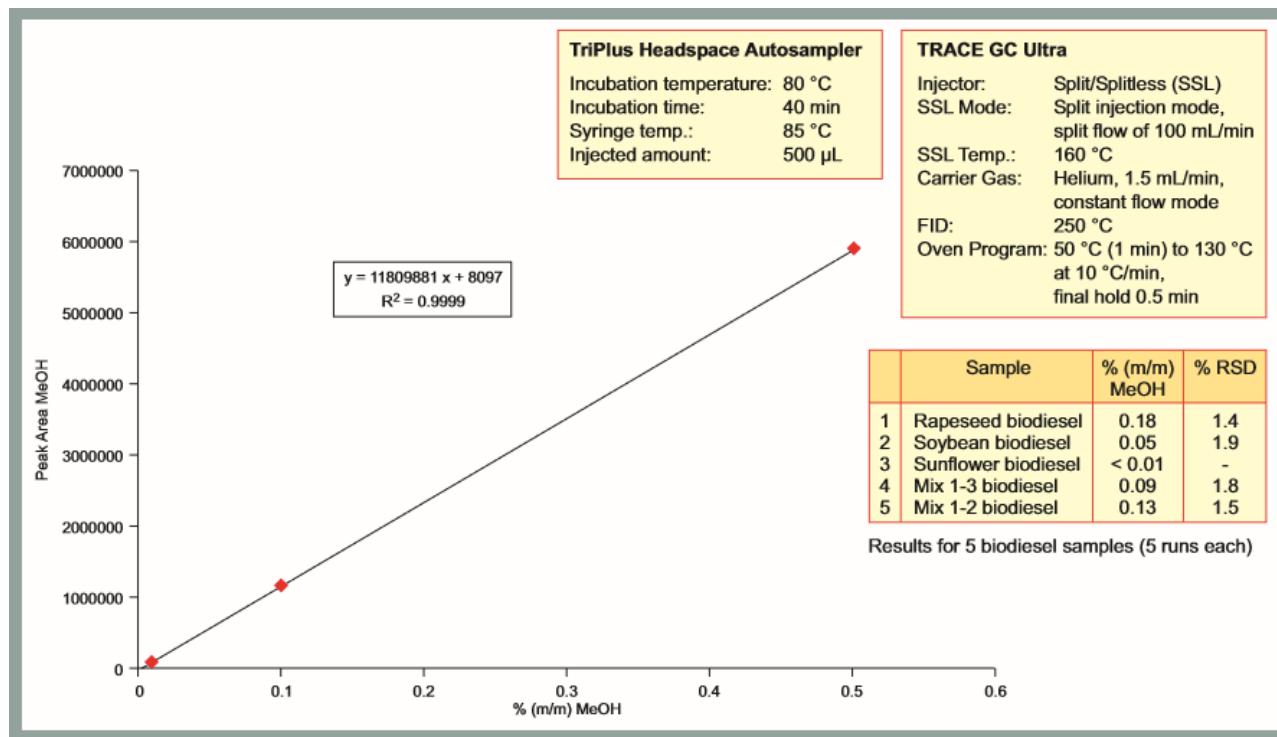
PTV Backflush



PTV Backflush (reverse flow device): the glycerides fraction is vented out without entering the column.

Methanol Content : EN 14110

Methanol in B100 is a matter of safety since even small amounts of this material can reduce the flash point of the biodiesel. Moreover, residual methanol can affect fuel pumps, seals and elastomers and can result in poor combustion properties. EN 14110 requires a headspace GC method, based on either polar or non-polar columns, and is applicable for a concentration range from 0.01% m/m to 0.5% m/m of methanol (MeOH).



Multi-Residue Pesticide Analysis in Herbal Products Using Accelerated Solvent Extraction with a Triple Quadrupole GC-MS/MS System



Sample Preparation

Dried leaves , fruits or seeds and other herbal products

Weight 10 g of sample.

Mixed with DE and load into the extraction cells.

Concentrated Sample and injection with GC



ASE™ 350

Sample weight	10 g
Extraction solvent	Ethylacetate/cyclo-Hexane 1:1, same as GPC solvent
Temperature	120 °C
Pressure	100 bar
Extraction time	5 min, 1 cycle
Flushing with solvent	60% of cell volume
Flushing with nitrogen	100 s

Multi-Residue Pesticide Analysis in Herbal Products Using Accelerated Solvent Extraction with a Triple Quadrupole GC-MS/MS System



GC : Condition

Injector PTV	Splitless mode
Base temperature	50 °C
Transfer	10 °C/s to 250 °C, until end of run
Flow	Constant flow, 1.2 mL/min, helium
Analytical column	40 m, ID 0.18 mm, 0.18 µm film, 5%-phenyl phase (5MS type)
Pre-column	5 m, ID 0.18 mm, empty deactivated, no backflush
Column oven	Temperature programmed
Start	70 °C, for 1.50 min
Ramp 1	15 °C/min to 190 °C
Ramp 2	7 °C/min to 290 °C, 12 min
Transfer line	250 °C

MS/MS : Condition

Ion source temperature	220 °C
MRM Detection	Timed SRM mode (see Appendix)

SRM : More than 80 compound

Pesticide Name	RT (min)	Precursor Mass (m/z)	Product Mass (m/z)	Collision Energy (V)
Difluorobenzamid Degradation (Isocyanat)	6.93	152.93	90.01	20
Difluorobenzamid Degradation (Isocyanat)	6.93	152.93	125.01	20
Carbofuran 1	8.80	149.06	121.05	10
Carbofuran 1	8.80	164.08	149.07	10
Difluorobenzamid Degradation	8.62	141.00	63.11	25
Difluorobenzamid Degradation	8.62	141.00	113.09	15
Biphenyl-d10, ISTD	9.28	160.00	160.16	10
Biphenyl	9.28	154.08	153.08	15
Biphenyl	9.28	153.08	152.08	15
Carbofuran-3-hydroxy 1	10.43	137.05	81.01	18
Carbofuran-3-hydroxy 1	10.43	180.05	137.01	15
Tetrahydropthalimid	10.84	151.04	79.01	25
Tetrahydropthalimid	10.84	151.04	122.09	10
O-Phenylphenol	11.00	170.07	141.06	20
O-Phenylphenol	11.00	170.07	115.05	20
Molinate	11.10	187.10	126.07	10
Molinate	11.10	126.07	98.05	5
Chlorfenprop methyl	11.59	196.00	165.00	10
Chlorfenprop methyl	11.59	165.00	137.00	10
Fenobucarb	11.20	121.07	77.05	15
Fenobucarb	11.20	150.09	121.07	10
Propachlor	11.76	176.06	120.04	10
Propachlor	11.76	120.04	92.03	10
Propachlor	11.76	169.06	120.04	10
Propachlor	11.76	196.07	120.04	10
Cyclone	11.98	154.10	83.05	10
Cycloate	11.98	215.13	154.10	5
Diphenylamin	11.49	169.01	168.09	20
Diphenylamin	11.49	169.01	167.09	20
Chloropropham	12.26	213.06	127.03	15
Chloropropham	12.26	213.06	171.04	10
Phosmet-oxon	12.09	160.00	132.96	15
Phosmet-oxon	12.09	104.00	75.88	10
Phosmet-oxon	12.09	160.00	76.96	20
Prometon	13.10	225.16	183.13	10
Prometon	13.10	225.16	210.15	10
Carbofuran 2	13.13	149.06	121.05	10
Carbofuran 2	13.13	164.08	149.07	10
Profluralin	13.22	318.10	199.06	15
Profluralin	13.22	330.23	252.45	25
Swep	13.46	187.05	123.95	18
Swep	13.46	219.11	174.02	15
Trietzazine	13.48	229.14	200.14	15
Trietzazine	13.48	214.14	186.10	15
Dimethipin	13.53	117.98	57.97	10

Pesticide Name	RT (min)	Precursor Mass (m/z)	Product Mass (m/z)	Collision Energy (V)
Dimethipin	13.53	210.10	76.02	10
Terbutylazin	12.97	214.10	132.06	10
Terbutylazin	12.97	214.10	104.05	10
Propyzamid	13.04	173.01	145.01	15
Propyzamid	13.04	173.01	109.01	18
Propyzamid	13.04	175.02	147.01	15
Propyzamid	13.04	254.02	226.02	15
Isocarbamide	13.67	142.03	70.01	15
Isocarbamide	13.67	142.03	113.01	10
Dinosob	13.92	211.13	116.99	15
Dinosob	13.92	211.13	163.11	10
Terbazil	13.42	161.05	88.03	15
Terbazil	13.42	160.05	76.02	15
Beomoculen	14.37	358.79	242.85	15
Beomoculen	14.37	356.93	241.24	15
Dimesethanamid	14.60	230.06	154.04	10
Dimesethanamid	14.60	232.06	154.04	10
Dimethachlor	14.61	197.08	148.06	10
Dimethachlor	14.61	199.08	148.06	10
Acetochlor	14.65	174.11	146.15	15
Acetochlor	14.65	223.19	147.17	10
Desmetryn	14.68	213.11	171.08	10
Flurprimidol	14.77	269.12	106.98	20
Alachlor	14.26	188.10	160.07	10
Alachlor	14.26	188.10	130.12	25
Alachlor	14.26	237.14	160.15	10
Metribuzin	14.14	198.08	82.03	20
Metribuzin	14.14	198.08	89.04	16
Propanil	15.00	217.01	161.00	10
Propanil	15.00	219.01	163.00	10
Fipronildesulfanyl	14.15	333.00	231.20	20
Fipronildesulfanyl	14.15	333.00	281.30	20
Carbofuran-3-hydroxy 2	15.02	137.05	81.01	18
Carbofuran-3-hydroxy 2	15.02	180.05	137.01	15
Prometryn	14.49	241.14	184.10	15
Prometryn	14.49	226.13	184.10	12
Tridiphan	15.18	186.94	158.94	15
Tridiphan	15.18	219.09	184.09	20
Ethofumesat	14.80	206.82	180.86	10
Ethofumesat	14.80	285.75	206.82	12
Pentanochlor	15.73	141.05	106.05	15
Pentanochlor	15.73	239.05	141.05	15
Chloropyrifos	15.78	257.97	165.98	20
Chloropyrifos	15.78	314.05	258.18	15
Bromacil	15.03	206.01	188.01	15
Bromacil	15.03	207.01	190.01	15

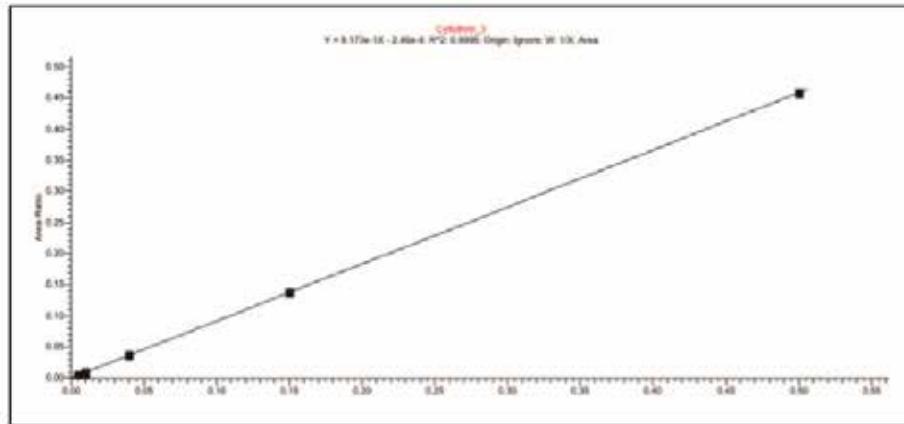
Pesticide Name	RT (min)	Precursor Mass (m/z)	Product Mass (m/z)	Collision Energy (V)
Anthrachinon	15.44	207.97	151.99	20
Anthrachinon	15.44	180.04	152.05	15
Anthrachinon	15.44	207.97	180.10	10
Nithrothal isopropyl	16.09	236.08	194.07	10
Nithrothal isopropyl	16.09	236.08	148.05	20
Triadimenfon	15.41	208.07	181.06	10
Triadimenfon	15.41	210.07	183.06	10
Tiocarbazil	16.15	156.08	100.05	8
Tiocarbazil	16.15	279.10	156.07	6
Tetraconazol	15.39	336.02	218.01	20
Tetraconazol	15.39	338.02	220.01	20
Butralin	15.54	266.14	220.11	15
Butralin	15.54	265.14	190.10	15
Dicaphthon	15.44	262.00	262.00	9
Crufomat	16.30	256.20	226.15	25
Crufomat	16.30	276.20	182.09	10
Allethrin	16.17	123.07	80.98	10
Allethrin	16.17	136.04	92.98	10
Dinobuton	16.89	163.06	116.04	15
Dinobuton	16.89	211.07	117.04	18
Pencosazol	16.89	243.06	157.04	25
Pencosazol	16.89	245.06	192.04	15
Pyrifenoxy 1	16.17	262.03	192.02	20
Pyrifenoxy 1	16.17	262.03	200.02	20
Pyrifenoxy 2	16.81	262.03	192.02	20
Pyrifenoxy 2	16.81	262.03	200.02	20
Famophos (Famphur)	20.16	218.07	108.94	15
Famophos (Famphur)	20.16	218.07	126.95	20
Iprodion Degradation	18.63	166.87	123.99	20
Iprodion Degradation	18.63	166.87	159.02	15
Iprodion Degradation	18.63	243.94	187.02	10
Iprodion	20.57	314.06	245.25	15
Iprodion	20.57	166.99	123.87	20
Iprodion	20.57	316.00	247.35	15
Iprodion	20.57	316.00	273.11	10
Propiconazol 1	19.38	259.02	173.02	20
Propiconazol 1	19.38	172.94	144.91	15
Propiconazol 2	19.54	259.02	173.02	20
Propiconazol 2	19.54	172.94	144.91	15
Pyrafenof-ethyl	20.30	412.02	349.02	15
Pyrafenof-ethyl	20.30	349.02	307.02	15
Cledinafop-propargyl	20.36	349.05	266.04	15
Cledinafop-propargyl	20.36	349.05	238.04	15
Lenail	20.70	153.05	136.06	15

Pesticide Name	RT (min)	Precursor Mass (m/z)	Product Mass (m/z)	Collision Energy (V)
Pacllobutrazole	17.75	238.11	127.06	15
Chlormethicost	17.75	206.06	147.98	15
Chlornethicost	17.75	234.08	206.06	10
Napropamide	18.07	271.16	128.07	5
Napropamide	18.07	128.07	72.04	10
Eflutriafol	18.11	219.07	123.04	15
Eflutriafol	18.11	123.04	75.03	15
Fluroditen	18.14	190.02	126.01	10
Fluroditen	18.14	190.02	146.01	5
Bisphenol A	18.17	213.14	119.06	15
Bisphenol A	18.17	213.14	164.99	20
Bisphenol A	18.17	228.15	213.07	10
Chlortenson ISTD	18.20	302.00	110.90	20
Hexaconazol	18.22	214.08	159.07	20
Hexaconazol	18.22	214.08	151.98	25
Imazalil	18.24	172.96	144.96	15
Imazalil	18.24	172.96	108.95	25
Isoprothiolan	18.24	203.99	117.95	7
Isoprothiolan	18.24	203.99	84.90	25
Isoprothiolan	18.24	290.06	118.03	15
Flamprop-methyl	18.39	230.05	170.04	10
Flamprop-methyl	18.39	276.06	105.02	10
Kresoxim-methyl	18.48	206.10	131.09	15
Kresoxim-methyl	18.48	206.10	116.01	10
Buprofezin	18.51	175.08	116.96	20
Buprofezin	18.51	175.08	131.99	15
Buprofezin	18.51	249.16	105.93	20
Pesticide Name	RT (min)	Precursor Mass (m/z)	Product Mass (m/z)	Collision Energy (V)
Azinphosmethyl	22.95	160.00	132.00	10
Azinphosmethyl	22.95	160.00	104.64	10
Pyriproxyfen	23.06	136.00	77.92	20
Fenamiro	23.55	251.02	159.01	15
Fenamiro	23.55	330.03	139.01	10
Pyridaben	24.50	364.14	309.12	5
Pyridaben	24.50	309.12	147.06	15
Fluquinconazol	24.59	340.01	298.01	22
Fluquinconazol	24.59	342.01	300.01	22
Etofenprox	26.05	163.09	107.06	16
Etofenprox	26.05	163.09	135.07	10
Etofenprox	26.05	376.14	135.02	30
Etofenprox	26.05	376.14	163.09	10
Silaffuolen	26.25	179.00	151.00	7
Difenaconazol 1	26.91	323.05	265.04	15
Difenaconazol 1	26.91	325.05	267.04	20
Difenaconazol 2	27.05	323.05	265.04	15
Difenaconazol 2	27.05	325.05	267.04	20
Indoxacarb	28.55	264.02	176.14	10
Indoxacarb	28.55	264.02	148.03	20
Indoxacarb	28.55	321.05	259.34	10

Your Scientific Specialist

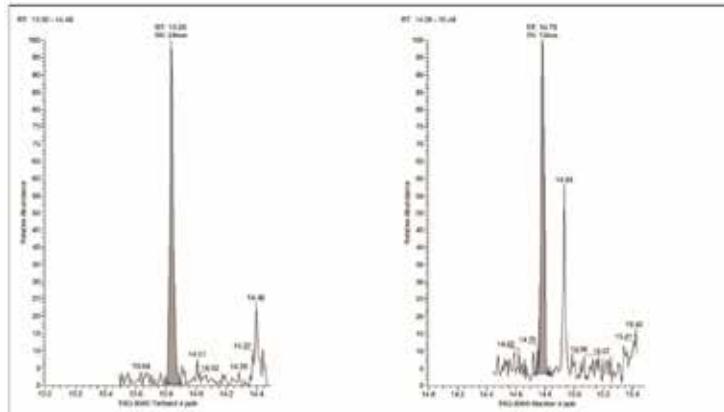
Calibration and Detection limit.

Calibration level : 0.004 µg/mL to 1.0 µg/mL (This range represents an analyte concentration of 0.01 to 2.5 mg/kg in the samples)

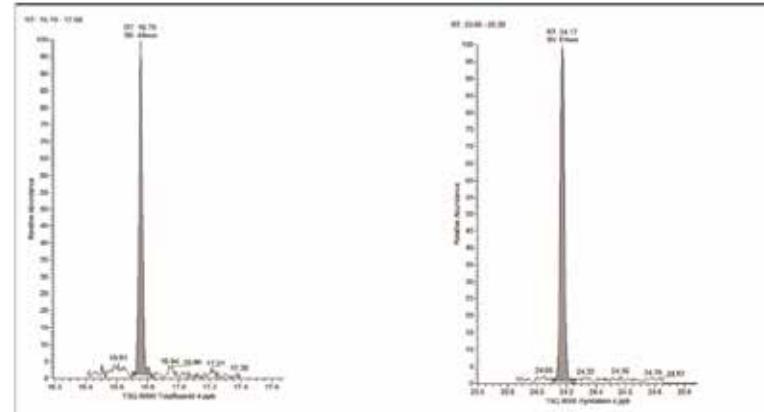


Pesticide	RT [min]	S/N @ 4 ppb
Terbacil	13:83	24
Alachlor	14:78	12
Tolyfluanid	16:75	44
Pyridaben	24:17	83

Sensitivity (LOD)



Terbacill



Sample Result.....

Sample Matrix	Pesticide Residues Found	Concentration (mg/kg)
Dried Herbs	o-Phenylphenol	0.017
Dried Herbs	Tebuconazol	0.023
Dried Fruit	Diflubenzuron	0.049
Dried Fruit	Myclobutanil	0.023
Dried Fruit	Propargit	0.479
Dried Fruit	Tebuconazol	0.081
Dried Fruit	Difenconazol	0.013
Dried Herbs	Picoxystrobin	0.228
Dried Herbs	Picoxystrobin	0.233
Dried Herbs	o-Phenylphenol	0.011
Herbal Tea	o-Phenylphenol	0.014
Herbal Tea	o-Phenylphenol	0.011
Herbal Tea	Terbutylazin	0.016



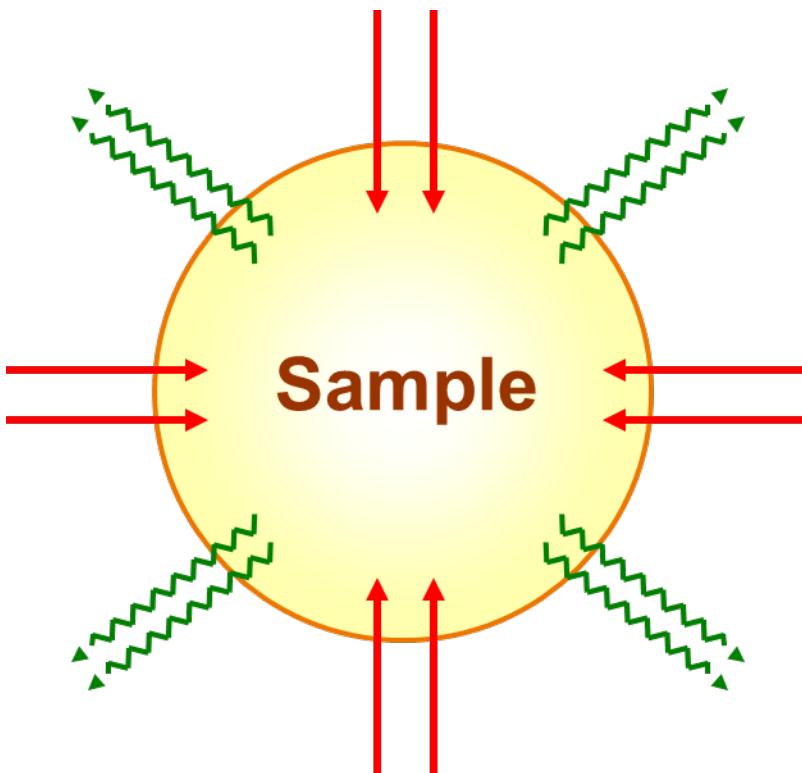
Application note 52291

Your Scientific **Specialist**

- PY-GCMS



Information from polymeric Materials by Heating

**Information**

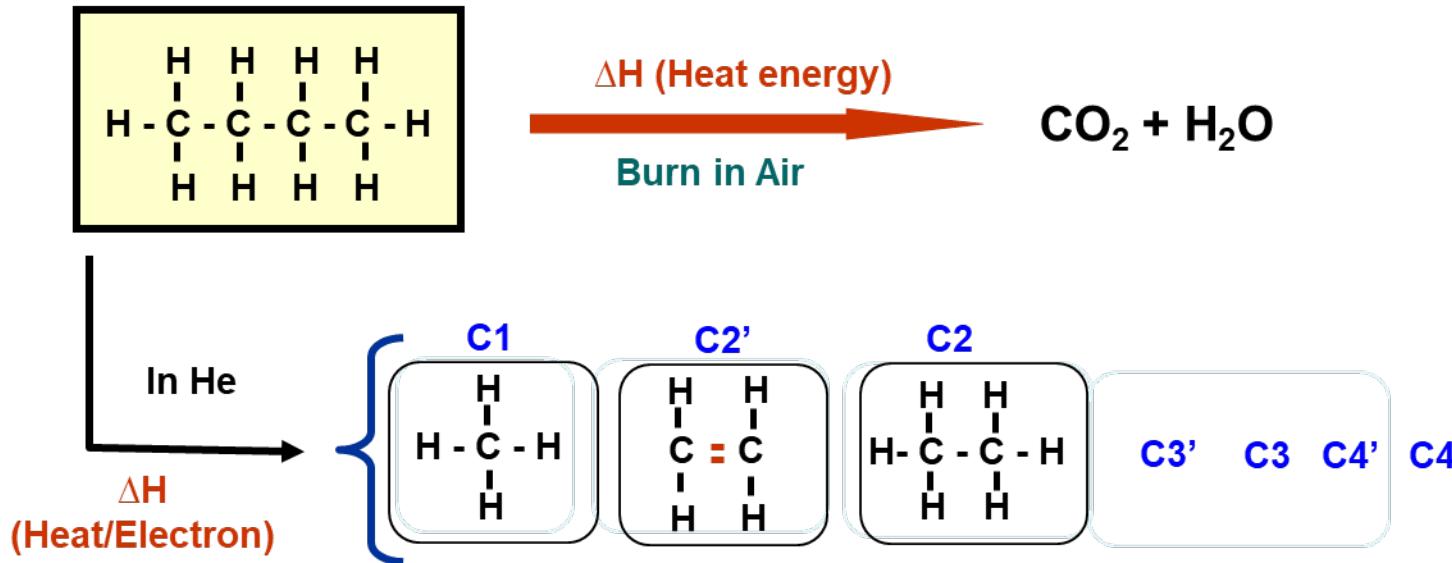
- Weight loss: **TGA**
- Enthalpy change: **DTA, DSC**
- Mechanical change: **TMA, Dilatometry**
- Evolved gas

volume: **EGA (volume of gas)**
qualification & quantification:

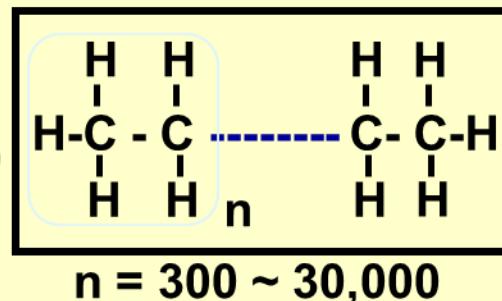
Py-GC/MS
TD-GC/MS
UV/Py-GC/MS
EGA/MS
Py/MS

Pyrolyzer

Pyrolysis of Polymeric materials and pyrolyzates



Polyethylene
(MW: 10,000 ~1,000,000)

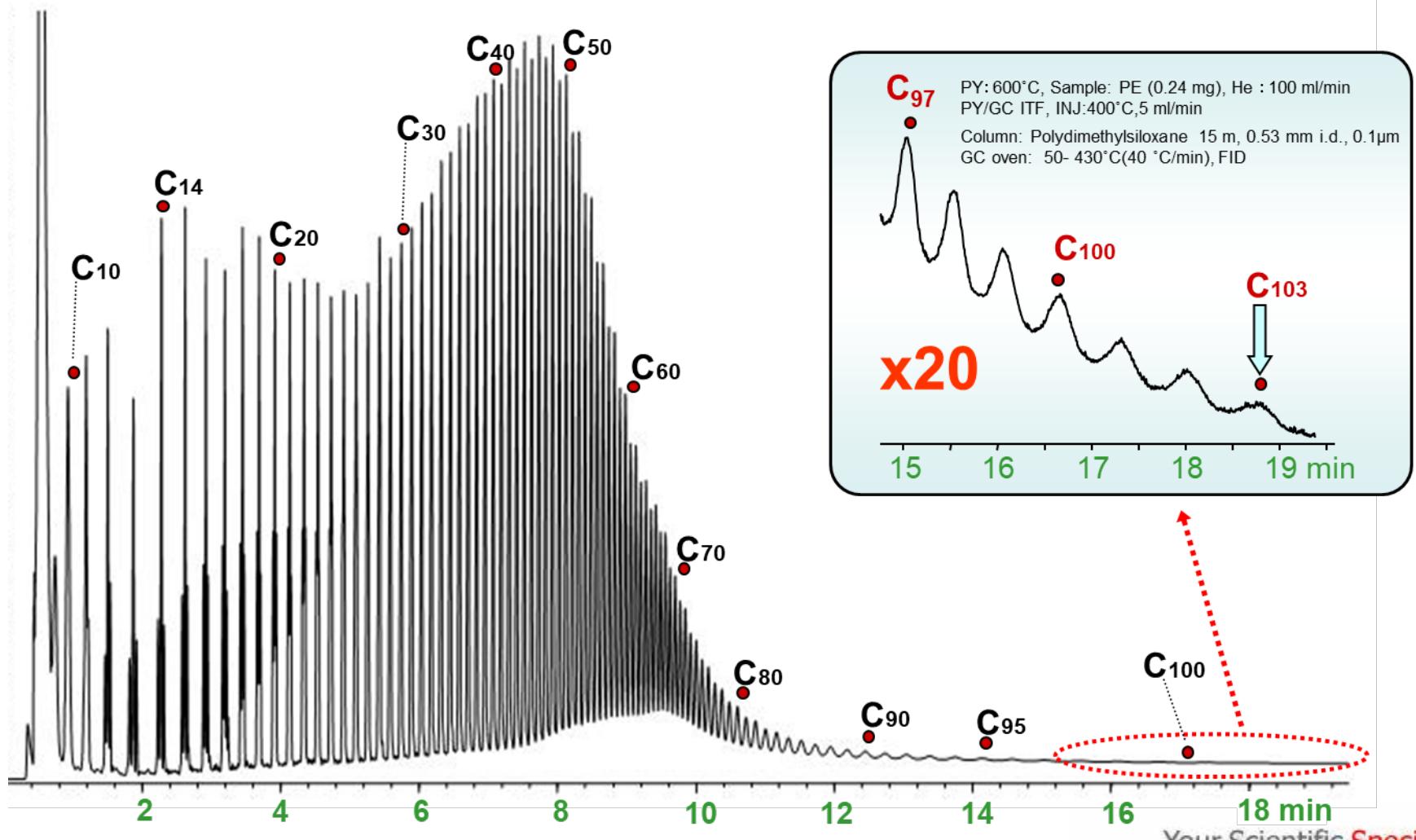


ΔH
(Heat/Energy) 

C1, C2', C2, C3', C3
C4', C4" ... Cx

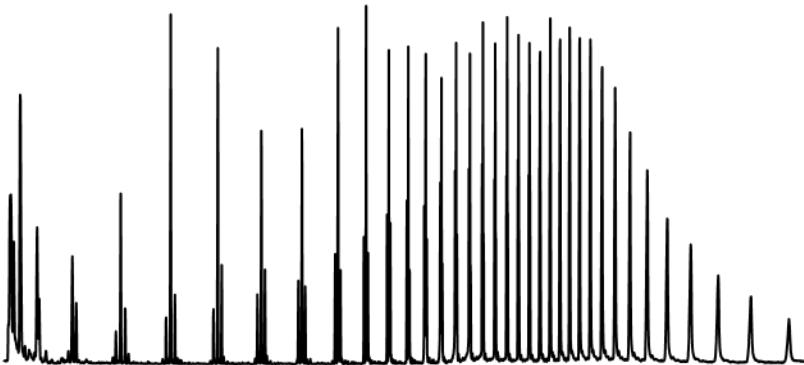
Pyrolyzer

Typical pyrogram of polyethylene at 600°C

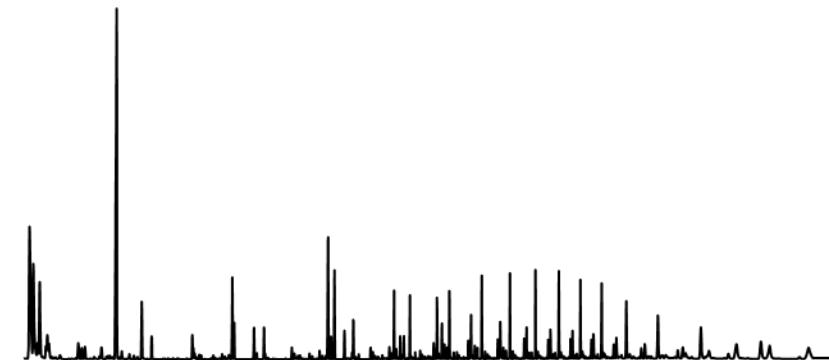


Typical pyrograms

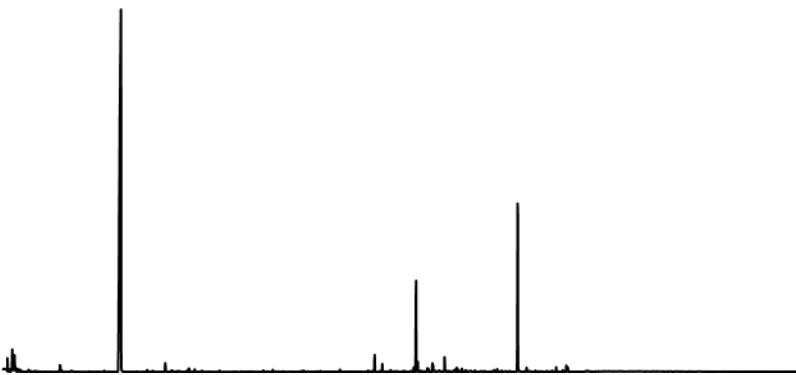
Polyethylene



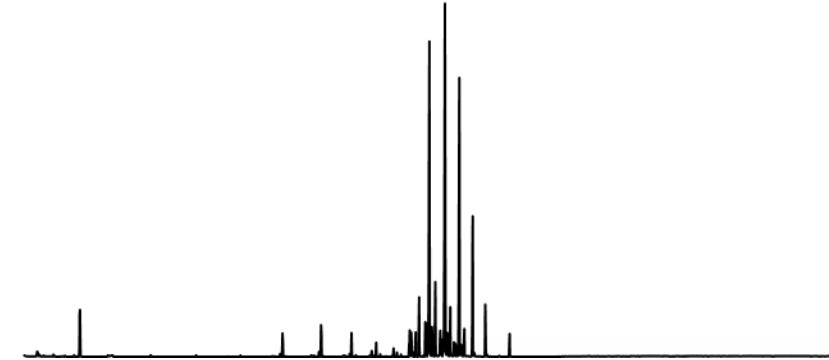
Polypropylene



Polystyrene



Higher methacrylate copolymer

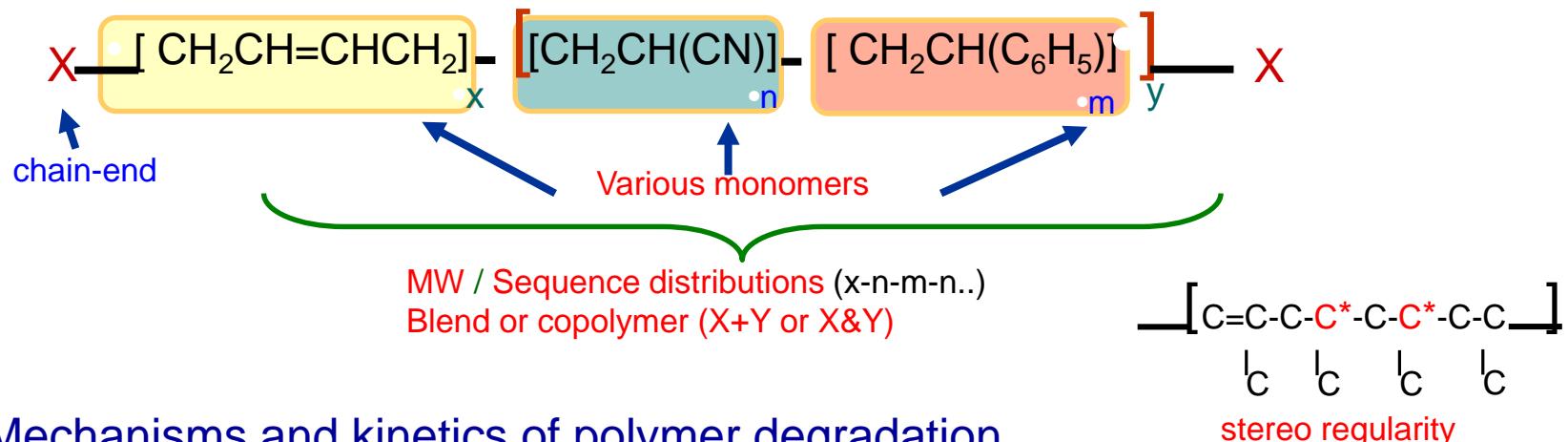


Characterization of Polymers by PY-GC/MS

A: Identification of polymeric materials

Unknown materials (PP/ PVC/ SBR?)

B: Structural characterization of polymers



C: Mechanisms and kinetics of polymer degradation

D: Qualitative and quantitative analysis of additives

ສກາວະເຄົ່ອງ GCMS

- Injector
 - Temperature 300 °C
 - Split 200:1
 - Carrier gas flow 1.0 ml/min
- Oven
 - Initial 70 °C hold 1min ramp 1 ; 10 °C/min to 320 °C hold 8 min.
- MS
 - Temperature 250 °C
 - Scan 35-550 amu.

ສກາວະເຄົ່ອງ Pyrolyzer

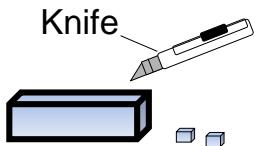
- Single-Shot Analysis
- Furnace Temperature 600 °C
- Interface Temperature 300 °C



Sample cup

ขั้นตอนการเตรียมตัวอย่าง

Step 1



Step 2

Place a sample
in the sample cup

0.1- 0.5mg

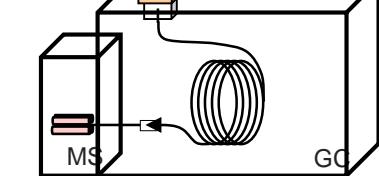


No solvent extraction

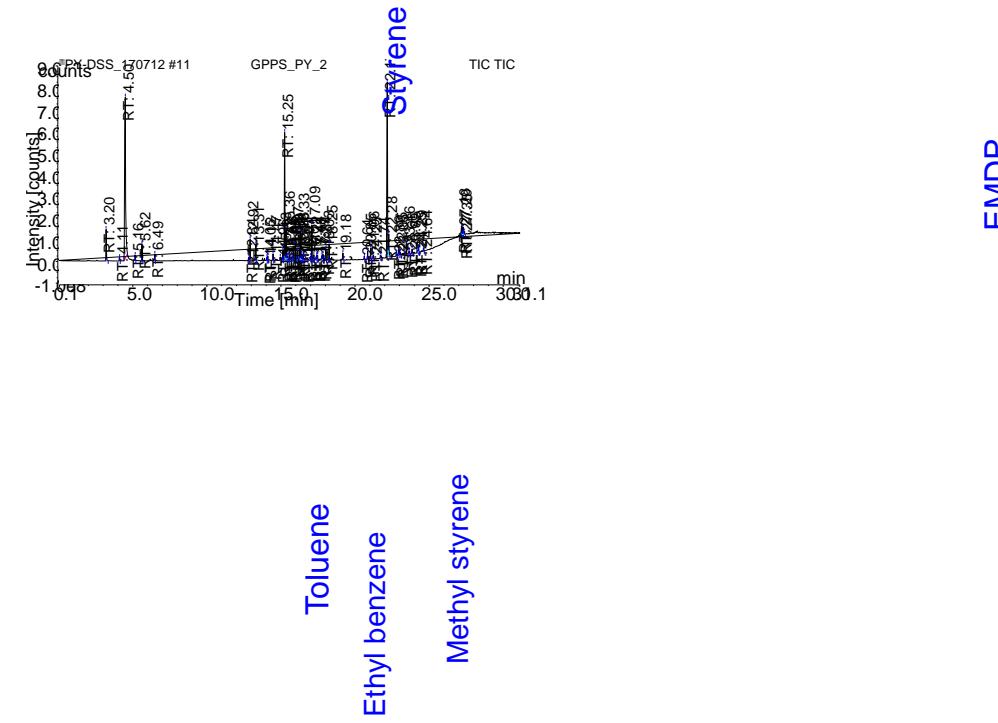
Step 3

Pyrolyzer

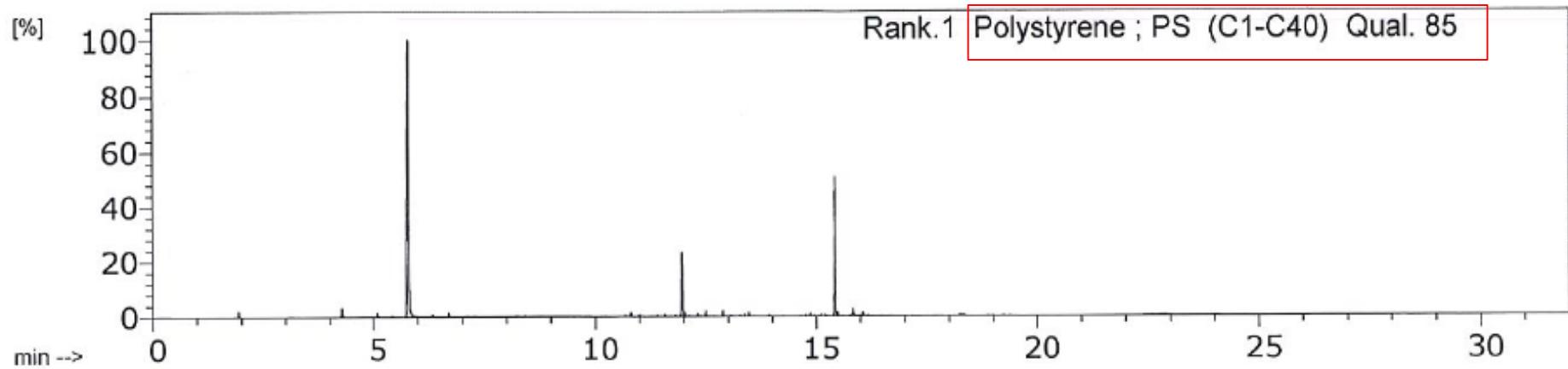
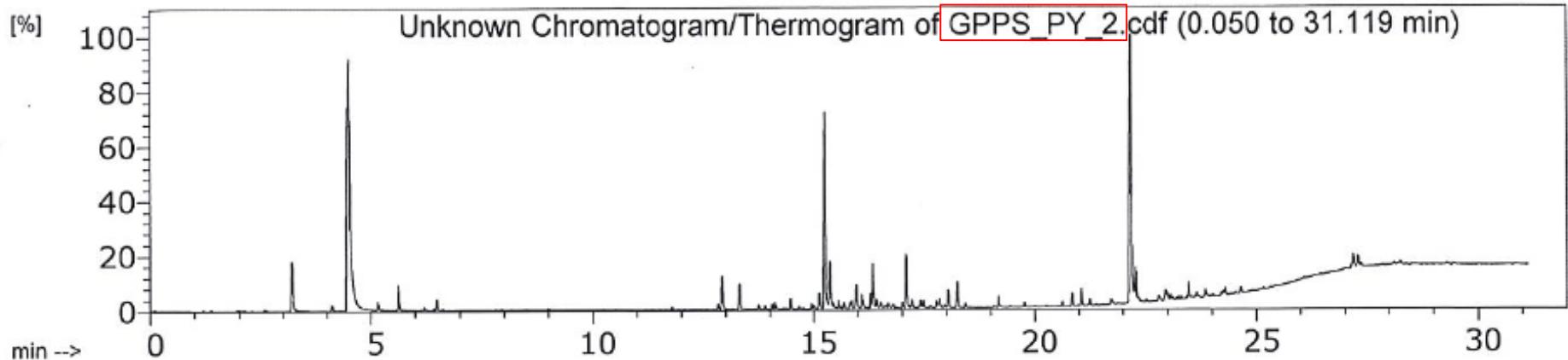
Sample cup



ผลการวิเคราะห์ตัวอย่างที่ 1



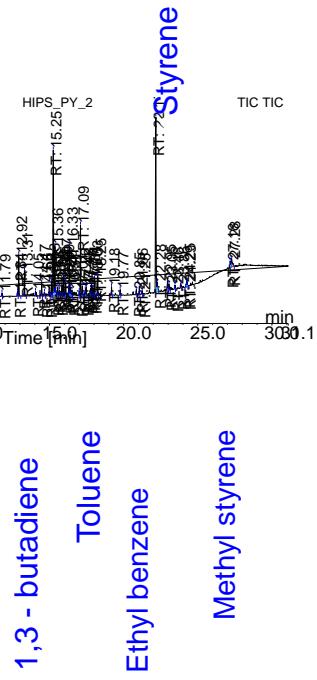
1/40 «Í ¿áÇAF-Search



Rank.2 : Styrene-butadiene copolymer ABA block, 85% styrene (C1-C40) Qual. 85

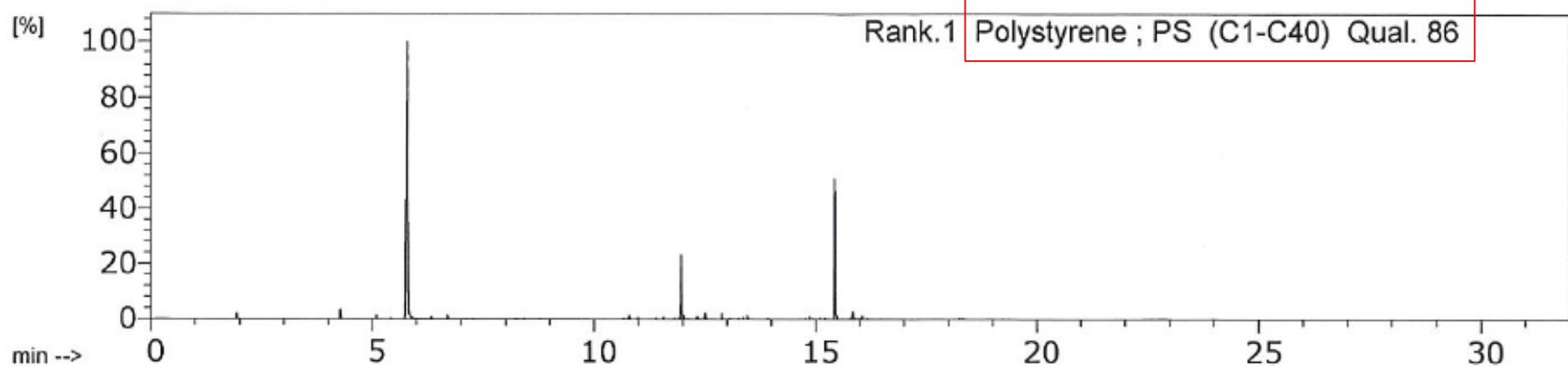
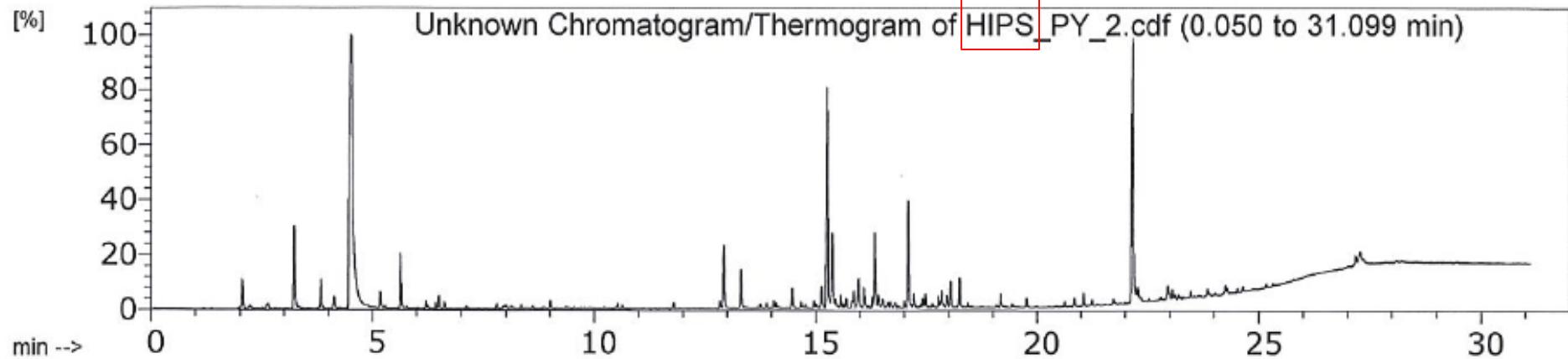
Rank.3 : Acrylonitrile-Butadiene-Styrene copolymer ; ABS (C1-C40) Qual.84

ผลการวิเคราะห์ตัวอย่างที่ 2



(2,3-diphenylcyclopropyl) methyl phenyl sulfoxide, *trans*

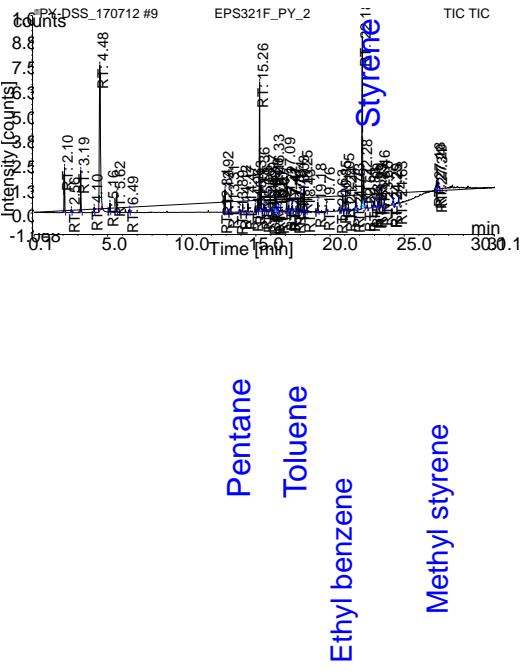
140 «ÍçáÇÁF-Search



Rank.2 : Acrylonitrile-Butadiene-Styrene copolymer ; ABS (C1-C40) Qual.86

Rank.3 : Styrene-butadiene copolymer ABA block, 85% styrene (C1-C40) Qual. 86

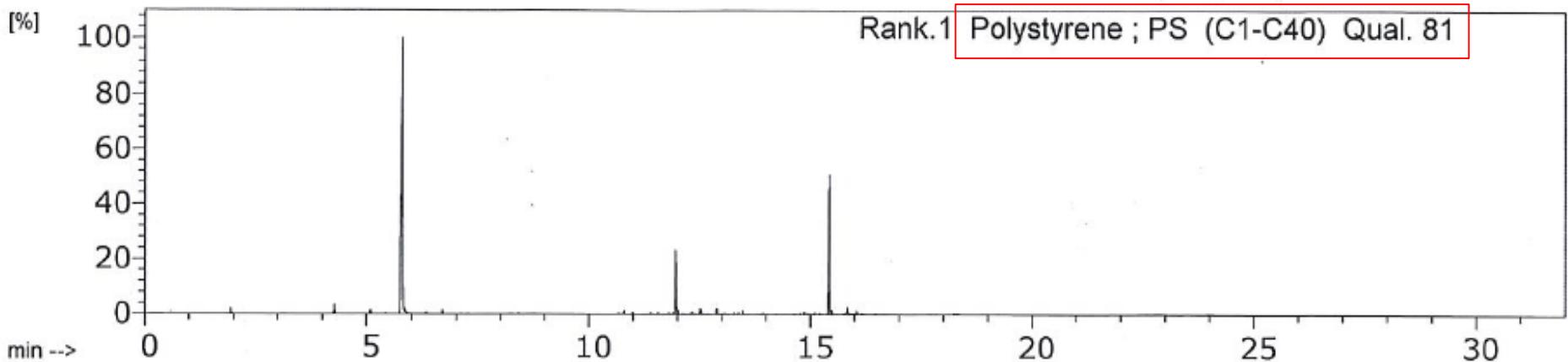
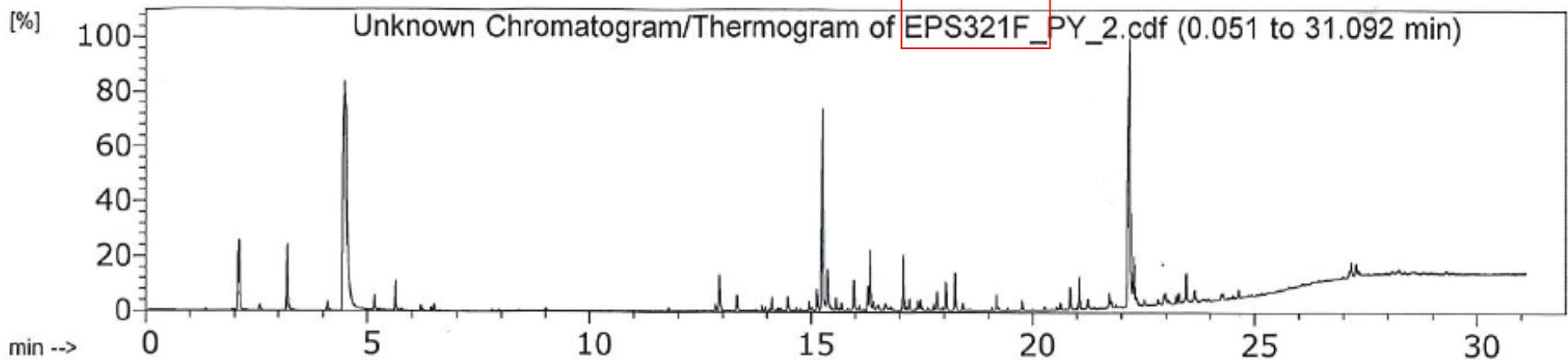
ผลการวิเคราะห์ตัวอย่างที่ 3



EMDP

(2,3-diphenylcyclopropyl) methyl phenyl sulfoxide · trans

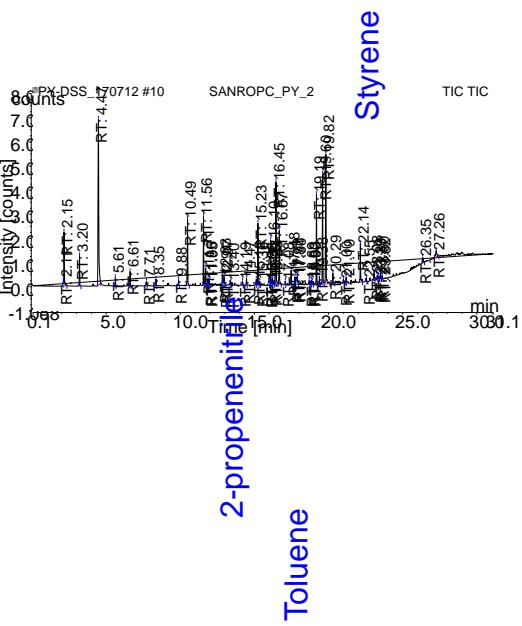
100% F-Search



Rank.2 : Acrylonitrile-Butadiene-Styrene copolymer ; ABS (C1-C40) Qual.81

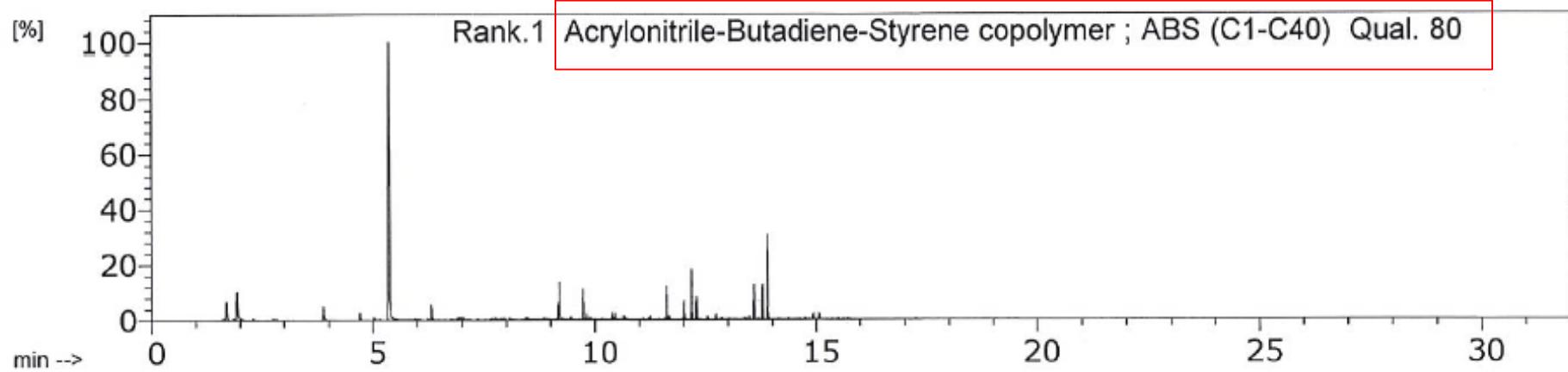
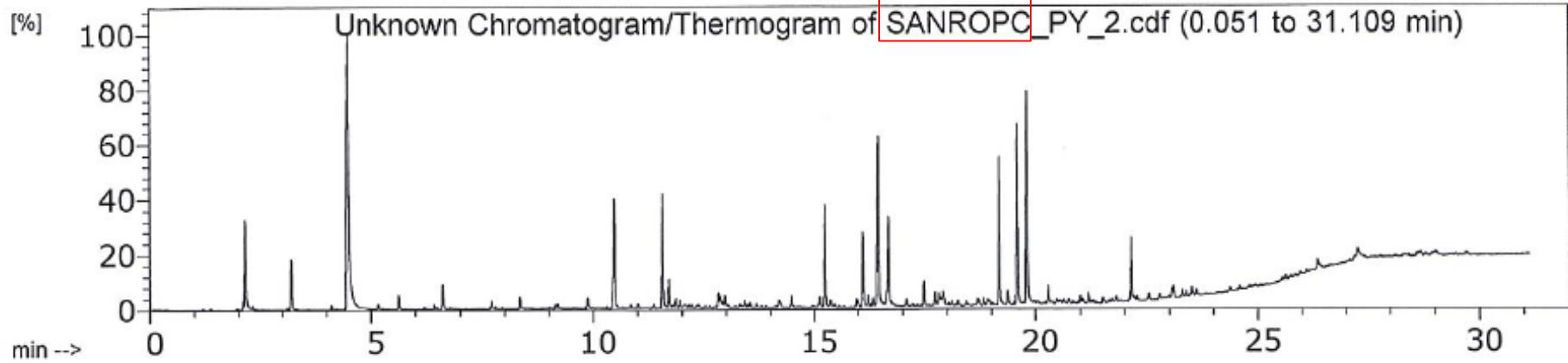
Rank.3 : Styrene-butadiene copolymer ABA block, 85% styrene (C1-C40) Qual. 81

ผลการวิเคราะห์ตัวอย่างที่ 4



EMDP

140 «Í ão F-Search



Rank.2 : Acrylonitrile-Butadiene-Styrene copolymer ; ABS (C1-C40) Qual.79

Rank.3 : Acrylonitrile styrene copolymer ; AS (C1-C40) Qual.76

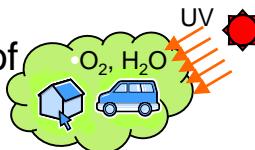
1: Characterization of polymers



2: Quality control



3: Degradation/life evaluation of polymeric materials



4: Recycling of polymeric materials, biomass utilization



5: Organic geochemistry and soil chemistry



6: Clinical science, pathology



7: Biochemistry, microbiology



8: Coal liquefaction, energy conservation



9: Forensic science



10: Wood science, pulp industry



11: Tobacco smoke, toxicology



12: Extraterrestrial science



13: Environmental science



● **Analysis PAHs in extender oils**



Topics to be discussed

- Introduction PAHs
- Sample Preparation
- GCMSMS method
- Analysis PAHs
- LOD&LOQ
- Example of sample result
- Comment

Introduction

- Polycyclic aromatic hydrocarbons (PAHs) in extender oils and tyres are produced using extender oils that may contain PAHs not added intentionally.
- PAHs are considered as toxic substances classified according to Directive 67/548/EEC as carcinogenic, mutagenic and toxic for reproduction.

Scope for analysis.

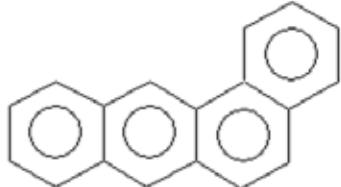
- EU standard specifies a procedure for determination of benzo(a)pyrene and sum of the eight individual polycyclic aromatic hydrocarbons in extender oils. ***listed in Table 1***
- Sample Preparation Method : BS EN 16143:2013

Name of PAH	Abbreviation	CAS Registry number
Benzo(a)pyrene	BaP	50-32-8
Benzo(e)pyrene	BeP	192-97-2
Benzo(a)anthracene	BaA	56-55-3
Chrysene	CHR	218-01-9
Benzo(b)fluoranthene	BbFA	205-99-2
Benzo(j)fluoranthene	BjFA	205-82-3
Benzo(k)fluoranthene	BkFA	207-08-9
Dibenzo(a,h)anthracene	DBahA	53-70-3

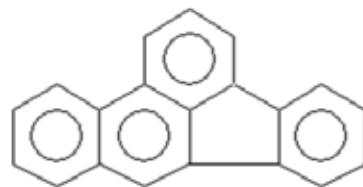
Table 1- List of individual PAHs in extender oils

PAHs...

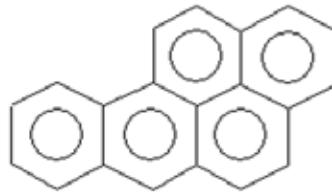
Consists of 8 natives of PAHs
MW range 228-278 amu (16PAHs could be up to 300+)



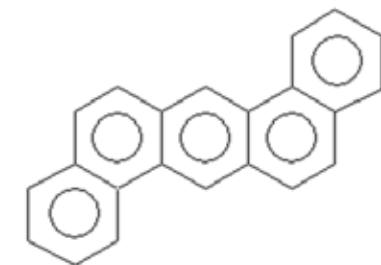
Benzo(a)anthracene



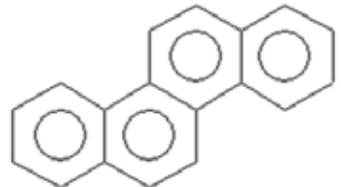
Benzo(b)fluoranthene



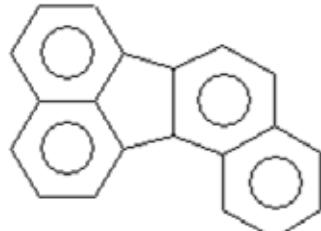
Benzo(a)pyrene



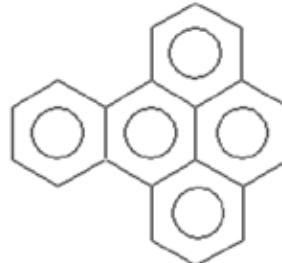
Dibenzo(a,h)anthracene



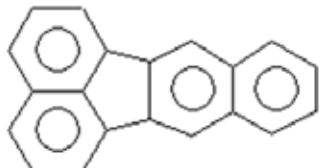
Chrysene



Benzo(j)fluoranthene



Benzo(e)pyrene



Benzo(k)fluoranthene

$C_{18}H_{12}$
MW. 228 g/mol

$C_{20}H_{12}$
MW. 252 g/mol

$C_{20}H_{12}$
MW. 252 g/mol

$C_{22}H_{14}$
MW. 278 g/mol

Sample Preparation Process

(1) Prepares sample solution

Weight Sample 70 ± 0.1 mg into Vol. flask 5 ml

Dissolve with 2 ml of n-Pentane and Spike internal Std. (deuterated IS)



Pack column

Extracting

Deactivate Silica gel by stirring with 7% (m/m) of water for 24 h.

(3) 1st sample extraction (8 Hours)

3.1 Mix deactivated silica (in 2) 5 g with n-Pentane

3.2 Load silica gel into chromatographic column (16 cm. L X 1 cm. ID)*

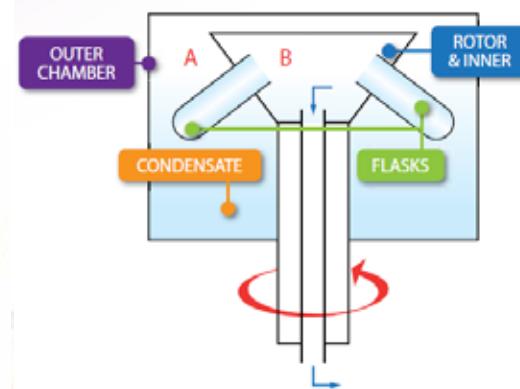
3.3 Flush silica gel with 10 ml n-Pentane through the column (discard)

3.4 Load sample (1) into column (before n-Pentane vanish form silica gel surface).

3.5 Rinse sample container with 2 ml n-Pentane. (not critical) and pour into column.

3.6 Elute sample by Cyclohexane 75 ml (several portion) and collect the eluents. **

3.7 Evaporate eluent (3.6) under 35 C till final volume 1ml.



*extended length of column to 25 cm. convenient for sample loading

** pressurized with N2 (1 bar est.) for faster elution

Sample Preparation Process

(4) Sample clean up (Sephadex LH20) (6 hours)

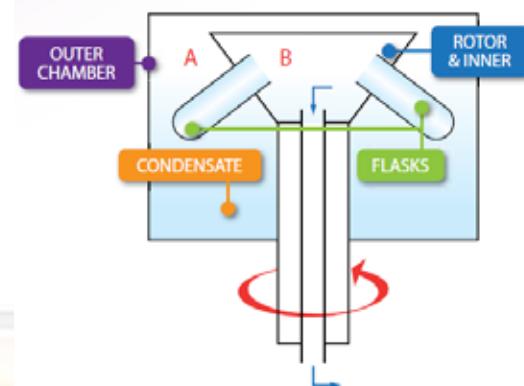
- 4.1 Mix 5 g. of Sephadex with IPA .. leave for overnight.
- 4.2 Load Sephadex into chromatographic column (12 cm L X 2.3 cm ID)
- 4.3 Add 1 ml IPA into (3.7) and load into column.
- 4.4 Rinse sample vessel with IPA (1 ml) and load into column.
- 4.5 Elute with IPA at 1 ml/min, Discard the first 24 ml eluent.
- 4.6 Collect eluent portion (@24-70 ml) in drying vessel
- 4.7 Evaporate eluent (4.6) under 35 C till nearly dry.
- 4.8 Add 2 ml Acetone and evaporate till dry.
- 4.9 Dissolve with CycloC6 and transfer into 1 ml Vol.Flask
- 4.10 Add injection standard (DE)* 0.2 ml and make up volume to 1 ml with CycloC6
- 4.11 Make up volume to 1 ml wth Cyclohexane.
- 4.12 Analyze with GCMSMS.



Fraction collecting



Dissolved Solution



*DE = Decafluorodiphenyl

Instrument Method

GC parameters

Parameter	Value
GC-column	60 m x 0.25 mm ID x 0.25 µm
Stationary phase	17% phenyl-methylpolysiloxane
Temperature program	Initial 90 °C hold 1min 20°C /min to 250 °C 4°C /min to 330 °C hold 10 min
Injection	PTV, Splitless
Injection temperature	275 °C
Injection Volume	1 µL
Carrier gas	He UHP grade 1.2 ml/min

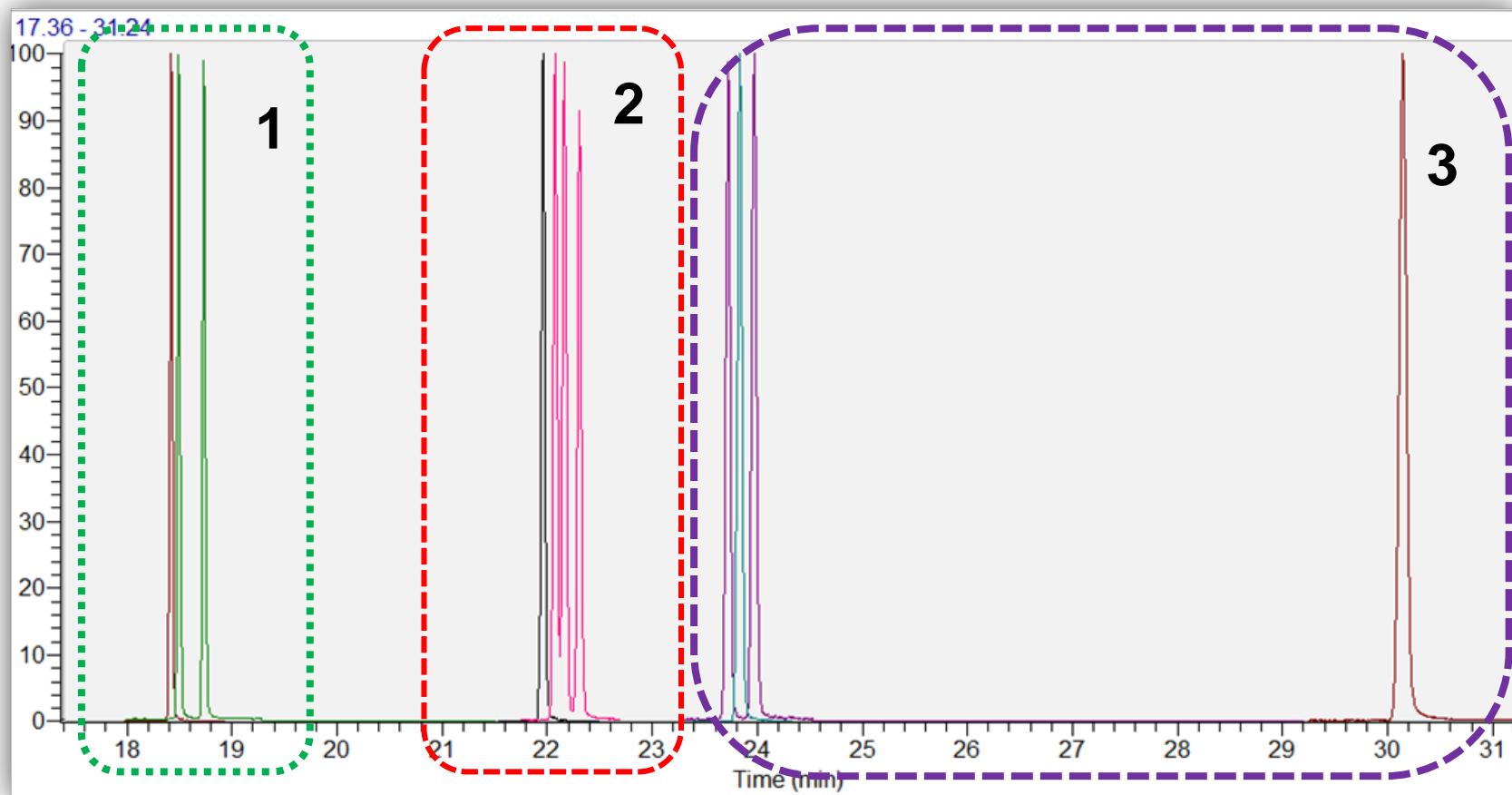
Instrument Method

- Mass Spectrometer : EI – Temp 250 C/ TL Temp 330/
- MSMS – SRM Q1 resolution 0.7 FWHM, Q3 Resolution 0.7 FWHM

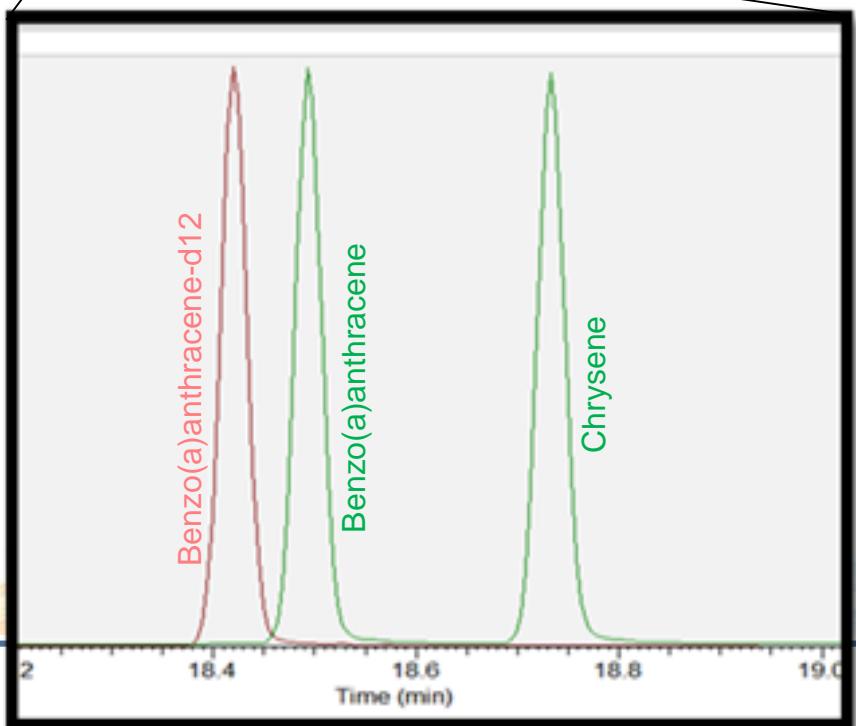
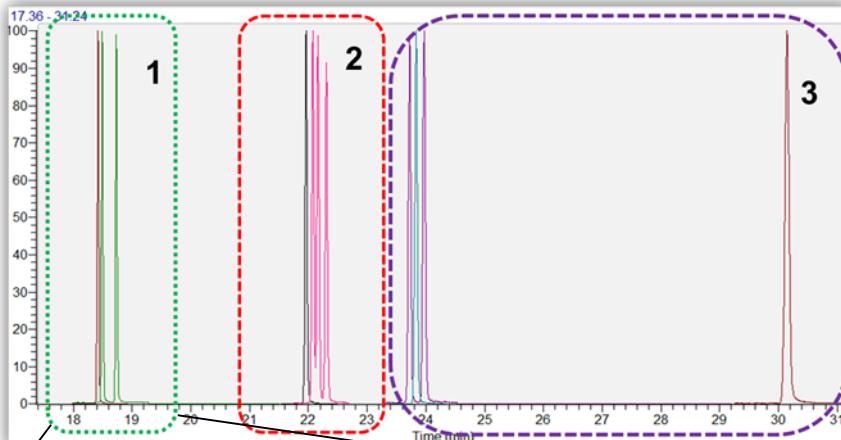
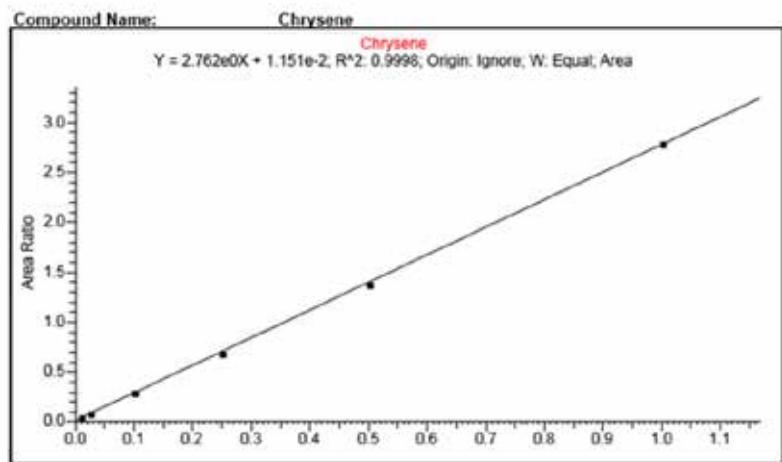
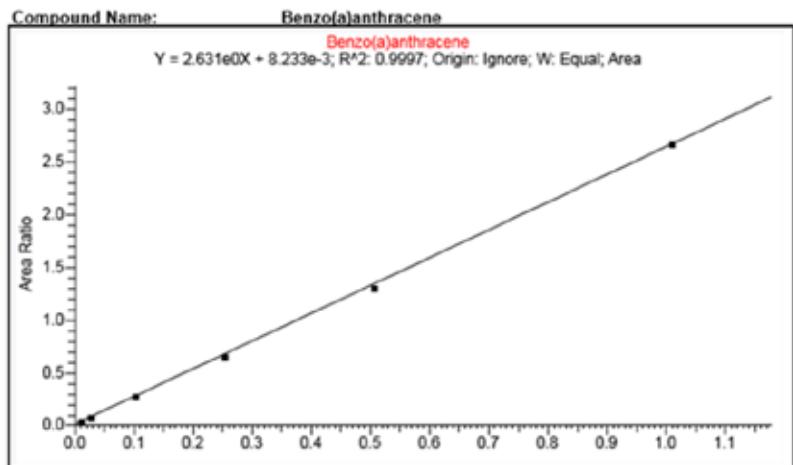
Component	RT	mass	product mass	Collision energy
Decfluorodiphynly	5.84	333.9	233.9	35
		333.9	264.9	25
Benzo(a)antracene-D12	18.46	240.1	212.1	25
		240.1	236	30
Benzo(a)antracene	18.53	228.1	202	25
		228.1	226	30
Chrysene	18.77	228.1	202	25
		228.1	226	30
Benzo(b)Fluoranthene-D12	22.02	264.1	236	30
		264.1	260	35
Benzo(b)fluoranthene	22.13	252.1	226	25
		252.1	250	30
Benzo(k)fluoranthene	22.22	252.1	226.1	25
		252.1	250	35
Benzo(j)fluoranthene	22.36	252.1	226	25
		252.1	250	30
Benzo(e)pyrene	23.78	252.1	226.1	30
		251.1	250	30
Benzo(a)pyrene-D12	23.89	264.2	236.1	30
		264.2	260	35
Benzo(a)pyrene	24.03	252.1	226.1	35
		251.1	250	30
Dibenzo(a,h)anthracene	30.23	278.1	276	35
		278.1	276.2	50

8 PAHs Standard

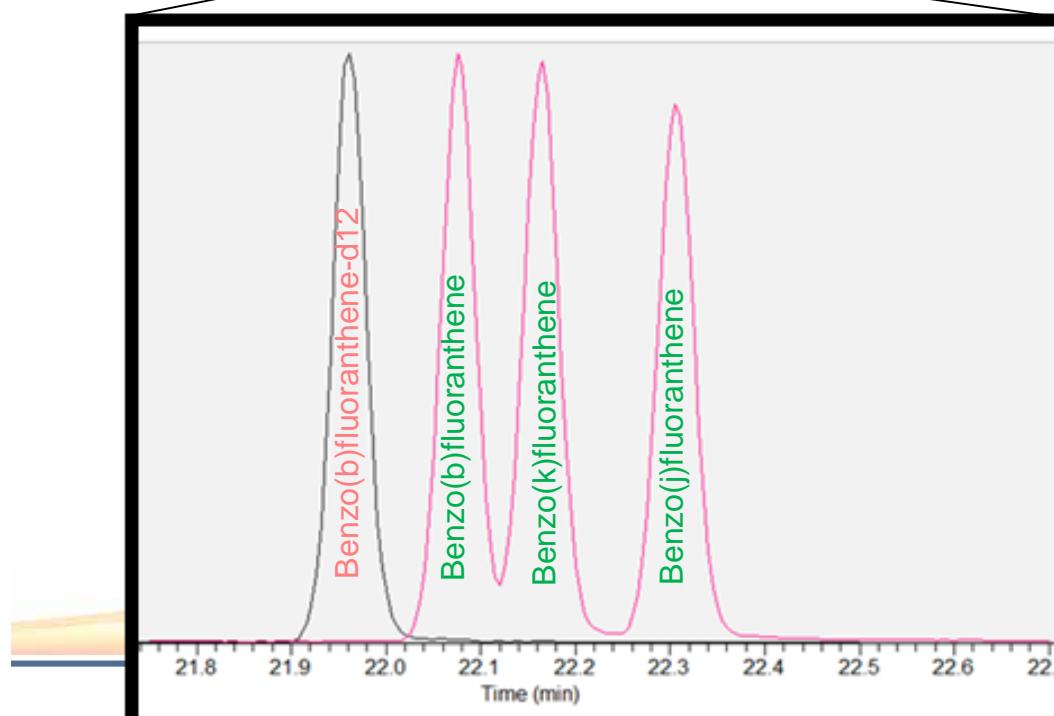
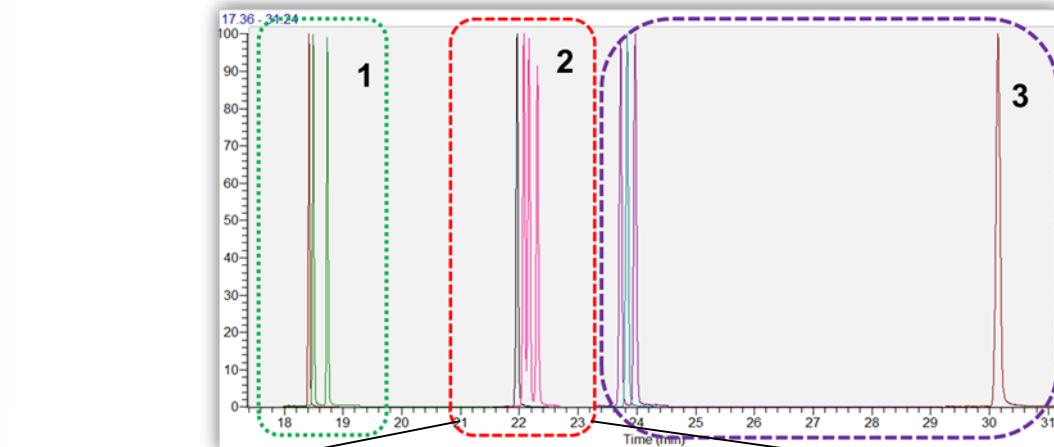
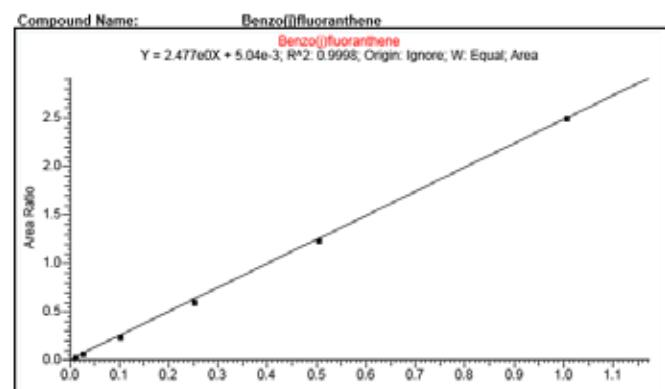
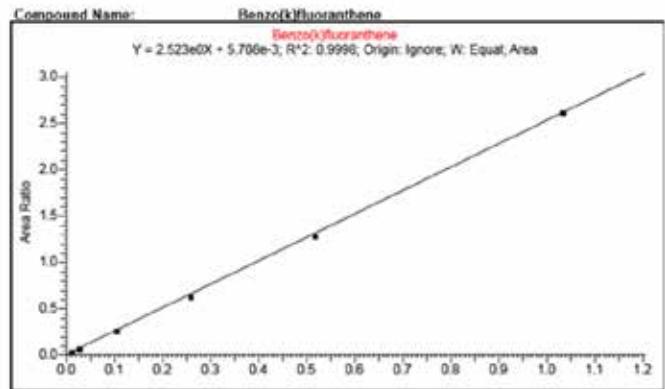
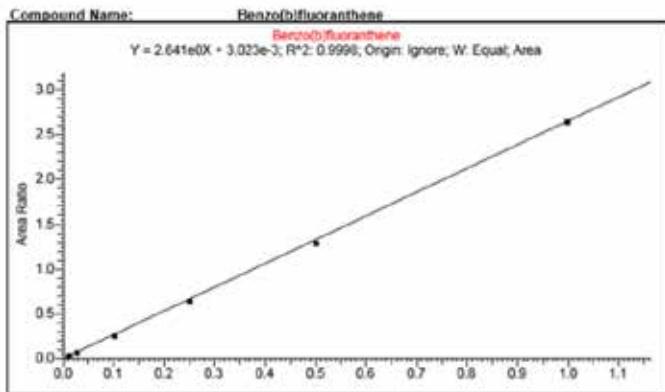
TIC



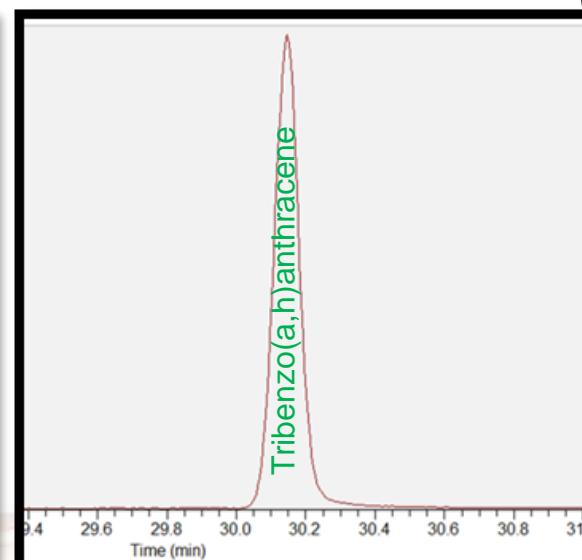
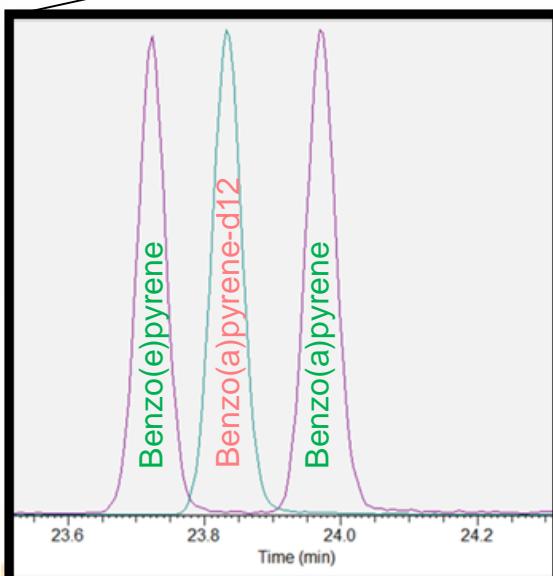
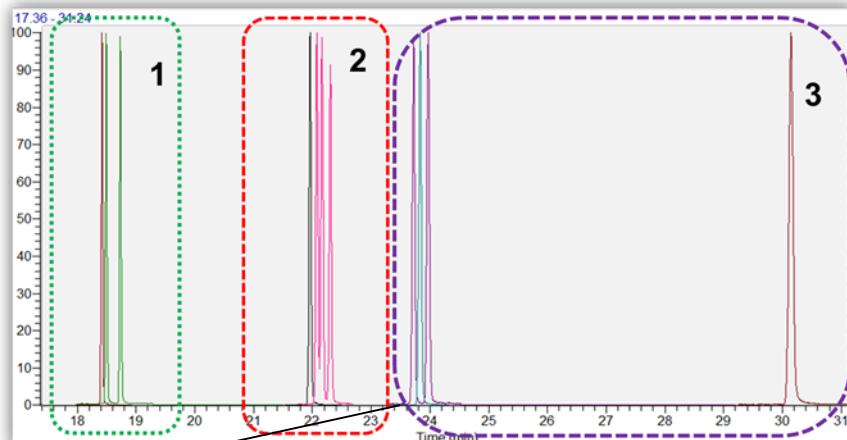
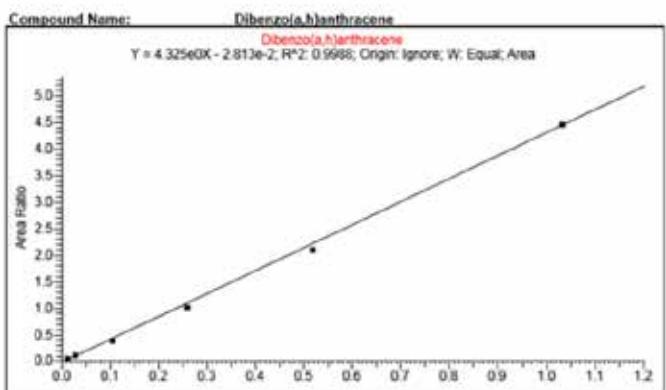
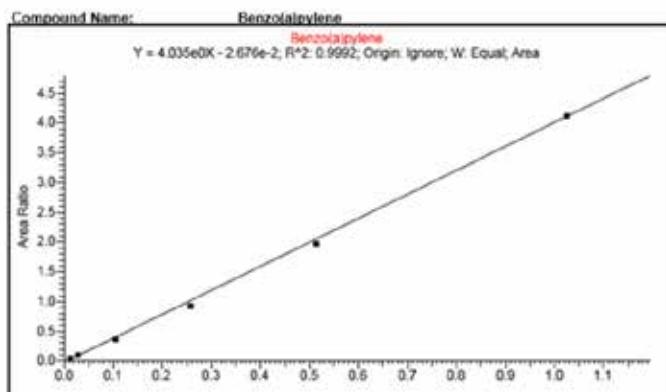
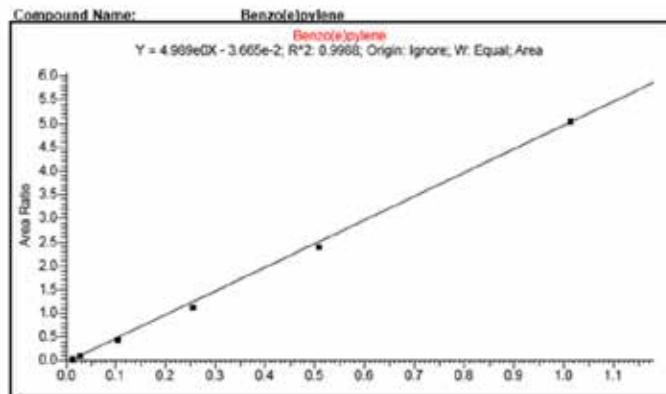
Chromatogram (1) –Standard 8 PAHs with 3 IS(d12)



Chromatogram (2) –Standard 8 PAHs with 3 IS(d12)



Chromatogram (3) –Standard 8 PAHs with 3 IS(d12)



LOD/LOQ

- Calculated from 10 replicate runs of TDAE sample (Treated Distillate Aromatic Extracted)

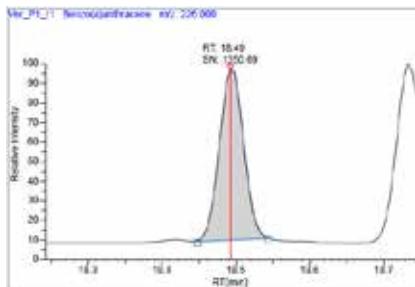
No.	PAHs (mg/kg)							
	Benzo(a)anthracene	Chrysene	Benzo(b)fluoranthene	Benzo(k)fluoranthene	Benzo(j)fluoranthene	Benzo(e)pylene	Benzo(a)pylene	Dibenzo(a,h)anthracene
1	0.226	0.370	0.198	0.186	0.103	-0.507	0.144	0.125
2	0.220	0.367	0.177	0.165	0.117	-0.510	0.130	0.148
3	0.222	0.361	0.184	0.182	0.127	-0.507	0.137	0.124
4	0.236	0.375	0.194	0.178	0.136	-0.511	0.147	0.149
5	0.221	0.372	0.204	0.168	0.118	-0.518	0.129	0.150
6	0.224	0.366	0.189	0.180	0.117	-0.510	0.129	0.142
7	0.236	0.363	0.192	0.194	0.123	-0.535	0.122	0.139
8	0.221	0.368	0.204	0.178	0.133	-0.509	0.126	0.135
9	0.247	0.369	0.181	0.166	0.118	-0.509	0.125	0.144
10	0.231	0.362	0.202	0.169	0.130	-0.507	0.115	0.147
SD	0.0089	0.0045	0.0097	0.0095	0.0097	0.0086	0.0098	0.0095
LOD	0.0267	0.0134	0.0291	0.0285	0.0291	0.0258	0.0294	0.0286
LOQ	0.0891	0.0447	0.0969	0.0951	0.0972	0.0860	0.0980	0.0955

8 compounds of PAHs have LOQ less than 0.1 mg/kg

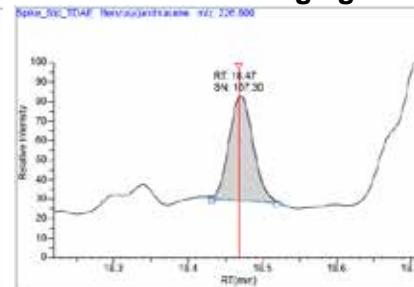
Peak Confirmation

Benzo(a)anthracene

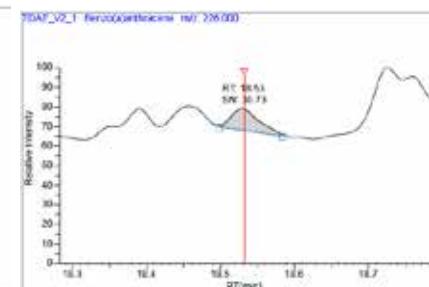
QC Check



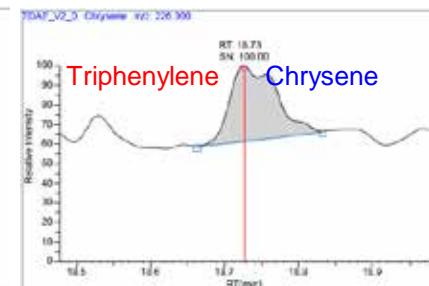
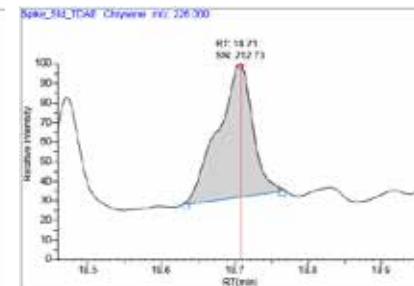
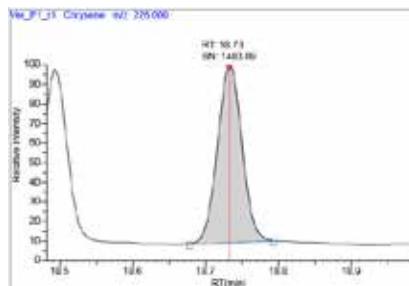
Sample spiked
3 ul of 0.5 mg/kg



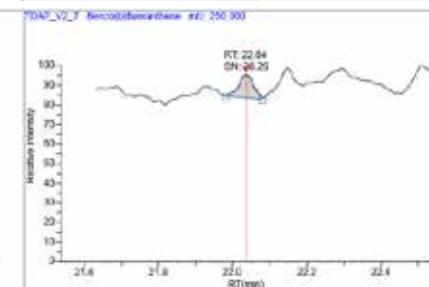
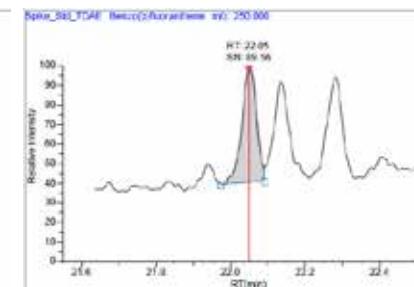
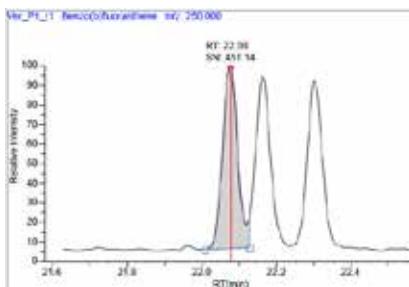
Sample(TDAE)



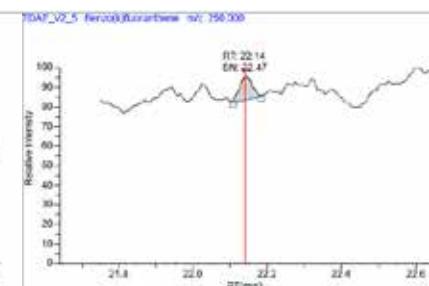
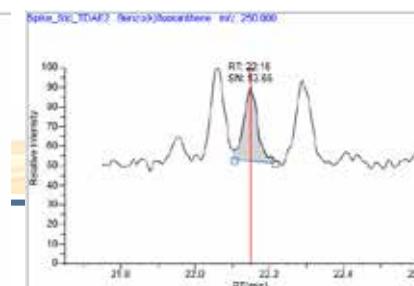
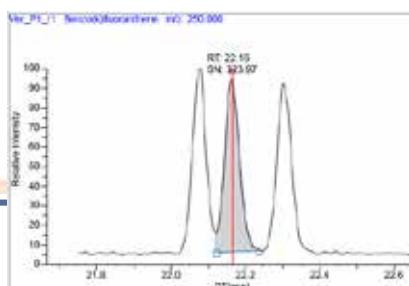
Chrysene



Benzo(b)fluoranthene



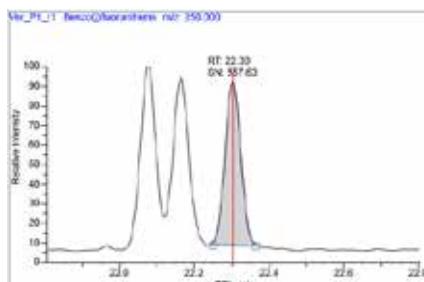
Benzo(k)fluoranthene



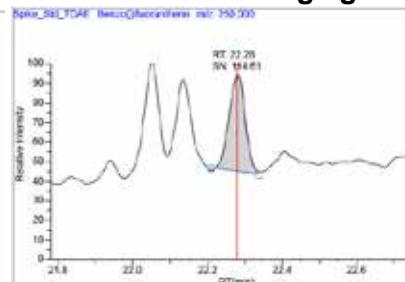
Peak Confirmation

Benzo(j)fluoranthene

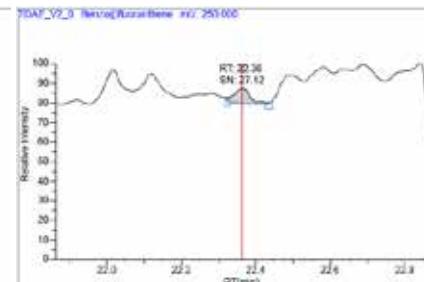
QC Check



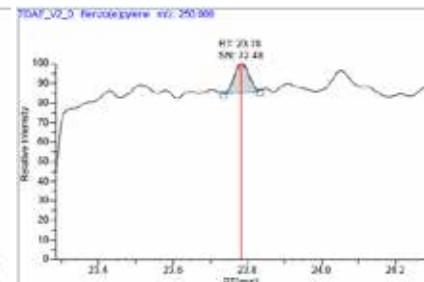
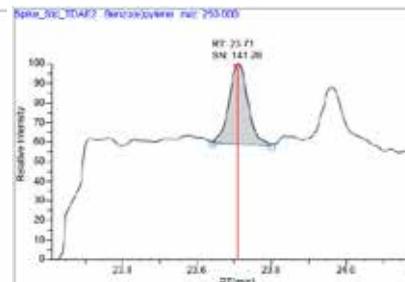
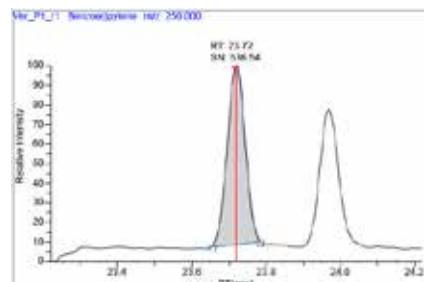
Sample spiked 3 ul of 0.5 mg/kg



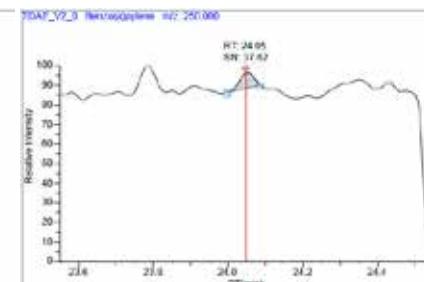
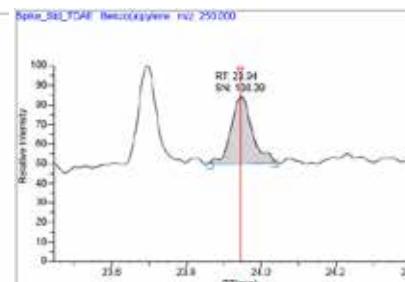
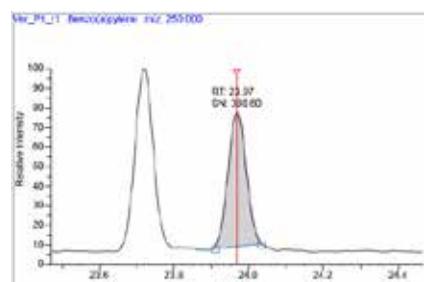
Sample-TDAE



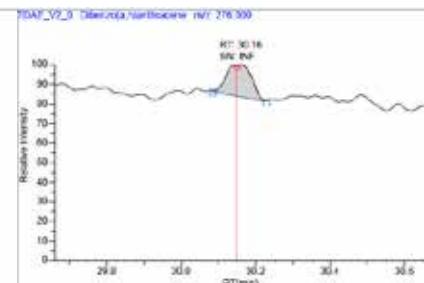
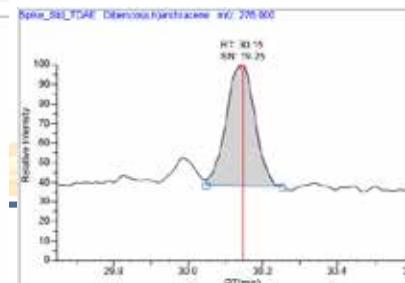
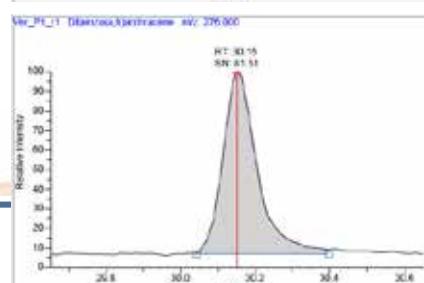
Benzo(e)pyrene



Benzo(a)pyrene



Dibenzo(a,h)anthracene



Result.. Recovery

- Two batches of analysis (2 replicates for each batch) from same sample (RPO)
- Recovery of PAHs : Deuterated IS vs. Injection Standard (Decafluorodiphenyl)
- BIU acceptable recovery is between 50% and 150%

Internal standard	Standard amount (mg)	Sample		Calculated amount (mg)	%Recovery	Acceptable Criteria of %Recovery	Verified
		RPO_V1_Re01	RPO_V1_Re02				
Benzo(a)anthracene-d12	4008	4663.572	4719.434	4691.503	117.05	(50-150)	Pass
Benzo(b)fluoranthene-d12	4216	5684.548	5493.625	5589.087	132.57	(50-150)	Pass
Benzo(a)pyrene-d12	4060	5389.764	5301.968	5345.866	131.67	(50-150)	Pass

		RPO_V2_Re01		RPO_V2_Re02	3532.543	88.14	(50-150)	Pass
		RPO_V2_Re01	RPO_V2_Re02					
Benzo(a)anthracene-d12	4008	3532.543	3532.543	3532.543	3532.543	88.14	(50-150)	Pass
Benzo(b)fluoranthene-d12	4216	3249.254	3249.254	3249.254	3249.254	77.07	(50-150)	Pass
Benzo(a)pyrene-d12	4060	3319.878	3319.878	3319.878	3319.878	81.77	(50-150)	Pass

Comments

- Complicated & time consuming sample preparation – requires skills and prone to error
- Improvement in separation (triphenylene vs. chrysene) can be done upon availability of standard (triphenylene).
- Comparison study of purification between the two steps i.e. Silica Gel vs. Silica Gel & Sephadex are not so much different.
- New development on sample prep in order to reduce work loads and improve analysis result.

Q&A

